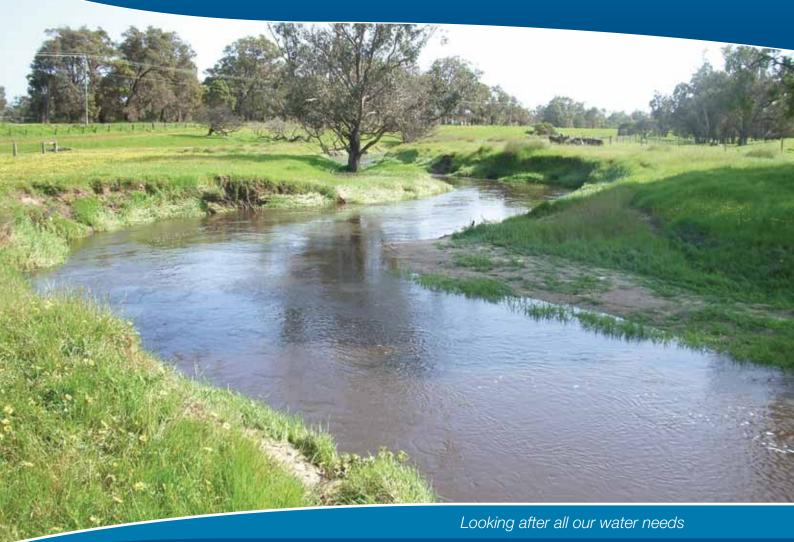


Assessing the influence of acid sulfate soils on water quality in south-western Australian catchments and estuaries

A technical report for the project: Tackling acid sulfate soils on the Western Australian coast





Report no. WST 19 August 2011

Assessing the influence of acid sulfate soils on water quality in south-western Australian catchments and estuaries

A technical report for the project: *Tackling acid sulfate* soils on the Western Australian coast

Looking after all our water needs

By K Kilminster, S Norton and F Miller

Department of Water Water Science Technical Series Report no. 19

August 2011

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Summary

Acid sulfate soil (ASS) is the common name given to soil and sediment that contain iron sulfides (e.g. pyrite). Sulfides are formed by bacterial sulfate-reduction, a process requiring anaerobic conditions, sulfate and degradable organic matter. Oxidised or disturbed ASS release acidity and mobilise metals. Metals and acidity released into the aquatic environment may negatively affect the ecosystem by causing adverse impacts on organisms. This study analysed water collected from catchments and estuaries in south-western Australia for abiotic signals of acidic drainage. This was part of the project *Tackling acid sulfate soils on the Western Australian coast*, funded by the Australian and Western Australian governments through the National Action Plan for Salinity and Water Quality and Natural Heritage Trust initiatives.

As part of this project we developed a new indicator (the sulfur isotope indicator) to identify water affected by acidic drainage from disturbed ASS. With this new indicator we sampled estuaries, rivers, streams and drains in south-western Australia. Drainage from ASS was found to be impacting both fresh water and estuarine water in the region. The impact observed was more subtle when compared with the large quantities of acid produced behind floodgates reported for many ASS-affected sites in Australia's eastern states. Forty-one sites in the current study were identified as hotspot sites, with either unusually high aluminium and/or iron concentrations (within the range expected from reported ASS-affected waterways) or with signals of acidic drainage identified by the sulfur isotope indicator.

The sulfur isotope indicator categorises samples into iso-groups, using measurements of sulfate and chloride concentrations and sulfur stable isotope ratio. This indicator appears to provide a useful screening tool for assessing the influence of acidic drainage. Water samples identified as affected by ASS had deteriorated water quality; median concentrations of many metals (AI, Co, Cr, Cu, Fe, Mn, Ni, V, Zn) and phosphorus were higher in ASS-affected samples (iso-group 3) than in other samples not influenced by ASS. The methodology is more sensitive for identifying acidic drainage compared with other indicators previously used in ASS management. Iso-group categorisation also showed that sulfate-reduction was the dominant process for two-thirds of samples, most of which were associated with extra sources of sulfur. It is likely the supply of extra sulfate to these waterways results in enhanced monosulfidic black ooze production, however further work is required to confirm this.

Reported blackwater events (with resuspended metastable monosulfides) on Australia's east coast have been catastrophic – with hundreds to thousands of fish and prawns reported killed as a result of just one event. Further investigation is required to assess the risk of blackwater events in Western Australia.

Water quality guidelines were frequently exceeded, particularly for nutrients and some metals (AI, Fe and less often Co). It is probable that various sources contributed to the metals observed in waterways and estuaries in south-western Australia, including urban, agricultural and industrial influences, as well as disturbed ASS sources. In 70% of freshwater samples aluminium concentrations exceeded the ANZECC & ARMCANZ (2000) guideline for 95%

freshwater species protection. It is not known if aluminium is naturally high in the region, or if this presents a serious environmental problem.

Signals of chronic acidic drainage were observed in this project, most likely resulting in chronic long-term effects on aquatic ecosystems. These effects may ultimately modify the community structure of species within the aquatic ecosystem, with subsequent effects on the trophic food web.

The priorities for further research were identified as:

- 1 further investigation of hotspot sites identified in this study, including development of management and/or remediation plans for these sites
- 2 additional work to refine the sulfur isotope indicator for assessing ASS influence in waterways
- 3 further work at sites identified as probably having enhanced sulfate-reduction to assess the risk associated with blackwater events at these locations
- 4 ecotoxicology specifically targeted at Western Australian native species to understand whether aluminium concentrations in south-western Australian waterways are presenting a widespread ecological problem.

1 Introduction

1.1 Background to project

The project *Tackling acid sulfate soils on the Western Australian coast* was funded by the Australian and Western Australian governments through the Natural Heritage Trust and the National Action Plan for Salinity and Water Quality. Project funding was administered through the State Natural Resource Management Office with the Department of Water as the lead agency for the project.

This study of water quality within south-western Australian estuaries and catchments was carried out as a surveillance of abiotic indicators to assess the influence of acid sulfate soils on the region's coastal waterways.

1.2 Acid sulfate soil

Acid sulfate soil (ASS) is the common name given to soil and sediment that contains iron sulfides (e.g. pyrite). These are a natural feature of many lowland environments and if left undisturbed under anoxic conditions, they are stable sulfur stores. Sulfides are formed by bacterial sulfate-reduction, a process requiring anaerobic conditions, sulfate and degradable organic matter (Cook et al. 2000). Modern-day ASS formation occurred in the Holocene period after the last major sea-level rise, more than 10 000 years ago (Dent 1986).

When pyrite is disturbed the anoxic sediments are oxidised, producing acidity. This anthropogenically generated acidity may be due to disturbance of sulfide-containing soils during dredging, building or dewatering activities – which may include extraction of groundwater for potable water supplies or forestry practices that result in a lowering of the watertable. The oxidation of iron sulfide produces acid through a number of chemical and biochemical pathways (Langmuir 1997). The overall oxidation reaction of pyrite is described below. Iron hydroxide ($Fe(OH)_3$) is formed when the reaction goes to completion, and in solution iron hydroxide will precipitate as an inert reddish-brown iron-floc.

Equation 1 $\operatorname{FeS}_2 + {}^{15}\!/_4 O_2 + {}^{7}\!/_2 H_2 O \rightarrow \operatorname{Fe}(OH)_3 + 4H^+ + 2SO_4^{2-}$

Often the oxidation of pyrite does not reach completion under typical environmental conditions. In the Australian coastal environment, incomplete oxidation products such as goethite, ferrihydrite, jarosite and schwertmannite have been identified (Sullivan & Bush 2004). Alunite, an aluminium analogue of jarosite, has also been identified in Western Australian soils (Miller et al. 2010).

1.3 Environmental effects of ASS

Oxidised ASS may have negative impacts on land as well as water resources.

• Vegetation scalds may indicate acidity in the topsoil, inhibiting growth of plants and crops.

- ASS may severely damage infrastructure with acid dissolution of concrete and accelerated corrosion of iron, steel and certain aluminium alloys.
- Water quality may deteriorate as a result of oxidised ASS runoff, with possible effects on aquatic life.
- Acidic discharge has been implicated in fish kills (Brown et al. 1983; Hart et al. 1987). Damage to fish gills and skin from acid exposure increases the susceptibility of fish to fungal infections such as 'red spot' (Sammut et al. 1996; Kiryu et al. 2003).

1.4 Metal mobilisation and toxicity

Soils containing sulfides can also contain significant quantities of metals. Metals commonly associated with ASS are aluminium (AI), cadmium (Cd), cobalt (Co), copper (Cu), manganese (Mn), nickel (Ni) and zinc (Zn) (Nordmyr 2008) and iron (Fe), of course, is also prominent since pyrite contains Fe (Cook et al. 2000; Green et al. 2006). Peat soils in Western Australia have also been shown to contain arsenic (As) (Appleyard et al. 2006; Miller et al. 2010). Most metals are more water-soluble in acidic conditions, resulting in leaching of metals as acidic water passes through the soil. Oxidation of metal sulfides may liberate metals associated with sulfides (most commonly FeS₂, although low concentrations of other metal sulfides are typically present). Metals leached from sulfidic soils can greatly contribute to metal concentrations in waterbodies (Sundström et al. 2002) and subsequently may negatively affect the ecosystem by causing adverse impacts on organisms; for example, death, changes in abundance, morphology or appearance, bioaccumulation of metals and/or invasion of tolerant species (Phillips & Rainbow 1994).

1.5 Assessing the influence of ASS on water quality

This project sampled natural waters (estuarine and fresh) for abiotic signals of acidic drainage from ASS. Approximately 60 physical and chemical variables were analysed in each water sample. Little information exists for many of these variables in natural waters; thus the project also aimed to provide baseline data by investigating relatively undisturbed as well as disturbed sites. Approximately 150 sites across five freshwater catchment areas (Swan Canning, Peel Harvey, Leschenault, Scott Coastal Plain and the south coast) and approximately 90 sites within 13 estuaries in south-western Australia were investigated. Estuarine sites were typically strongly brackish (>20 psu) to hypersaline (>40 psu) in the summer months, while freshwater flows during winter resulted in some slightly brackish water samples. Catchment samples were fresh to slightly brackish, with most samples <0.5 psu.

2 Potential indicators of acidic drainage

Potential indicators of acidic drainage explored in this project included the chloride:sulfate ratio, sulfur stable isotope ratio, total acidity and metal concentrations (Fe, Al particularly). These potential indicators often need to be considered collectively as many interactions are possible and may confound interpretation.

Chloride:sulfate ratios

Seawater is comprised of many ions: the major ions include chloride (CI⁻), sodium (Na⁺), magnesium (Mg⁺), sulfate (SO₄²⁻), calcium (Ca²⁺) and potassium (K⁺). In undiluted seawater, the concentration of chloride is approximately 19 400 mg L⁻¹ and sulfate is approximately 2700 mg L⁻¹, producing a ratio between the two ions of 7.2:1 (for CI⁻:SO₄²⁻). Chloride is considered a conservative ion, so the ratio will be conserved with no other source of sulfate present in a natural water sample (other than seawater). With other inputs of sulfur (e.g. acidic drainage, fertiliser or greywater) this ratio will be reduced. Since sulfate is released by the oxidation of pyrite (Equation 1), low chloride:sulfate ratios (<4 or <3) have previously been used to indicate drainage from disturbed ASS (Sammut et al. 1996; Hinwood et al. 2006; NSW Department of Primary Industries 2008). Note: geochemists will often report the reverse ratio (e.g. SO₄²⁻:Cl⁻).

Total acidity

Measuring acidity by pH alone does not account for all sources of acidity in natural water. Measures of pH quantify the concentration of hydrogen ions (H⁺) in solution on a logarithmic scale, and only measure accurately the acidity from fully dissociated acids (i.e. strong acids). Alternatively, the measure of total acidity is a titration-based measurement that provides information on the total amount of base needed to completely neutralise the solution. Total acidity reflects all acidity-generating components in the water and includes H⁺, acidity from hydrolysing metals and acidity from weak acids that do not fully dissociate (e.g. formic, acetic acid and those carboxylic groups associated with humic acid).

Metal concentrations (iron and aluminium)

High concentrations of metals are usually associated with acidic drainage, as the solubility of metals increases with decreasing pH. Aluminium and iron are commonly associated with ASS (Cook et al. 2000): soil-leaching studies have shown that these elements are leached in orders-of-magnitude higher concentrations than other metals present within the soil (Miller et al. 2010). The use of metal concentrations in water as an indicator of acidic drainage is complicated by other influences, such as urban and industrial sources of metals and changes in metal solubility related to mixing with buffered estuarine waters. Metals such as Fe and AI can also contribute acidity depending on the form (chemical species) which is present in the waterway; for example, hydrolysis of iron (II) to iron hydroxide produces H⁺ (i.e. acid) and this acidity is sometimes referred to as 'stored acidity' (Sullivan & Bush 2004).

Sulfur stable isotope ratio

Measuring δ^{34} S value of sulfate in water provides information on the source of the sulfate. Sulfur occurs naturally in four stable isotope forms, of which ³²S and ³⁴S are most abundant (95% and 4.2% respectively). Differences in the relative abundance of ³⁴S are determined relative to a standard and δ^{34} S is expressed on the per mil scale (‰) (Clarke & Fritz 1997). Sulfur-34 abundance varies between sulfur compounds due to physical processes and biological cycling such as fractionation by sulfate-reducing bacteria (Kaplan & Rafter 1957; Clarke & Fritz 1997). Reduction of sulfate by sulfate-reducing bacteria results in isotopically light δ^{34} S (i.e. relatively more ³²S). The δ^{34} S value of seawater is highly positive (~20 to 21 ‰) compared with that of sulfide deposits such as pyrite, which are highly negative (-18 to -52 ‰) (Coomer & Robinson 1976; Jowett et al. 1991).

The potential to use sulfur isotope analysis as an indicator of acidic drainage and sulfatereduction processes within natural waters is investigated in this project. This is a novel and innovative approach to assessing acidic drainage from disturbed ASS.

3 Methods

3.1 Sampling locations

More than 300 surface water samples were collected from estuaries and catchment waterways (main rivers, tributaries and drains) in south-western Australia. The sampling locations included freshwater, brackish and saline systems, but only rarely lakes or wetlands. The regions sampled were:

- Swan-Canning estuary (16 sites) and Swan-Canning catchment (30 sites) (Figure 1)
- Peel-Harvey estuary (30 sites, including shallow sites at the margin of the estuary where fauna was also sampled) and Peel-Harvey catchment (31 sites) (Figure 2)
- Leschenault Estuary (10 sites) and Leschenault catchment (28 sites) (Figure 3),
- Vasse-Wonnerup estuary (seven sites) (Figure 4)
- Hardy Inlet (12 sites) and Scott Coastal Plain catchment (28 sites) (Figure 5)
- Broke, Walpole Nornalup and Irwin inlets (10 sites) (Figure 6)
- Wilson Inlet, Princess Royal Harbour and Oyster Harbour (nine sites) and Albany catchment (30 sites) (Figure 7).

Typically, estuarine sites were sampled twice. These are nominally called 'summer' and 'winter', and notated throughout the text with –s or –w after the site name (e.g. ELL-s). For the summer sampling period, actual dates range from January to March; for the winter sampling period, May to September (with most samples taken in June to August). Freshwater catchment sites were sampled only once (spring 2007). Details of site locations and sampling dates can be found in Appendix A.

3.2 Sampling method and analysis

Samples were collected using a grab pole sampler from a depth of 0–0.5 m. A subsample was filtered on-site through a 0.45 µm filter. At each site, in situ measurements of dissolved oxygen (DO), pH, conductivity, salinity and oxidation-reduction potential (freshwater samples only and converted to Eh for reporting) were obtained with a multi-parameter probe (either Hydrolab Quanta or Hydrolab minisonde 5). The collected water samples from each site were analysed to determine nutrients – total nitrogen (TN), nitrate and nitrite (NOx), ammonium (NH₄⁺), dissolved organic nitrogen (DON), total phosphorus (TP), filterable reactive phosphorus (FRP), total acidity, total alkalinity, dissolved organic carbon (DOC), sulfur stable isotope (δ^{34} S), colour (freshwater samples only) and a suite of total and filtered (or soluble) metals. Metals analysed generally included aluminium (AI), arsenic (As), calcium (Ca) cadmium (Cd), cobalt (Co), chromium (Cr), copper (Cu), iron (Fe), mercury (Hg), magnesium (Mg), manganese (Mn), molybdenum (Mo), sodium (Na), nickel (Ni), lead (Pb), selenium (Se), vanadium (V), and zinc (Zn). Total metal suite (i.e. metals determined from an unfiltered sample) was not analysed for the summer estuary snapshot. Analysis detail is described in full in Appendix B.

Procedural blanks (de-ionised water samples prepared in the field) and field replicate samples were collected throughout the sampling events. Quality control evaluation was also carried out on laboratory-reported blanks, duplicates and spike recoveries. Data from quality assurance and quality control are reported in Appendix C.

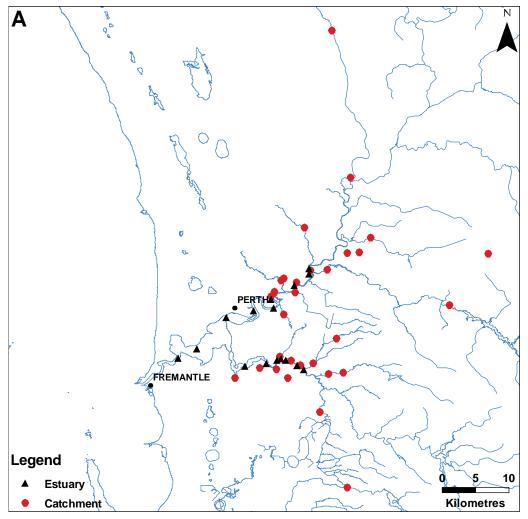


Figure 1 Surface water sampling sites for the Swan-Canning region. Red dots indicate catchment sampling sites and black triangles indicate estuary sampling sites.

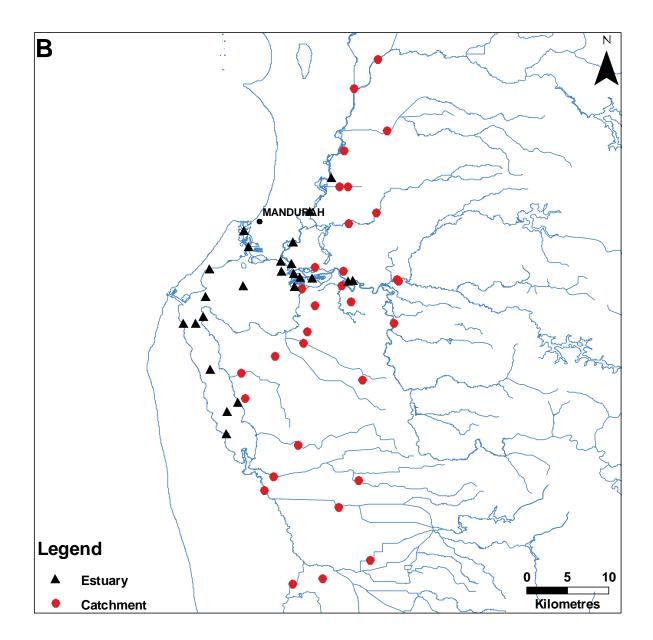


Figure 2 Surface water sampling sites for the Peel-Harvey region. Red dots indicate catchment sampling sites and black triangles indicate estuary sampling sites.

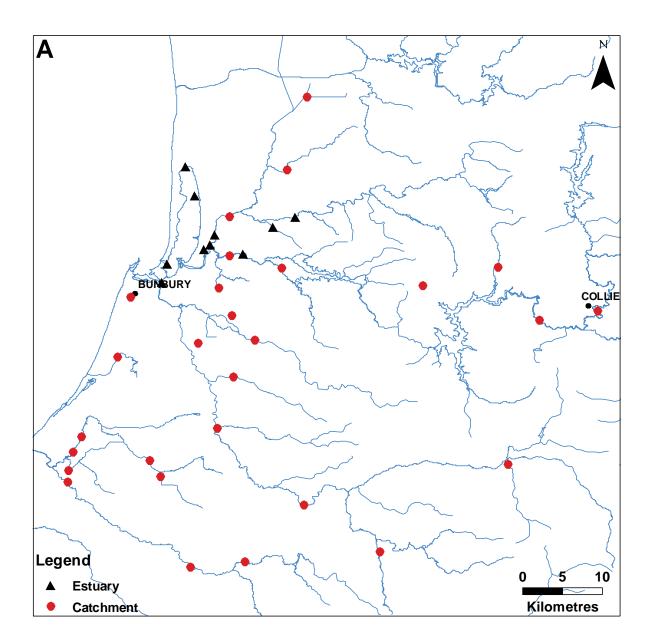


Figure 3 Surface water sampling sites for the Leschenault region. Red dots indicate catchment sampling sites and black triangles indicate estuary sampling sites.

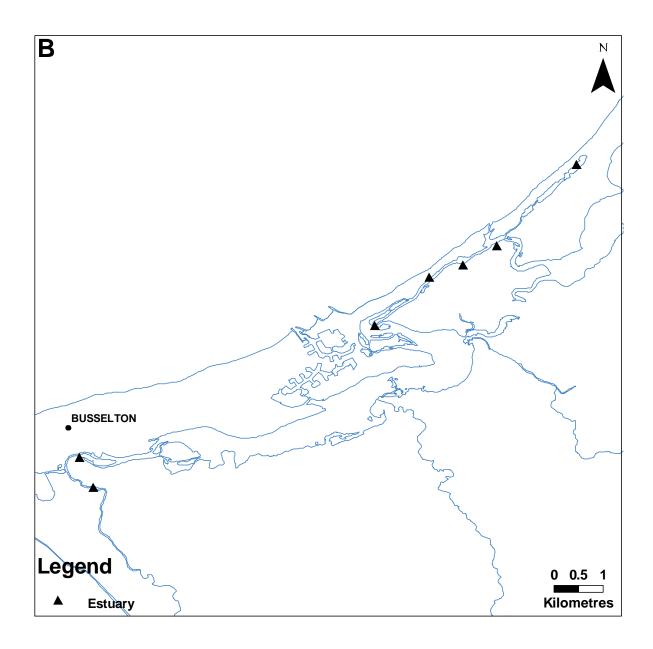


Figure 4 Surface water sampling sites for the Vasse-Wonnerup estuary (black triangles indicate sampling sites).

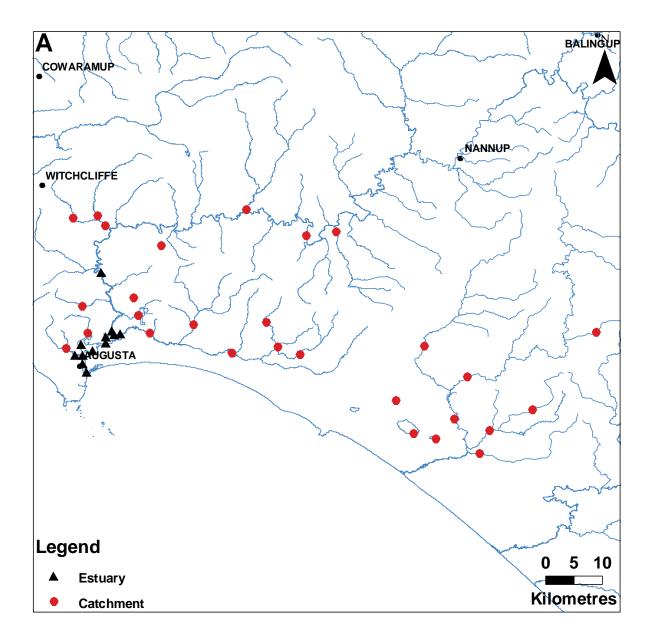


Figure 5 Surface water sampling sites for the Hardy Inlet and Scott Coastal Plain. Red dots indicate catchment sampling sites and black triangles indicate estuary sampling sites.

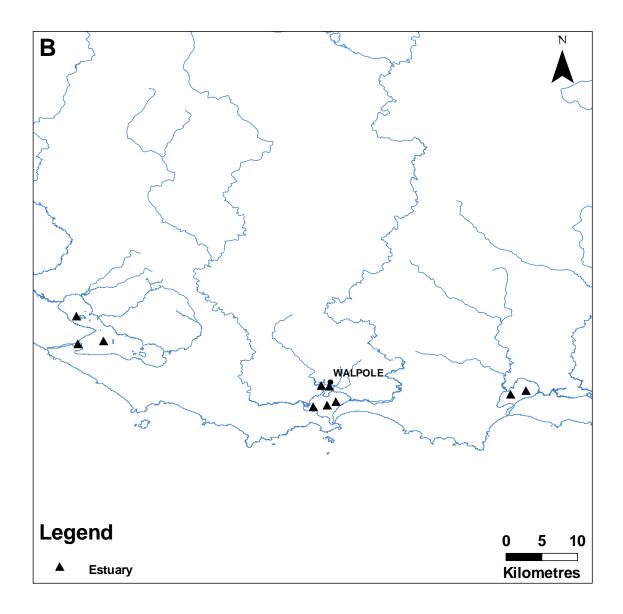


Figure 6 Surface water sampling sites for Broke, Walpole Nornalup and Irwin inlets. Red dots indicate catchment sampling sites and black triangles indicate estuary sampling sites.

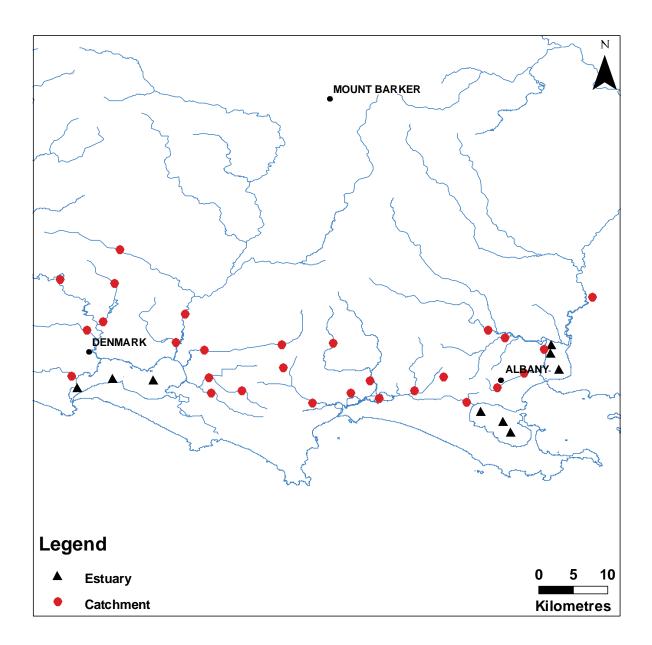


Figure 7 Surface water sampling sites for Wilson Inlet, Princess Royal Harbour, Oyster Harbour, and south coast catchment. Red dots indicate catchment sampling sites and black triangles indicate estuary sampling sites.

3.3 Data storage

Data for this project are housed within the Department of Water's Water Information Network (WIN) database. Data reported here are associated with the following project codes: WA-R-WAACIDEST, SG-C-ACIDCATSC, KP-C-WAACIDCATP, SW-C-ACIDCATL, SW-C-ACIDCATSCP, SC-A-ACIDCATA.

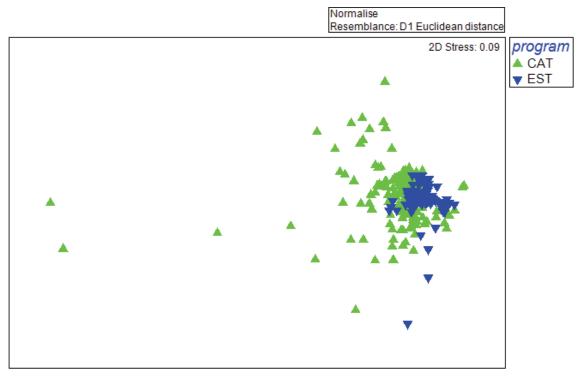
3.4 Data analysis

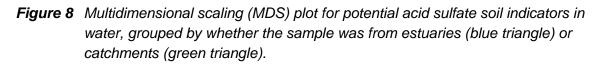
Data were transformed and normalised before statistical analysis. The following programs were used for data manipulation, graphical representation and statistical analysis: Excel 2003, PRIMER v6 with PERMANOVA+, Sigmaplot version 10 (Systat Software Inc) and Statistica version 8 (StatSoft Inc). Where necessary, resemblance matrixes were calculated using Euclidian-distance on transformed and normalised data. Statistical routines used are stated in the text; however, further detail on PRIMER and PERMANOVA+ routines can be found in Clarke and Warwick (2001) and Anderson et al. (2008).

4 Results: regional variation

4.1 Potential ASS indicators by region

The variables outlined in Section 2 as potential indicators of ASS were examined for their variation across different regions. Firstly estuarine and catchment data, pooled from all geographical regions, were compared. A multidimensional scaling (MDS) plot for the variable subset of potential ASS indicators – total acidity, aluminium (soluble), iron (soluble), pH and chloride:sulfate ratio – showed clear differences between estuaries and catchments (Figure 8).





Salinity is likely to have a major effect on many of these potential ASS indicators; for example, saline water in estuaries buffers pH, and consequently metals that might be soluble in acidic conditions will precipitate at more neutral pH. To account for this, electrical conductivity was used as a co-variate in a permutational multivariate analysis of variance (PERMANOVA) to analyse differences in the variable subset of potential ASS indicators between catchments and estuaries. Catchment and estuarine data were found to be significantly different (pseudo-F=0.5, p>0.0001). The different distributions of these potential indicators are shown as boxplots in Figure 9. These boxplots are a non-parametric way to display the whole sample distribution, without assumptions about data distribution. Much higher concentrations of Fe and AI were observed in catchment samples than in estuarine samples (Figure 9). In addition, catchment samples showed much higher variability of AI, Fe and CI⁻:SO₄²⁻ than estuarine samples.

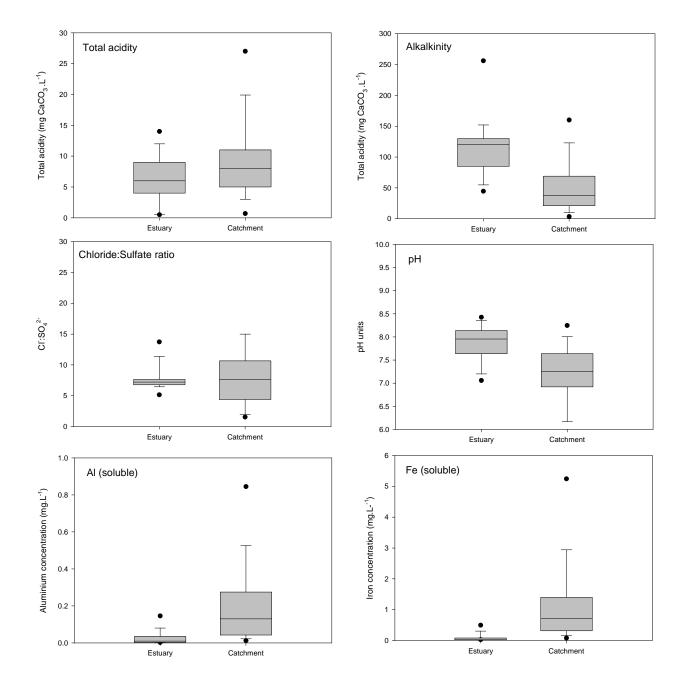


Figure 9 Boxplots of estuarine and catchment water quality for potential acid sulfate soil indicators – total acidity, alkalinity, chloride:sulfate ratio, pH, AI (soluble) and Fe (soluble), (line = median, edges of box 25th and 75th, whiskers 10th and 90th, and dots 5th and 90th percentile).

The data were then explored for differences between estuaries (e.g. Swan-Canning vs Hardy) and between catchments (e.g. Peel Harvey vs Leschenault). The median and range of total acidity, total alkalinity, pH, Eh, concentrations of sulfate and chloride, $C\Gamma:SO_4^{2^-}$ and the $\delta^{34}S$ are shown in Table 1 for estuarine data and Table 2 for catchment (freshwater) data. Univariate analysis of variance (ANOVA) showed that estuary regions were significantly different (F>7, p<0.00001) for all variables in Table 1 except $\delta^{34}S$. Univariate ANOVA showed that catchment regions were also significantly different (F>3.5, p<0.01) for all variables in Table 2 except $\delta^{34}S$. Major differences between estuaries with these parameters are most likely to reflect differences in location (e.g. the Swan-Canning estuary has more up-river sites than Oyster Harbour) and the relative influence of salinity (which may vary with season). The regional differences observed highlight the importance of using electrical conductivity (or salinity) as a co-variate in multivariate analysis of this dataset.

A large range in values was observed for most variables, and in most geographical regions (Table 1 and Table 2). Lower extremes of pH were observed in catchments (lowest-value pH 2.8) than in estuaries (lowest-value pH 6.5), and higher median values of alkalinity were measured in estuaries compared with catchments, demonstrating the higher buffering capacity of estuarine water. Furthermore, higher extremes of total acidity were also seen in catchments (maximum 290 mg CaCO₃ L⁻¹), compared with estuaries (maximum 18 mg CaCO₃ L⁻¹).

Sites were classified as either 'acidic' or 'reducing' based solely on the $C\Gamma:SO_4^{2-}$ ratio determined for each site. This indicator is a ratio of two chemical analyses, so the combined error associated with each measurement in the ratio was considered when categorising sites as either acidic or reducing (Table 3). Acidic sites were those with the $C\Gamma:SO_4^{2-}$ ratio <4 (indicating excess sulfate), and reducing sites were those with a ratio greater than expected for seawater (indicating missing sulfur). Estuarine sites were generally similar to seawater (~7.2 ± error). Only one estuarine site (within the Peel-Harvey estuary) was classified as acidic. The Hardy Inlet had many reducing sites (83%) followed by the Leschenault, Peel-Harvey and Vasse-Wonnerup estuaries (~20%). Within catchment sites, the Swan Canning had the greatest number of acidic sites (70%). Acidic sites were also common in the south coast and Scott Coastal Plain catchments (13% and 11% respectively). Reducing sites in the catchment areas were most often found in the Scott Coastal Plain (25%) and the Peel Harvey (16%).

| Variable (unit) | | Swan Canning | Peel Harvey | Leschenault | Vasse Wonnerup | Hardy | South coast estuaries |
|---|---|--------------------------|---------------------------|---------------------------|-------------------------|------------------------|----------------------------|
| Total acidity (mg CaCO ₃ L ⁻¹) | median range % <lor< td=""><td>10 4–18 0%</td><td>4 <1–14 33%</td><td>6 <1–18 0%</td><td>8 <1–16 21%</td><td>4.5 3–5 0%</td><td>6 2–11 0%</td></lor<> | 10 4–18 0% | 4 <1–14 33% | 6 <1–18 0% | 8 <1–16 21% | 4.5 3–5 0% | 6 2–11 0% |
| Alkalinity (mg CaCO ₃ L ⁻¹) | median range % <lor< td=""><td>130 66–20 0%</td><td>120 59–170 0%</td><td>58 22–330 0%</td><td>140 42–180 0%</td><td>53 45–63 0%</td><td>120 44–140 0%</td></lor<> | 130 66–20 0% | 120 59–170 0% | 58 22–330 0% | 140 42–180 0% | 53 45–63 0% | 120 44–140 0% |
| pH (pH units) | median range % <lor< td=""><td>7.7 7.0–8.1 0%</td><td>8.0 6.5–9.3 0%</td><td>7.7 6.7–8.2 0%</td><td>8.0 7.3–8.7 0%</td><td>7.6 7.2–8.6 0%</td><td>8.1 7.7–8.5 0%</td></lor<> | 7.7 7.0–8.1 0% | 8.0 6.5–9.3 0% | 7.7 6.7–8.2 0% | 8.0 7.3–8.7 0% | 7.6 7.2–8.6 0% | 8.1 7.7–8.5 0% |
| Chloride (mg L ⁻¹) | median range % <lor< td=""><td>14000 170–19000 0%</td><td>18000 1700–43000 0%</td><td>1030 * 100–30000 0%</td><td>11600 160–2400 0%</td><td>1650 140–2200 0%</td><td>19000 13000–22000 0%</td></lor<> | 14000 170–19000 0% | 18000 1700–43000 0% | 1030 * 100–30000 0% | 11600 160–2400 0% | 1650 140–2200 0% | 19000 13000–22000 0% |
| Sulfate (mg L ⁻¹) | median range % <lor< td=""><td>2000 33–2800, 0%</td><td>2600 110–5,900 0%</td><td>112 * 15–4600 0%</td><td>1750 10–3200 0%</td><td>120 29–230 0%</td><td>2650 1700–3100 0%</td></lor<> | 2000 33–2800, 0% | 2600 110–5,900 0% | 112 * 15–4600 0% | 1750 10–3200 0% | 120 29–230 0% | 2650 1700–3100 0% |
| Cl ⁻ :SO ₄ ²⁻ (ratio) | median range | 7.1 4.9–9.1 | 7.2 0.7–15.5 | 7.0 4.1–16.5 | 6.8 5.0–47 | 12.0 4.8–14.5 | 7.4 6.5–8.2 |
| δ^{34} S (per mil, ‰) | median range | 20.3 9.8–22.2 | 20.7 -7.0–23.6 | 20.4 3.4–27.9 | 20.8 10.1–22.8 | 22.5 20.5–23.4 | 20.5 18.6–22.8 |

Table 1 Summary data (median and range) for water quality in estuaries across south-western Australia for some potential indicators of acidic drainage. <LOR = less than limit of reporting; * indicates bimodal distribution.</th>

| Variable (unit) | | Swan Canning | Peel Harvey | Leschenault | Scott Coastal Plain | South coast |
|---|---|-----------------------|----------------------|-----------------------|------------------------|----------------------|
| Total acidity (mg CaCO ₃ L ⁻¹) | median | 4.5 | 9 | 6 | 10 | 11 |
| | range | <1–31 | <1–57 | 1–27 | 6–62 | 4–290 |
| | % <lor< td=""><td>13%</td><td>6%</td><td>0%</td><td>0%</td><td>0%</td></lor<> | 13% | 6% | 0% | 0% | 0% |
| Alkalinity (mg CaCO ₃ L ⁻¹) | median | 65 | 56 | 41 | 18 | 23.0 |
| | range | 17–250 | 3–180 | 12-240 | <1–200 | <1–260 |
| | % <lor< td=""><td>0%</td><td>0%</td><td>0%</td><td>4%</td><td>10%</td></lor<> | 0% | 0% | 0% | 4% | 10% |
| pH (pH units) | median range % <lor< td=""><td>7.4 6.2 –9.3 0%</td><td>7.1 4.8–9.5 0%</td><td>7.16 6.1–7.7 0%</td><td>6.8 4.1–7.6 0%</td><td>7.2 2.8–8.1 0%</td></lor<> | 7.4 6.2 –9.3 0% | 7.1 4.8–9.5 0% | 7.16 6.1–7.7 0% | 6.8 4.1–7.6 0% | 7.2 2.8–8.1 0% |
| Redox (Eh) | median | 311 | 343 | 345 | 347 | 366 |
| (mV) | range | 243–386 | 179–419 | 262–389 | 213– 583 | 249–700 |
| Chloride (mg L ⁻¹) | median range % <lor< td=""><td>140 80–450 0%</td><td>190 60–9000 0%</td><td>105 50–480 0%</td><td>130 50–1500 0%</td><td>220 60–2300 0%</td></lor<> | 140 80–450 0% | 190 60–9000 0% | 105 50–480 0% | 130 50–1500 0% | 220 60–2300 0% |
| Sulfate (mg L ⁻¹) | median range % <lor< td=""><td>62 15–340 0%</td><td>25 <5–1100 10%</td><td>15 6–43 0%</td><td>12 <5–290 14%</td><td>30 <5–630 3%</td></lor<> | 62 15–340 0% | 25 <5–1100 10% | 15 6–43 0% | 12 <5–290 14% | 30 <5–630 3% |
| Cl ⁻ :SO4 ²⁻ | median | 3.4 | 8.2 | 6.9 | 10.3 | 8.0 |
| (ratio) | range | 0.7–15 | 3.2–88 | 3.3–23 | 0.9–96 | 0.6–24 |
| δ^{34} S | median | 17.3 | 22.0 | 20.8 | 22.5 | 19.7 |
| (per mil, ‰) | range | 11.7–25.1 | 7.9-31.4 | 16.2–26.2 | -6.6–27.8 | 3.2–27.7 |

Table 2 Summary data (median and range) for water quality in catchments across south-western Australia for some potential indicators of acidic drainage. <LOR = less than limit of reporting.</td>

| Region | n | Acidic sites (extra sulfate) | Reducing sites (missing sulfate) |
|------------------------|----|---------------------------------|-------------------------------------|
| Estuaries | | | |
| Swan-Canning estuary | 32 | 0 | 3 |
| Peel-Harvey estuary | 51 | 1 | 10 |
| Leschenault Estuary | 20 | 0 | 4 |
| Vasse-Wonnerup estuary | 14 | 0 | 3 |
| Hardy Inlet | 12 | 0 | 10 |
| South coast estuaries | 28 | 0 | 1 |
| Catchments | | | |
| Swan Canning | 30 | 21 | 1 |
| Peel Harvey | 37 | 1 | 6 |
| Leschenault | 28 | 2 | 3 |
| Scott Coastal Plain | 28 | 3 | 7 |
| South coast | 31 | 4 | 2 |

Table 3 Number of sites (or samples) classified as either 'acidic' ($C\Gamma:SO_4^{2^-} < 4$) or 'reducing' ($C\Gamma:SO_4^{2^-} > 7.2$) using only the $C\Gamma:SO_4^{2^-}$ ratio for both estuarine and catchment regions.

4.2 Electrical conductivity by region

Electrical conductivity measured in the field was converted to temperature compensated conductivity (i.e. EC at 25°C) for reporting. Figure 10 shows the distribution of conductivity measured in samples from estuarine areas and samples from the catchment (i.e. rivers, streams, drains). There is some overlap in the conductivity measured between these two groups. Catchment samples with measured electrical conductivity suggest either saltwater intrusion into lowland rivers or highly contaminated sites. Conversely, samples within estuarine reaches showing low conductivity reflect the influence of fresh water in the upper estuary (usually a sample collected in winter). Median conductivities and ranges are shown for each sampled region in Table 1 and Table 2.

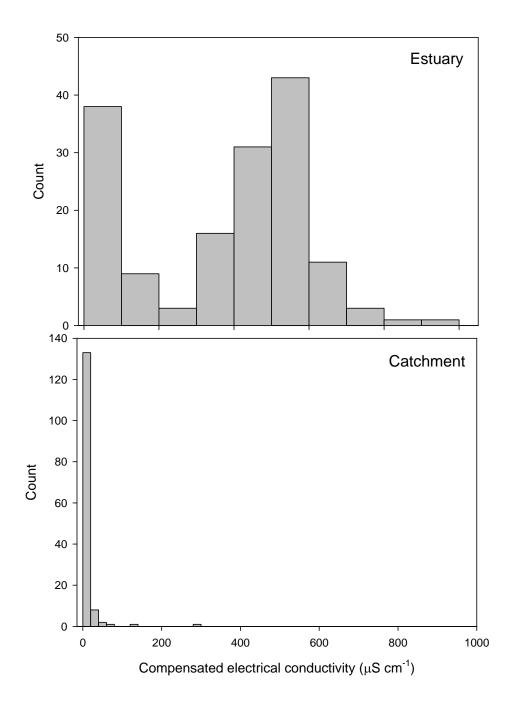


Figure 10 Histograms of compensated conductivity for samples classified as estuarine and those classified as catchment (freshwater). Samples were designated as either estuarine or catchment prior to sampling.

5 Results: seasonal differences

Seasonal differences within the data were explored for estuarine samples only (catchment samples were obtained once as a snapshot). PERMANOVA was used to test for differences among groups of variables with electrical conductivity as a co-variate. Including electrical conductivity as a co-variate removes the effect of salinity from the statistical analysis for season. PERMANOVA for the acidity subset of variables (included pH, total acidity, alkalinity and $CI^{-}SO_4^{-2}$) was significant (pseudo-F=3.7, p<0.01). PERMANOVA for the soluble metals subset of variables (included AI, As, Cd, Cr, Co, Cu, Fe, Hg, Mn, Mo, Ni, Pb, Se, V, Zn) was significant for season (pseudo-F=5.4, p<0.01). Total metals were only analysed in the second sampling period, so seasonal trends could not be determined.

Soluble Fe, Al and Mn were all higher in winter than in the summer water samples across all south-west estuaries (Figure 11). Soluble Cu, Ni and Zn did not show a seasonal trend, however there was variation across estuary region in these elements, with the highest concentrations of Cu and Zn found in the Swan-Canning estuary (Figure 12).

PERMANOVA for the nutrient subset of variables (included TN, NOx, NH_4^+ , DON, TP, FRP) was not significant for season (pseudo-F=0.4, p=0.7). The lack of a significant difference for season is most likely due to high geographical regional variation; that is, differences in nutrient concentrations between sites overwhelmed any overall seasonal variation present.

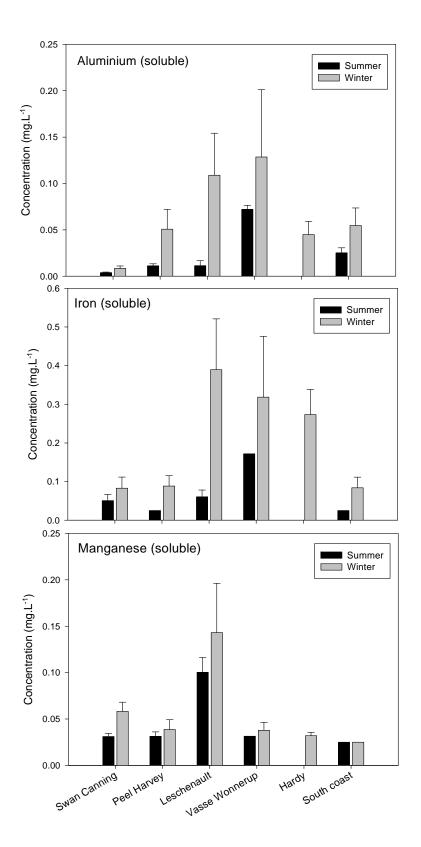


Figure 11 Average soluble aluminium, iron, and manganese concentrations in summer and winter sampling events across south-western Australian estuaries (mean + st. err.).

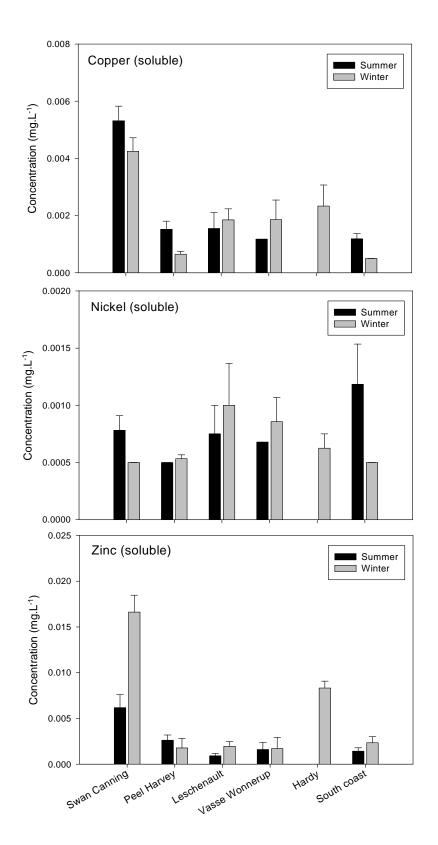


Figure 12 Average soluble copper, nickel, and zinc concentrations in summer and winter sampling events across south-western Australian estuaries (mean + st. err.).

6 Results: data relationships

6.1 Correlations of variables

Estuarine and catchment data were combined to compare the relationships of soluble metals with total metals. There were strong positive relationships between soluble and total concentrations for AI, Co, Mn and Mo (R>0.99). Additionally, major ions Mg and Ca showed very strong correlations between soluble and total measurements (R>0.999). Soluble and total concentrations for As, Cr, Cu, Fe, Ni, Pb, V and Zn were not strongly correlated. Dissolved organic carbon was strongly correlated with colour (R=0.93), dissolved organic nitrogen (R=0.86), total phosphorus (R=0.75) and filterable reactive phosphorus (R=0.83).

6.2 Distribution of variables

Not all variables determined in this study have appropriate guideline values against which their concentrations can be compared. In the absence of such guidelines, the distribution of each variable was considered in terms of the percentile range to provide cut-off values for high and low (extreme) values within the dataset (Figure 13). As a lower limit, the 10th percentile value was chosen, representing the value at which 10% of samples were less than the value. Similarly, the 90th percentile value indicates the value at which 10% of samples were higher. The percentile ranges were calculated separately for estuarine and catchment data (Table 4). As expected, acidity was lower and, correspondingly, alkalinity was higher in the estuaries than the catchments; while dissolved organic carbon was higher in the catchments than the estuaries. Median values for AI in both estuaries and catchments were above recommended guideline values. Interestingly, for the estuarine data the 90th percentile value for soluble Fe was 0.3 mg L⁻¹, and for total Fe it was 1.1 mg L⁻¹ – values very similar to the recent guidelines proposed for Fe in British Columbia, Canada (Phippen et al. 2008) of 0.35 mg L⁻¹ for soluble iron and 1.0 mg L⁻¹ for total iron. Higher concentrations of Fe were observed in the catchment samples with medians of 0.72 and 1.35 mg L⁻¹ for soluble and total Fe respectively.

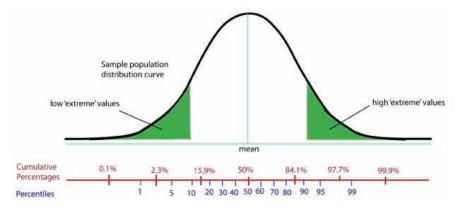


Figure 13 Diagram showing high and low extreme values from the distribution of the dataset for each variable at 10th and 90th percentiles.

| | Estuaries | | | | | Catchments | | | |
|---|--------------------|--------------------|--------|------|--------------------|--------------------|--------|-----|--|
| Variables (units) | 10th percentile | 90th percentile | median | n | 10th percentile | 90th percentile | median | n | |
| Total acidity (mg CaCO ₃ L ⁻¹) | <1.0 | 12 | 6.0 | 167 | 3.0 | 19 | 8.0 | 146 | |
| Alkalinity (mg CaCO ₃ L^{-1}) | 55 | 150 | 120 | 167 | 10 | 120 | 38 | 146 | |
| pH (pH units) | 7.20 | 8.35 | 7.95 | 156 | 6.11 | 7.79 | 7.17 | 146 | |
| Dissolved organic carbon (mg L ⁻¹) | 2.0 | 24 | 7.0 | 167 | 4.0 | 72 | 18 | 146 | |
| Colour (TCU) | n.d. | n.d. | n.d. | n.d. | 20 | 790 | 110 | 137 | |
| Redox (Eh) (mV) | n.d. | n.d. | n.d. | n.d. | 280 | 401 | 341 | 131 | |
| Aluminium (sol) (mg L ⁻¹) | <0.005 | 0.10 | 0.10 | 167 | 0.024 | 0.51 | 0.13 | 145 | |
| Aluminium (tot) (mg L ⁻¹) | 0.022 | 0.31 | 0.084 | 93 | 0.059 | 0.93 | 0.27 | 146 | |
| Arsenic (sol) (mg L ⁻¹) | <0.001 | 0.001 | 0.001 | 164 | <0.001 | 0.002 | <0.001 | 145 | |
| Arsenic (tot) (mg L ⁻¹) | <0.001 | 0.008 | 0.001 | 85 | <0.001 | 0.002 | <0.001 | 146 | |
| Copper (sol) (mg L ⁻¹) | <0.001 | 0.001 | 0.001 | 164 | <0.001 | 0.002 | <0.001 | 145 | |
| Copper (tot) (mg L ⁻¹) | <0.001 | 0.007 | 0.001 | 85 | <0.001 | 0.003 | <0.001 | 146 | |
| Iron (sol) (mg L ⁻¹) | <0.05 | 0.3 | <0.05 | 167 | 0.17 | 2.9 | 0.72 | 145 | |
| Iron (tot) (mg L ⁻¹) | <0.050 | 1.1 | 0.28 | 93 | 0.36 | 4.9 | 1.35 | 146 | |

Table 4Summary data for estuarine and catchment water quality showing 10th and 90th
percentile values and median for the range of variables analysed. Values
preceded by the < symbol did not exceed limit of reporting.</th>

| Manganese (sol) (mg L ⁻¹) | <0.050 | 0.098 | <0.050 | 164 | 0.008 | 0.13 | 0.030 | 145 |
|--|--------|-------|--------|-----|--------|-------|--------|-----|
| Manganese (tot) (mg L ⁻¹) | 0.007 | 0.16 | 0.028 | 85 | 0.011 | 0.14 | 0.038 | 146 |
| Nickel (sol) (mg L ⁻¹) | <0.001 | 0.001 | 0.001 | 152 | <0.001 | 0.002 | <0.001 | 145 |
| Nickel (tot) (mg L ⁻¹) | <0.001 | 0.002 | 0.001 | 73 | <0.001 | 0.003 | <0.001 | 146 |
| Zinc (sol) (mg L⁻¹) | <0.001 | 0.012 | 0.001 | 167 | <0.001 | 0.014 | 0.002 | 145 |
| Zinc (tot) (mg L ⁻¹) | <0.001 | 0.019 | 0.002 | 91 | <0.001 | 0.015 | 0.002 | 146 |

7 Results: guideline comparison

The estuarine and catchment water quality data were compared with appropriate guideline values. Physical and nutrient guidelines for aquatic ecosystem stress are ecosystem-specific (e.g. estuarine, lowland rivers etc.), hence guidelines were chosen according to the ecosystem sampled. There are no specific guidelines available for metals in estuaries so, as recommended by ANZECC & ARMCANZ (2000), the lowest-available guideline value was chosen for comparative purposes from either the freshwater or marine guideline values. The freshwater samples (catchment snapshots) were compared with ANZECC & ARMCANZ (2000) trigger values for metals for 95% freshwater species protection. In all cases, the guideline or trigger concentration used for comparison is clearly stated.

7.1 Estuarine water quality: physical guidelines

The pH and dissolved oxygen measured in south-western Australian estuaries were compared with the ANZECC & ARMCANZ (2000) trigger values for estuarine ecosystem stress (Table 5). Where a pH guideline value was triggered, in most cases the sample was below the lower limit of 7.5. The exception to this was the Vasse Wonnerup, where almost a third of sites were above the upper-limit trigger value of 8.5. The pH guideline value was not exceeded for any of the south coast estuaries.

Dissolved oxygen was generally depleted rather than enhanced in the estuarine water samples. The Swan-Canning and Leschenault estuaries showed the lowest oxygen concentrations with 69% and 81% of samples (respectively) having oxygen concentrations below 90% oxygen saturation. High concentrations of dissolved oxygen (i.e. greater than 110% oxygen saturation) generally indicate eutrophic systems where oxygen has been enhanced by primary production (e.g. algal blooms). Instances where samples exceeded the upper-limit trigger value for dissolved oxygen occurred in samples from the Vasse-Wonnerup (26%) and Peel-Harvey (16%) estuaries, as well as Hardy Inlet (8%).

Table 5Comparison of estuarine water quality data for pH and dissolved oxygen data
against guideline values by area. Guideline values are ANZECC & ARMCANZ
(2000) trigger values for estuarine ecosystem stress (pH, DO%).

| Variable (units) | Trigger value | Samples exceeding trigger values | | |
|---|------------------------|---|--|--|
| Swan Canning | | | | |
| pH (pH units) | ≤7.5 & ≥8.5 | Samples exceeding guideline: 34% below ^W ; 0% above range. | | |
| DO (% saturation) | ≤90 & ≥110% | Samples exceeding guideline: 69% below; 0% above range. | | |
| Peel Harvey | | | | |
| pH (pH units) | ≤7.5 & ≥8.5 | Samples exceeding guideline: 14% below and 6% above range. | | |
| DO (% saturation) | ≤90 & ≥110% | Samples exceeding guideline: 39% below and 16% above range. | | |
| Leschenault | | | | |
| pH (pH units) | ≤7.5 & ≥8.5 | Samples exceeding guideline: 31% below ^s and 0% above range. | | |
| DO (% saturation) | ≤90 & ≥110% | Samples exceeding guideline: 81% below and 0% above range. | | |
| Vasse Wonnerup | | | | |
| pH (pH units) | ≤7.5 & ≥8.5 | Samples exceeding guideline: 14% below and 29% above range. | | |
| DO (% saturation) | ≤90 & ≥110% | Samples exceeding guideline: 57% below and 29% above range. | | |
| Hardy | | | | |
| pH (pH units) | ≤7.5 & ≥8.5 | Samples exceeding guideline: 50% below and 8% above range. | | |
| DO (% saturation) | ≤90 & ≥110% | Samples exceeding guideline: 50% below and 8% above range. | | |
| South coast (includes Oyster and Princess Royal harbours, Wilson, Broke, Walpole Nornalup and Irwin inlets) | | | | |
| pH (pH units) | ≤7.5 & ≥8.5 | Guideline range not exceeded. | | |
| DO (% saturation) | ≤90 & ≥110% | Samples exceeding guideline: 3% below and 0% above range. | | |

^w indicates the majority of samples which exceeded guidelines were collected in winter

^S indicates the majority of samples which exceeded guidelines were collected in summer.

7.2 Estuarine water quality: nutrient guidelines

The nutrient concentrations measured in south-western Australian estuaries were compared with the ANZECC & ARMCANZ (2000) trigger values of physical and chemical stressors using the estuaries ecosystem classification (Figure 14). Nutrient concentrations exceeded guideline values (TN, NOx, NH_4^+ , TP, FRP) in the majority of samples for most estuaries. The south coast estuaries (including Oyster and Princess Royal harbours and Wilson, Broke, Walpole Nornalup and Irwin inlets) were the least disturbed in terms of nutrient concentrations, with only a low percentage of sites exceeding nitrogen and phosphorus guideline values. However, filterable reactive phosphorus (FRP) was elevated in the south coast estuaries with 76% of samples exceeding the guideline. In the Swan-Canning, Peel-Harvey and Leschenault estuaries, TP and FRP exceeded the guidelines more often than nitrogen (total nitrogen (TN), nitrate + nitrite (NOx) and ammonium (NH₄⁺)). The Hardy Inlet exceeded both TN and NOx guideline values for all samples. The number of times the trigger value was exceeded for the oxidised form of soluble nitrogen (NOx) was lower than that of the reduced form (NH₄⁺) in all estuaries except for the Hardy.

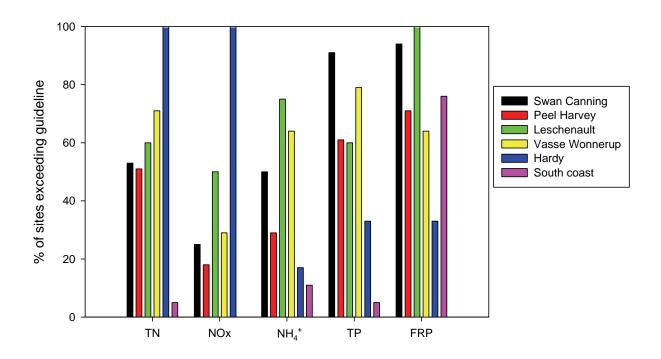


Figure 14 Percentage of sites exceeding ANZECC & ARMCANZ (2000) guideline values for pH, total nitrogen (TN), nitrate + nitrite (NOx), ammonium (NH₄⁺), total phosphorus (TP) and filterable reactive phosphorus (FRP) for south-western Australian estuaries.

7.3 Estuarine water quality: metal guidelines

No specific guidelines for metals in estuaries are available; hence we used the lowest guideline value from either the freshwater or marine guideline values – as recommended by ANZECC & ARMCANZ (2000). The rationale behind this recommendation is that estuaries may contain biota from both fresh and marine systems and thus the lowest value should be adopted to ensure adequate species protection. In most cases the freshwater guideline gave the most conservative value at the 95% protection level, except for Cu and Hg where the marine values were used (because these were more sensitive). Trigger values were adjusted using the hardness algorithm for Cd, Cr, Cu, Ni, Pb, and Zn to derive hardness modified trigger values (HMTVs). The comparison of data with HMTVs was carried out for both total and soluble metal fractions, however biological concern is greater for cases where the soluble fraction exceeds guidelines (as this is more likely to be available to organisms).

A large number of estuarine sites exceeded the non-hardness adjusted trigger value for copper and zinc; however, when adjusted for the high hardness in estuarine waters, these measurements no longer exceeded guideline values. Aluminium and iron were the metals that most frequently exceeded the trigger values for the soluble fraction (Figure 15). The percentage of sites exceeding the trigger values for each estuary is shown in Appendix C. The aluminium guideline was exceeded for the total fraction in 47% to 93% of samples (per estuary) compared with that for the soluble fraction (less than 35%). Trigger values were not (or only rarely) exceeded for mercury, manganese, molybdenum, nickel, lead, selenium, zinc and vanadium. Total arsenic exceeded guideline values in the Peel Harvey, but the soluble concentration was lower than the guideline values in all cases. Total metals were not sampled in the first (summer) sampling period, and this sometimes led to reporting of a higher percentage of samples exceeding trigger values for soluble rather than total metals.

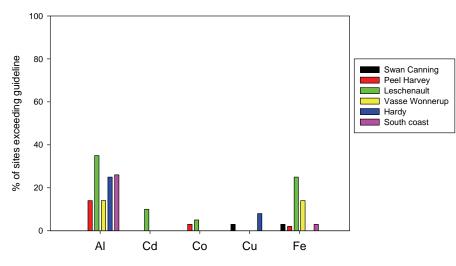


Figure 15 Percentage of sites exceeding trigger values (hardness modified where possible) for soluble aluminium (AI), cadmium (Cd), cobalt (Co) copper (Cu) and iron (Fe) for south-western Australian estuaries.

7.4 Catchment water quality: physical guidelines

The pH and dissolved oxygen measured in south-western catchment areas (main rivers, tributaries and drains) were compared with the ANZECC & ARMCANZ (2000) trigger values for lowland river ecosystem stress (Table 6). Both the low and high guideline values were exceeded for pH, with approximately 30% of samples in the Swan Coastal Plain and south coast catchments lower than pH 6.5. Dissolved oxygen was more often depleted (i.e. <80% saturation) than enhanced (>120% saturation) in the catchment water quality across all catchments (Table 6). The Peel Harvey and the Scott Coastal Plain catchments had the greatest number of samples exceeding the low guideline value with 45% and 39% of samples, respectively, below 80% oxygen saturation.

7.5 Catchment water quality: nutrient guidelines

The nutrient concentrations measured in south-western catchments were compared with the ANZECC & ARMCANZ (2000) trigger values for lowland river ecosystem stress (Figure 16). Nutrient concentrations were consistently higher than guideline values for a significant proportion of samples across all catchments. The Peel Harvey samples exceeded total nitrogen (TN) and total phosphorus (TP) guidelines in approximately 70% of samples. The nitrate and nitrite (NOx) guideline was exceeded in approximately 75% of sites in the Swan Canning, whereas for the Peel Harvey only 3% of sites exceeded the NOx guideline value, but a higher proportion of the latter sites exceeded the ammonium (NH_4^+) guideline value.

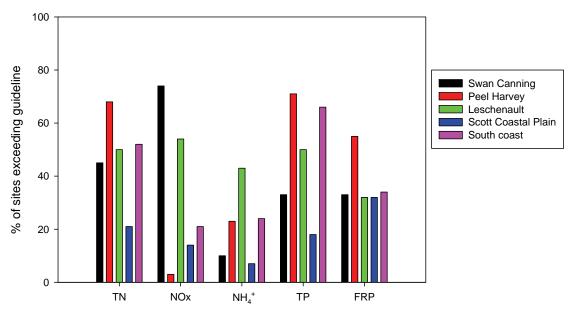


Figure 16 Percentage of sites exceeding guideline values for total nitrogen (TN), nitrate + nitrite (NOx), ammonium (NH₄⁺), total phosphorus (TP) and filterable reactive phosphorus (FRP) for south-western Australian catchments.

Table 6Comparison of catchment (fresh water) water quality data for pH and dissolved
oxygen data with ANZECC & ARMCANZ (2000) trigger values for lowland river
ecosystem stress (pH, DO%).

| Variable (units) | Trigger level | Samples exceeding trigger values |
|---------------------|------------------------|---|
| Swan Canning | | |
| pH (pH units) | ≤6.5 & ≥8.0 | Samples exceeding guideline: 7% below and 17% above range. |
| DO (% saturation) | ≤80 & ≥120% | Samples exceeding guideline: 20% below and 40% above range. |
| Peel Harvey | | |
| pH (pH units) | ≤6.5 & ≥8.0 | Samples exceeding guideline: 13% below and 10% above range. |
| DO (% saturation) | ≤80 & ≥120% | Samples exceeding guideline: 45% below and 10% above range. |
| Leschenault | | |
| pH (pH units) | ≤6.5 & ≥8.0 | Samples exceeding guideline: 7% below and 0% above range. |
| DO (% saturation) | ≤80 & ≥120% | Samples exceeding guideline: 29% below and 7% above range. |
| Scott Coastal Plain | | |
| pH (pH units) | ≤6.5 & ≥8.0 | Samples exceeding guideline: 32% below and 0% above range. |
| DO (% saturation) | ≤80 & ≥120% | Samples exceeding guideline: 39% below and 4% above range. |
| South coast | | |
| pH (pH units) | ≤6.5 & ≥8.0 | Samples exceeding guideline: 31% below and 7% above range. |
| DO (% saturation) | ≤80 & ≥120% | Samples exceeding guideline: 24% below and 17% above range. |

^w indicates the majority of exceedences occurred in winter

^S indicates the majority of exceedences occurred in summer

7.6 Catchment water quality: metal guidelines

Samples analysed to determine metals and metalloids were compared with ANZECC & ARMCANZ (2000) trigger values for 95% freshwater species protection (high reliability) unless otherwise stated. Trigger values were adjusted using the hardness algorithm for Cd, Cr, Cu, Ni, Pb and Zn to produce hardness modified trigger values (HMTVs). Al, Fe, Cu, Co, Zn and Cr in catchment samples were the metals that most frequently exceeded trigger values (Figure 17). The percentage of sites exceeding the trigger values for each catchment can be found in Appendix D.

The Al trigger value was exceeded in most samples across all catchments, with a slightly higher proportion of samples exceeding the trigger value for the total (77–100%) compared with the soluble fraction (45–89%). As, Cd, Hg, Ni, Pb, Se and Mo trigger values were not exceeded in any sample. N and V trigger values were exceeded occasionally. Fe commonly exceeded trigger values throughout all catchments. The south coast catchment samples exceeded the Fe guideline values most often (83% total and 72% soluble).

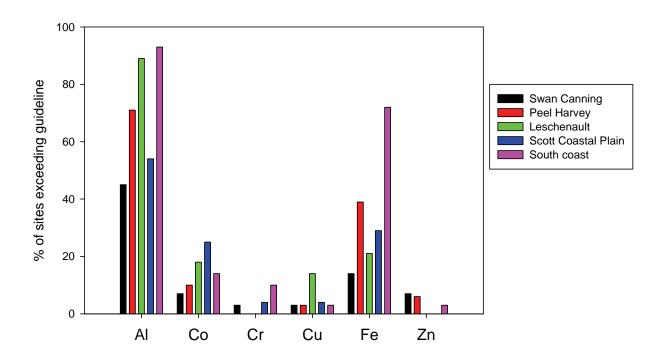


Figure 17 Percentage of sites exceeding trigger values (hardness modified where possible) for soluble aluminium (AI), cobalt (Co), chromium (Cr), copper (Cu), iron (Fe) and zinc (Zn) for south-western Australian catchments.

8 Results: analysis of sulfur stable isotopes

8.1 Background

Identifying sources of sulfur in natural waters is important for managing aquatic ecosystems potentially affected by ASS. Pyritic soils once oxidised may discharge high concentrations of metals, acid and sulfate to surrounding waterbodies. Sulfate in waterways may also be derived from seawater, rainfall, fertiliser and greywater. Analysing the isotopic composition of the sulfur (δ^{34} S) in sulfate may distinguish between these sources and provide a screening tool for waterway management to identify water resources affected by disturbed ASS.

Sulfur occurs naturally as four stable isotopes, of which ³²S and ³⁴S are most abundant (95%) and 4.2% respectively). Differences in the relative abundance of ³⁴S are determined relative to a standard and expressed on the per mil scale; that is, parts per thousand (‰), for δ^{34} S (Clarke & Fritz 1997). Sulfur-34 abundance varies between sulfur compounds due to physical processes and biological cycling such as fractionation by sulfate-reducing bacteria (Kaplan & Rafter 1957; Clarke & Fritz 1997), which results in isotopically light δ^{34} S for the sulfides (i.e. relatively more ³²S). Sulfur isotope ratios measured in brackish and more saline estuarine systems are likely to be mostly a mixture of sulfur from seawater sulfate and sulfate derived from oxidation of pyrite or other reduced sources. The sulfur in seawater has highly positive δ^{34} S values (~21 ‰) (isotopically enriched), whereas sulfur from reduced sources has negative δ^{34} S values (-18 to -52 ‰) (isotopically depleted). Other natural and anthropogenic sources of sulfur may also contribute to the measured isotopic ratio, including geothermal waters rich in sulfur, mineralised organically bound sulfur in soil, leached sulfate minerals and sulfate found in fertilisers and urban areas, as well as detergents, many of which contain sodium sulfate as a filler agent. Additionally, the degradation of sulfur-containing compounds in surfactants (e.g. sulfonates) may also contribute further sulfate to waterways. The average δ^{34} S often varies between different sources of sulfur and this property may be exploited to distinguish contributing sulfur sources in natural waters (Figure 18).

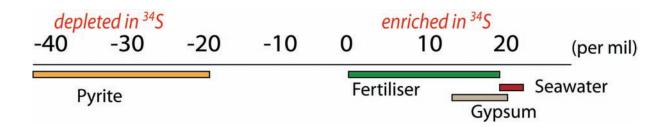


Figure 18 Approximate distribution of δ^{34} S values measured in pyrite, fertiliser, gypsum and seawater.

8.2 Study design and methods

Approximately 300 surface water samples were collected from estuaries and catchment waterways (main rivers, tributaries and drains) in south-western Australia, with locations including freshwater, brackish and saline systems. The Hardy Inlet, Swan-Canning, Peel-Harvey, Leschenault and Vasse-Wonnerup estuaries were sampled, along with the Broke, Walpole Nornalup, Irwin and Wilson inlets and the Princess Royal and Oyster harbours. Catchments were sampled in the Swan-Canning, Peel-Harvey, Leschenault, Vasse-Wonnerup, Scott Coastal Plain and south coast (near Albany) regions.

Filtered water (0.45 µm) for δ^{34} S isotope analysis was collected in HDPE bottles with 10% nitric acid added as a preservative in the field and transported to Monash University for analysis. Before analysis, samples were further acidified and concentrated by evaporation. Sulfate in the sample was precipitated as barium sulfate (BaSO₄) by the addition of barium chloride in excess. The dried BaSO₄ was analysed by continuous flow isotope ratio mass spectrometry. δ^{34} S values are expressed in per mil (‰) notation on the Canyon Diablo Troilite (CDT) standard scale; standardisation of δ^{34} S values used NBS-127 and IAEA-SO5 standards. Precision (1 σ) based on replicate analyses was ±0.2 ‰. Metals, nutrients, major ions and acidity measurements were also determined for these water samples, as described in Appendix B.

The sulfur isotope value was used to determine whether water quality was influenced by other sources of sulfate (acidic drainage, fertiliser or greywater) or dominated by sulfate-reduction processes. Samples were assigned to groups based on the criteria described in Table 7, with these so-called iso-groups categorising samples according to similar influences.

An isotopic mass-balance approach was used to categorise water samples into iso-groups. For more information about this approach see Kilminster and Cartwright (2011).

- Chloride was used as a conservative tracer to calculate the abundance of sulfur in the surface water derived from seawater (the chloride:sulfate ratio in seawater is approximately 7.2 (Clarke & Fritz 1997))
- Where sulfate was present in the water in excess of what could be derived from seawater, a mass-balance calculation was performed to determine what isotope ratio of the extra sulfate would be required to produce the measured δ³⁴S for the sample.
- A threshold for the calculated δ^{34} S for excess sulfur was determined by the lowest isotopic value of fertilisers (0 ‰) from a subset of those commercially available in Western Australia (see Appendix G).
- The two groupings most likely to be representative of acidic drainage (i.e. sulfur from pyrite oxidation) are iso-groups 3 and 5.

8.3 Iso-group proposed meaning and distribution

The proposed six iso-groups can be further grouped based on whether the sites are likely to be natural or disturbed in terms of external sulfur inputs. If disturbed, they can then be further divided into those likely to be receiving acidic drainage, and those dominated by sulfate-reduction.

| lso-group | Measured δ ³⁴ S | Extra sulfate? | Calculated δ^{34} S for excess S | Proposed meaning |
|-----------|-------------------------------|--------------------|---|--|
| 1 | 20-21 ‰ | No | n/a | Sulfur pristine |
| 2 | >21 ‰ | Sulfate missing | n/a | Sulfate-reducing dominant (with only seawater sulfate as a sulfate source) |
| 3 | <19.5 ‰ | Yes | <0 | Sulfate inputs from oxidised acid sulfate soils (acidic drainage) |
| 4a | >21 ‰ | Yes | >20.4 | Sulfate-reducing dominant and extra source of sulfur |
| 4b | <19.5 ‰ | Sulfate missing | n/a | Sulfate-reducing dominant and non-seawater sulfate |
| 5 | <19.5 ‰ | Yes | 0–20.0 | Extra source of sulfate but is a mixed signal (could be a combination of sulfate from oxidised pyrite and/or fertiliser/greywater) |

Table 7Categorisation criteria for iso-groups based on measured δ^{34} S value, presence of
extra sulfate, and the calculated isotopic value expected for the extra sulfate.

Natural sites: Iso-group 1 samples were considered pristine in terms of sulfur sources; that is, only showed signals related to seawater. Iso-group 2 samples were considered sulfate-reducing dominant, but without an extra source of sulfur (only sulfate from seawater) these were likely to be unperturbed systems.

Acidic and probably acidic sites: Samples falling into iso-group 3 were proposed as showing a clear signal of acidic drainage from disturbed ASS. These samples have extra sulfate (on average 9% extra sulfate relative to that expected from seawater) and the calculated δ^{34} S of the extra sulfate is highly negative (i.e. indicating a reduced sulfur source like pyrite). Iso-group 5 often showed significantly more extra sulfate than iso-group 3 (on average 50% extra), but the δ^{34} S of the extra sulfate suggested the sulfur could be derived from multiple sources (e.g. fertiliser, greywater or acidic drainage).

Disturbed, sulfate-reducing: Iso-group 4a and 4b samples have extra sources of sulfur (relative to seawater) contributing to the measured parameters, but sulfate-reduction

dominates in both cases. Samples in these groups come from sites that may pose risk of blackwater events (monosulfidic materials deoxygenating the water column following heavy rainfall).

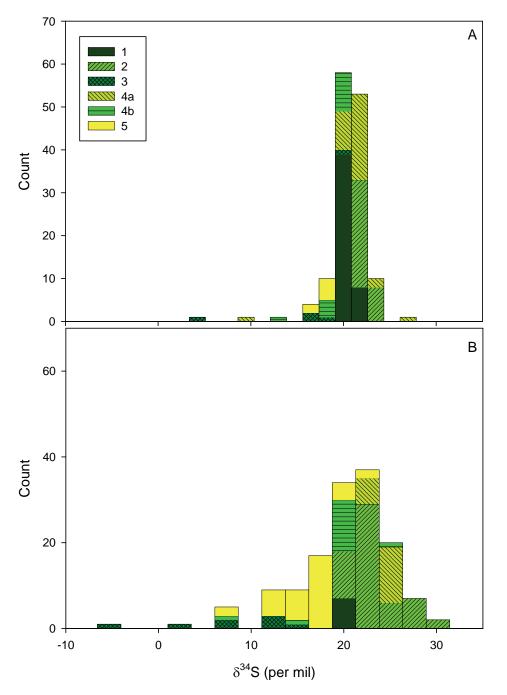


Figure 19 Histogram plots showing distribution of the stable sulfur isotope ratio measured in *A*) estuarine surface water and *B*) catchment surface water. Iso-group category is represented by column colour and shading.

Histogram plots showing distribution of the stable sulfur isotope ratio measured in both estuarine and catchment water, combined with the frequency at which each iso-group

occurs, are shown in Figure 19. These iso-group classifications suggest that sulfate-reducing sites (iso-groups 2, 4a and 4b) dominate most estuaries. The Swan-Canning catchment is dominated by iso-group 5 sites, suggesting multiple sources of external sulfur are contributing to the water quality measured in this catchment. The distribution of sites belonging to each iso-group according to region are shown in Table 8 and Table 9.

| lso-group | Swan Canning | Peel Harvey | Leschenault | Vasse Wonnerup | Hardy | south coast |
|-----------|-----------------|----------------|-------------|-------------------|-------|----------------|
| 1 | 6 | 17 | 3 | 3 | 0 | 8 |
| 2 | 2 | 13 | 3 | 1 | 10 | 2 |
| 3 | 1 | 2 | 3 | 0 | 0 | 0 |
| 4a | 7 | 10 | 6 | 5 | 2 | 7 |
| 4b | 4 | 4 | 2 | 2 | 0 | 4 |
| 5 | 8 | 3 | 3 | 2 | 0 | 9 |

Table 8Distribution of estuarine sites within iso-group categories across regions.

| Table 9 | Distribution of catchment (fresh water) sites within iso-group categories across |
|---------|--|
| | regions. |

| lso-group | Swan Canning | Peel Harvey | Leschenault | Scott Coastal Plain | south coast |
|-----------|-----------------|----------------|-------------|---------------------------|----------------|
| 1 | 0 | 0 | 3 | 2 | 0 |
| 2 | 4 | 15 | 6 | 17 | 12 |
| 3 | 1 | 1 | 1 | 1 | 4 |
| 4a | 3 | 4 | 8 | 2 | 2 |
| 4b | 0 | 2 | 5 | 2 | 6 |
| 5 | 22 | 5 | 4 | 3 | 5 |

8.4 Relevance of iso-groups

From an environmental management viewpoint, the most significant concern in relation to drainage from ASS is the input of acidic metal-rich water. This may deleteriously affect biota and ecosystem function (rather than the input of extra sulfate). Therefore, if the screening

tool is to be useful to management, we would also require it to inform us about metal distribution in the water, and these ideas are explored within this section.

Data analysis of the multivariate dataset collected was undertaken to see if the iso-group classification was meaningful with respect to the other variables analysed. Analysis was performed with PERMANOVA in the PRIMER v6 and PERMANOVA + software package. Data was transformed (log (x+1)), normalised and euclidian-distance resemblance matrices determined. PERMANOVA analyses were run on subsets of variables with electrical conductivity as a co-variate and iso-group as a fixed factor. It was important to consider electrical conductivity (or salinity) as a co-variate because the solubility of metals and pH is strongly affected by the salinity of the water, and analysis showed the co-variate was significant (p<0.0001) for all analyses run. Three variable subsets were considered: acidity, soluble metals, and total metals (Table 10). Within the acidity subset, variables considered to be describing some aspect of the acidity were included; for example, total acidity, alkalinity, pH and filtered iron and aluminium. Analysis of the data by PERMANOVA for the acidity subset was not significant for iso-group category, however both metal subsets showed significant differences between iso-group categories (p<0.05 and p<0.005 for filtered and unfiltered metals respectively).

When Fe and AI concentrations within water samples were considered individually, there was a relationship with the iso-groups. In estuarine samples, for both filtered AI and filtered Fe, iso-group 3 had a greater concentration than the other iso-groups (Figure 20). Iso-group 3 had an average of 0.12 mgL⁻¹ filtered AI compared with <0.05 mgL⁻¹ for the other groups. Similarly, filtered Fe was 0.49 mgL⁻¹ for iso-group 3 and <0.15 mgL⁻¹ for the other groups. For unfiltered AI, iso-group 3 and 5 were also higher on average (0.41 and 0.28 mgL⁻¹ respectively) than the other iso-groups (<0.18 mgL⁻¹). Unfiltered Fe was also higher for isogroup 3 and 5 (1.0 and 0.91 mgL⁻¹ respectively) than the other iso-groups (<0.54 mgL⁻¹). These variations are indicative only - due to small sample sizes in some of the iso-groups statistical significance was not met. Similarly, in the catchment samples iso-groups 3 and 5 had a higher average concentration of AI than the other iso-groups (Figure 21). For filtered Al, iso-groups 3 and 5 had an average concentration of 0.55 and 1.7 mgL⁻¹ respectively compared with <0.26 mgL⁻¹ for the other iso-groups. For unfiltered AI, iso-groups 3 and 5 had an average concentration of 0.74 and 2.1 mgL⁻¹ respectively compared with <0.45 mgL⁻¹ for the other iso-groups. There did not seem to be a strong pattern for iron concentrations in catchment samples across iso-group categories (Figure 21).

Median concentrations of many metals (AI, Co, Cr, Cu, Fe, Mn, Ni, V, Zn) and phosphorus were higher in ASS-affected samples (iso-group 3) than in other samples not influenced by ASS (Kilminster & Cartwright 2011). Additionally, total phosphorus and filterable reactive phosphorus had higher median concentrations (0.14 mgL⁻¹ and 0.057 mgL⁻¹ respectively) for iso-group 3 samples compared with the median concentrations for the other iso-groups combined (0.038 mgL⁻¹ and 0.01 mgL⁻¹ for TP and FRP).

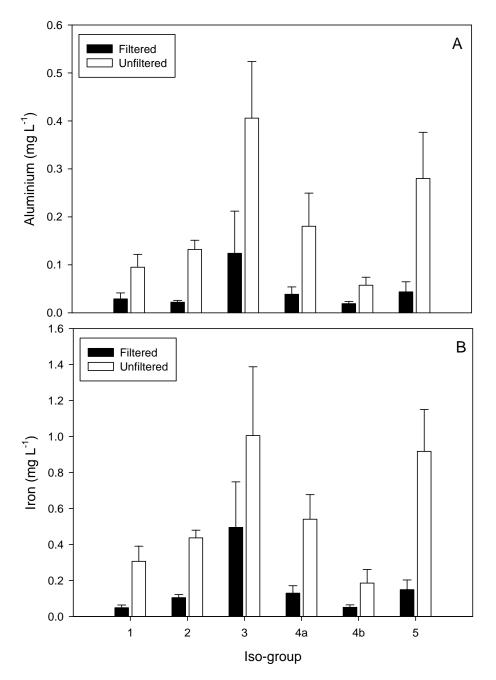


Figure 20 Concentrations of A) aluminium, and B) iron within estuarine surface water samples, where black columns represent the filtered (soluble) metal fraction and white columns represent the unfiltered (or total) metal fraction (mean + st. err.).

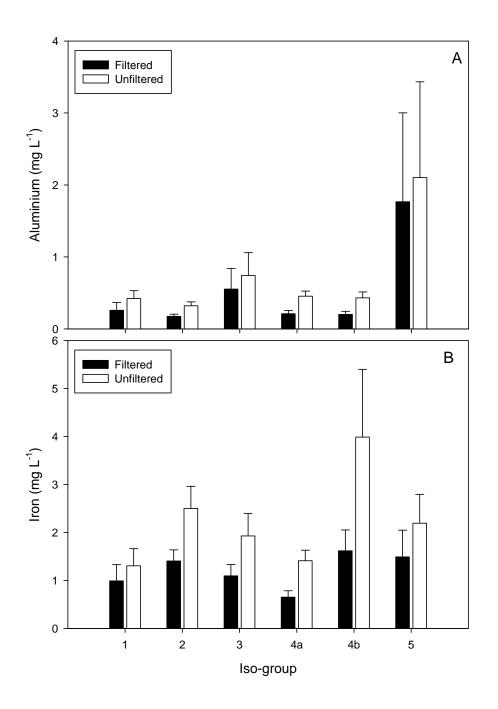


Figure 21 Concentrations of A) aluminium, and B) iron within catchment surface water samples, where black columns represent the filtered (soluble) metal fraction and white columns represent the unfiltered (or total) metal fraction (mean ± st. err.).

| Data Subset | Variables | Pseudo-F | <i>p</i> value |
|-----------------|---|----------|----------------|
| Acidity | total acidity, alkalinity, pH, Fe (filt) and AI (filt) | 1.62 | 0.10 |
| Filtered metals | filtered Al, As, Cd, Co, Cr, Cu, Fe, Mn, Mo, Ni, Pb, V, Zn | 1.72 | <0.05 |
| Total metals | unfiltered Al, As, Cd, Co, Cr, Cu, Fe, Mn, Mo, Ni, Pb, V, Zn | 2.00 | <0.005 |

Table 10 Results of PERMANOVA analyses of data for variable subset with iso-group as a
fixed factor and conductivity as a co-variate.

8.5 Conclusion regarding iso-groups

The categorisation of samples into iso-groups appears to provide a useful signal as to whether acidic drainage is influencing a waterbody. This methodology is more sensitive for identifying acidic drainage than other indicators such as the chloride:sulfate ratio or total acidity. These other indicators of acidic drainage are less suited to identifying the influence of ASS drainage in an estuary since the concentrations of sulfate from seawater are likely to override the input from acidic drainage and any acidity is likely to be neutralised by the seawater. This difference in sensitivity is seen in the plot of chloride:sulfate ratio *vs* iso-group for estuaries and catchments (Figure 22) where only the ratio for iso-group 5 (for catchments) is sufficiently low (i.e. <4) to suggest the influence of acidic drainage.

The iso-group categorisation also identified sites where sulfate-reduction was probably stimulated by a supply of excess sulfur (iso-group 4a and 4b). Approximately 59% of sites sampled appear dominated by sulfate-reduction (i.e. iso-groups 2, 4a and 4b), with half of these likely to have enhanced sulfate-reduction resulting from an anthropogenic source of additional sulfur (i.e. iso-group 4a and 4b). Possible sources of extra sulfur are acidic drainage as well as urban and agricultural sources such as fertiliser and greywater. It is also possible that sulfate-reduction may be enhanced through organic-matter enrichment associated with eutrophication in areas where sulfate is not limiting.

The process of sulfate-reduction efficiently removes metals from the water column, binding them into the sediment. However when this process is accelerated, the reduced sulfur is only bound in an easily oxidisable form, commonly called monosulfidic black ooze (MBO). MBO is a gelatinous black muck that accumulates on the sediment surface and is readily re-oxidised, stripping oxygen from the water column and remobilising the metals that were previously bound (Burton et al. 2006). MBO formation in drains and streams can cause major environmental problems as heavy rainfall mobilises the MBO – deoxygenating the water and resulting in blackwater events. Reported blackwater events on Australia's east coast have been catastrophic with hundreds to thousands of fish and prawns reported killed as a result of just one event (Sammut et al. 1995; Russell & Helmke 2002; ASSAY 2008). Further

investigation is required to assess the risk of blackwater events in Western Australian waterways.

Iso-group 3 is most likely influenced by acidic drainage. For catchments, the iso-group 3 samples often exceeded AI, Fe, Co and Cr and nutrient guidelines (Appendix D and E). Iso-group 5 samples are potentially influenced by acidic drainage, but the isotopic signature is mixed, suggesting that sources other than oxidised pyrite are potentially influencing the signal. Catchment samples from iso-group 5 frequently exceeded AI, Fe, Cr, Cu, Co, Zn and nutrient guidelines (Appendix D and E).

The iso-groups do not indicate the intensity of the acidic drainage influence. Also they do not accurately predict the occurrence of metals in a water sample. It seems plausible that this is due in part to the many other potential sources of metals from urban, agricultural and industrial land uses that may influence waterways.

The Western Australian coastal environment is particularly suited to the use of sulfur stable isotopes as an indicator of disturbed ASS. The landscape is geologically old, with no volcanic or geothermal inputs of sulfur. Additionally, unlike many other regions (especially those in the northern hemisphere) acid rain has little influence, while rainwater in Western Australia has δ^{34} S values very similar to seawater sulfate (18.2–20.0 ‰). Gypsum from Western Australia also reflects a seawater sulfur source (Bowen & Benison 2009). Australian fertiliser δ^{34} S values were previously reported to be between +6.4 and +8.5 ‰ (Mizota & Sasaki 1996), however the current study showed a greater range was present in commercially available fertilisers (-0.1 to +18.7 ‰). Regardless, there remains a large separation of the δ^{34} S values of sulfate from oxidised pyrite, seawater, precipitation and fertiliser, which has been used in the current screening tool for ASS-affected waterways. There is scope for this tool to be employed in other environments world-wide, however the δ^{34} S values of sulfur sources would need to be locally determined, and the iso-groups' threshold values adjusted accordingly.

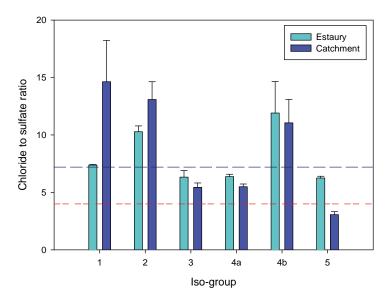


Figure 22 Chloride:sulfate ratio (mass) of estuarine and catchment water quality categorised by iso-group (mean + standard error). The blue long dashed line indicates the ratio (7.2) observed in seawater, and the red short dashed line indicates a value below which the influence of acid sulfate soil drainage is suggested.

9 Results: hotspot sites

A few sites within the dataset appeared to be significantly different to the rest of the dataset, and thus were considered hotspot sites. The water quality and appearance of these sites are discussed below. Sites showing extremely high values of aluminium and iron (>90th percentile values) are also highlighted.

9.1 Rich Road drain, near Capel

Site 6100095 (also known as BY01 or Minninup drain) is a drain that flows southward through low-lying agricultural land near Capel and the coast (Figure 23). Acidic water was identified in this drain, and water was sampled for chemical analysis on six occasions between 12 August and 25 September 2007. The drain had dried up by December 2007.



Figure 23 Northern view of Rich Road drain in September 2007.

Water quality was consistently poor. Al concentrations ranged from 2–4.5 mg L⁻¹ for soluble, and 2.1–4.7 mg L⁻¹ for total. Both ranges exceeded the 90th percentile values of 0.51 and 0.93 mg L⁻¹ observed for all catchment samples for soluble and total Al respectively (Table 4). Iron concentrations were also extremely elevated, ranging from 130–230 mg L⁻¹ and 140–240 mg L⁻¹ for soluble and total Fe respectively. The 90th percentile value for the catchment water quality was 2.9 mg L⁻¹ for soluble iron, and 4.9 mg L⁻¹ for total iron (Table 4). In addition

Fe precipitate (floc) was observed to be highly abundant in surficial sediments present in the bottom of the drain (Figure 24). Mn concentrations were also high in the drain, with a range of 3–4.1 mg L^{-1} for soluble, and 3.2–4.5 mg L^{-1} for total; again much higher than the 90th percentile value for catchment data of 0.13 and 0.14 mg L^{-1} for soluble and total Mn respectively (Table 4).

Low pH (2.7–3) and extremely high total acidity measurements (410–850 mg CaCO₃.L⁻¹) also characterised the water from the site. Sulfur isotope values measured in September 2007 were slightly lower than seawater (average ~18.2 ‰), and excess sulfur was present (97% more sulfur than would be expected from seawater mixing alone). For the September sampling, the site was classified as iso-group 5, and this indicates the extra sulfate is likely to have originated from a mixture of sources (e.g. fertiliser and acidic drainage).



Figure 24 Surficial sediments within the drain (site 6100095) showing the presence of iron floc.

9.2 Swan-Canning region

The Swan-Canning catchment had a large proportion of sites assigned to iso-group 5, suggesting potential impacts from acidic drainage with large quantities of extra sulfate. One of these, site BWMDBPI (a drain discharging to the Canning Estuary) (Figure 25) appeared impacted by disturbed ASS with 90% extra sulfate, relatively high total acidity (31 mg $CaCO_3.L^{-1}$) and high total Fe (11 mg L^{-1}) and total AI (3.1 mg L^{-1}). Only one site within the Swan-Canning region, SRBDS, showed a clear acidic drainage signal (i.e. iso-group 3), with 26% extra sulfate (Figure 26).



Figure 25 Location of BWMDBPI site in Swan-Canning catchment, adjacent to the Canning River.



Figure 26 Location of site SRBDS classified as iso-group 3 within the Swan-Canning region.

9.3 Peel-Harvey region

Most sites in the Peel-Harvey region were influenced by sulfate-reduction processes (mainly iso-groups 2 and 4a). Fe and Al were high at many sites within the Peel-Harvey catchment when compared with the 90th percentile values. Sites MR05005, MR05010, MR01010 and PATRDTRIB had concentrations of Fe (either soluble or total) higher than the 90th percentile value. Al concentrations (both total and soluble) were higher than the 90th percentile value for catchment samples MPH003, 613053, HR02005 and PHS7. Soluble Al was high at

SR04014 and 614120 and total AI was high at MR01010 and PATRDTRIB. See Figure 27 for the locations of these sites.

The MPH003 catchment sample showed a clear acidic drainage signal. This site is located on a drain that discharges directly into the Peel-Harvey estuary (Figure 28). The site was classified as iso-group 3 with 32% additional sulfate than would be expected from seawater alone. High total acidity (28 mg CaCO₃.L⁻¹) and high soluble and total aluminium were also measured at this site. Estuarine sites PHRS4-s and MPH015-w also exhibited a clear acidic drainage influence (iso-group 3) (Figure 29).

9.4 Leschenault region

Several sites in the Leschenault catchment displayed elevated Fe and Al. Sites exceeding the 90th percentile boundary level for soluble AI were ACID02 and LESCH04, and for total AI were GBC02 and ACID13 (Figure 30). Site ACID17 had extremely elevated Fe, both soluble and total, with measurements of 9.9 mg L⁻¹ for soluble Fe and 21 mg L⁻¹ for total Fe (both higher than the 90th percentile level of 2.9 mg L⁻¹ for soluble Fe and 4.9 mg L⁻¹ for total Fe (Table 4). The ACID17 sample site is located on the Collie River (Figure 31) downstream from a site discharging contaminated groundwater from an abandoned coal mine (Webb 2009).

Three estuarine sites (6121173-w, 6121162-w and 6121161-w) and one catchment site (LESCH05) were categorised as iso-group 3 (Figure 32), indicating they have received acidic drainage waters. Of these sites, the acidic drainage influence appeared strongest for site 6121173 (also known as Parkfield drain) with 43% extra sulfate than what would be expected from seawater alone. Site 6111043 had the next-greatest influence of acidic drainage with 16% extra sulfate present.

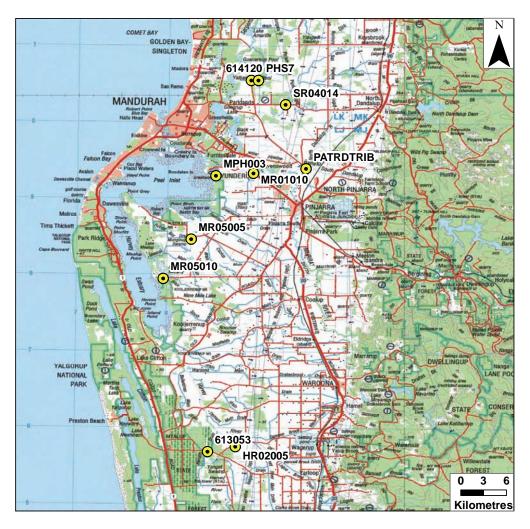


Figure 27 Catchment site locations in the Peel-Harvey region exhibiting extremely high concentrations of aluminium and/or iron in the water samples.



Figure 28 Photograph showing sampling at site MPH003, where the drain enters Peel-Harvey estuary.

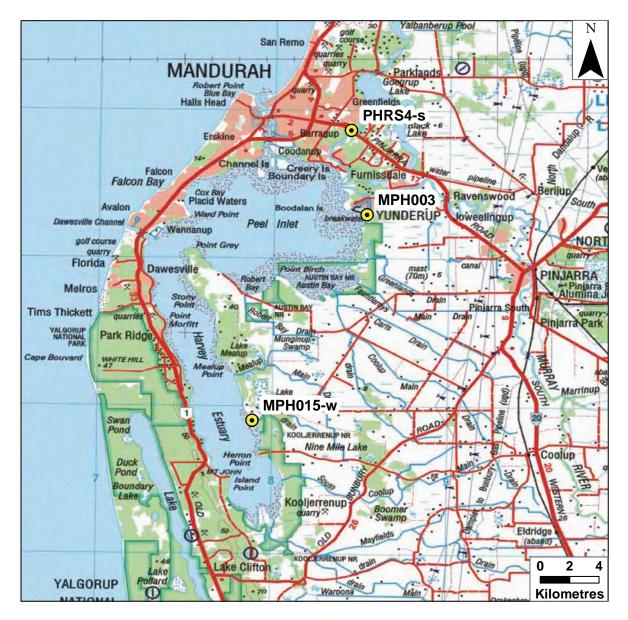


Figure 29 Location of sites in the Peel-Harvey region with clear acidic drainage influence (iso-group 3).



Figure 30 Location of sites in the Leschenault catchment displaying elevated aluminium concentrations in water samples.

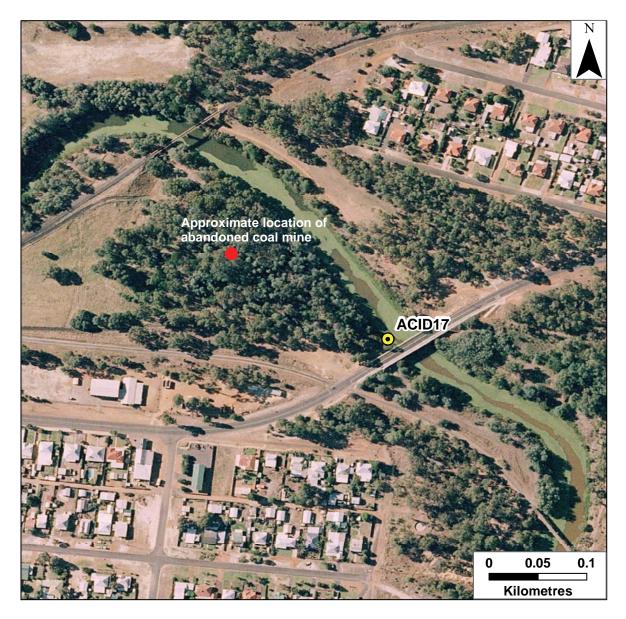


Figure 31 Location of site ACID17 along the Collie River, downstream from an abandoned coal mine.



Figure 32 Location of iso-group 3 sites within the Leschenault region.

9.5 Scott Coastal Plain

Several sites in the Scott Coastal Plain catchment showed elevated Fe and Al. Site 6091758 exceeded the 90th percentile value for Al with extremely high concentrations of 4.8 mg L⁻¹ and 5.9 mg L⁻¹ for soluble and total Al respectively. Sites 6091222, 6091223 and 6091596 exceeded the 90th percentile boundary value for Fe with concentrations of 3.8–4.2 mg L⁻¹ for soluble Fe and 5.3–8.8 mg L⁻¹ for total Fe. See Figure 33 for the locations of these sites with high concentrations of Fe or Al. One site showed clear signals of acidic drainage (i.e. iso-group 3), which was 6091755, with 26% extra sulfate (Figure 34).

9.6 South coast region

A number of sites in the south coast catchment displayed elevated AI and Fe. Site WOOD1 (Figure 35) had the most extreme AI concentrations (46 mg L⁻¹ and 50 mg L⁻¹) compared with the 90th percentile values of 0.51 and 0.93 mg L⁻¹ for soluble and total AI. Site TILP012 (also known as Elleker Grassmere drain) also had extremely elevated AI (11 mg L⁻¹ and 12 mg L⁻¹ for soluble and total AI). Sites TILP001 and TILP047 also exceeded the 90th percentile value for total AI. Elevated Fe was found at sites INFLOW, LEAY, MANCK, SUN, TILP012, TILP047 and WOOD1. Fe was highest at TILP012, with 19 mg L⁻¹ soluble Fe, and 21 mg L⁻¹ total Fe. Site locations can be seen in Figure 36 and Figure 37. Total acidity was particularly high for WOOD1 and TILP012 with measurements of 290 mg CaCO₃ L⁻¹ and 160 mg CaCO₃ L⁻¹ respectively. WOOD1 and TILP012 were the only two sites with pH <4 (2.98 and 2.84 respectively).

Four sites within the south coast region had a clear acidic drainage signal (iso-group 3) (Figure 38), and three of these are adjacent to estuarine environments. Sites LOWKG, TILP001 and MRD display a similar intensity of signal with 25–45% extra sulfate, while by comparison the extra sulfate signal of MTBARKRD was relatively minor (<1%) at this sampling time.



Figure 33 Location of sites on the Scott Coastal Plain with very high concentrations of either aluminium or iron.



Figure 34 Location of sites with iso-group 3 on the Scott Coastal Plain.



Figure 35 Site WOOD1, drain running along Woodies Road near Lake Powell.



Figure 36 Location of sites with elevated aluminium and/or iron in water samples within Torbay catchment near Albany.

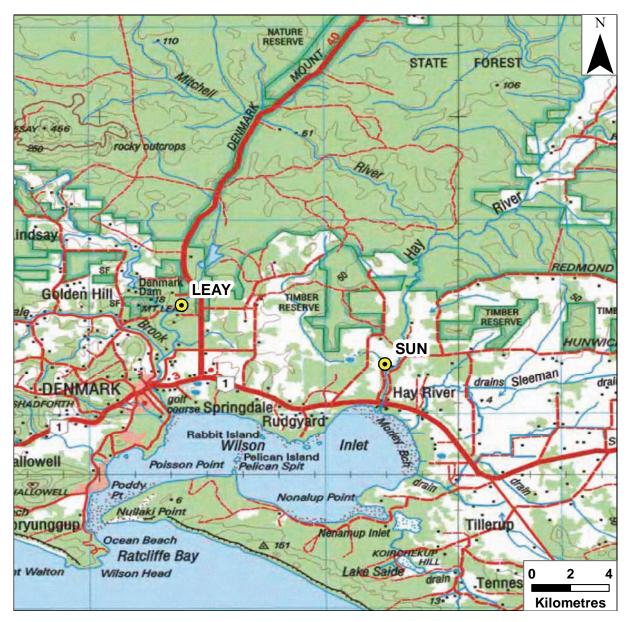


Figure 37 Location of sites SUN and LEAY near Wilson Inlet with elevated iron concentrations in water samples.



Figure 38 Location of sites with iso-group 3 in Albany region, with sites near Oyster and Princess Royal harbours and Wilson Inlet.

10 General discussion

10.1 Comparison to Australia's east coast

On Australia's east coast, deteriorated water quality and impacts on aquatic ecosystems associated with drainage from disturbed ASS have been well documented. Often these events have been rather dramatic. For example, the Richmond River experienced a massive fish kill of thousands of fish and prawns in January 2008, in a blackwater event that saw dissolved oxygen levels of the estuarine water remain below 1 mg L⁻¹ for about a week (ASSAY 2008). In another example, heavy rains in 1987 resulted in a 23 km stretch of the Tweed River Estuary acidifying – with possibly more than 1000 tonnes of aluminium being mobilised and the river remaining almost devoid of life for a further 18 months (White et al. 2006). Events at this scale have not been reported in Western Australia. Smaller episodic acidification events have also been reported on the east coast (Sammut et al. 1996; Russell & Helmke 2002).

The range in deteriorated water quality reported to be associated with acidic discharge from disturbed ASS in Australia's eastern states is shown in Table 11. While this list of locations is not exhaustive, it does provide a contextual comparison for data collected in this study. Typically pH values from ASS-affected drains and streams on the east coast were much lower (pH <4) than observed in the current study, where only two samples out of more than 300 samples were below pH 4. A couple of sites from this Western Australian study had elevated AI concentrations in the range observed for the east coast, (i.e. WOOD1 had soluble iron of 46 mg L⁻¹ and TILP012 had soluble iron of 11 mg L⁻¹), however higher extreme values were still reported from the east coast study sites of Firewood Creek, Qld, and Clothiers Creek, NSW. Fe showed a similar trend, with a number of sites displaying elevated Fe within the range reported in the literature (e.g. soluble iron was between 3–19 mg L⁻¹ for 14 sites in the current study), but higher extremes were still reported at Firewood Creek, East Trinity and Clothier Creek. Only two sites, PATRDTRIB and 6091223, showed significantly elevated Mn, with values between 2.2–2.6 mg L⁻¹ for soluble and total Mn.

In general, very low CI⁻:SO₄²⁻ ratios were observed for the study sites in the eastern states. CI⁻:SO₄²⁻ ratios lower than 4 were observed in 31 sites in the current study, of which 29 sites were iso-group 5 (suggesting sulfate influence from multiple sources, not only acidic drainage). Interestingly, no iso-group 3 sample had a low sulfate:chloride ratio (<4), suggesting that these clear signals of acidic drainage reflect chronic rather than acute acidic drainage.

10.2 Acidity measurements

Measures of pH quantify the concentration of hydrogen ions (H^+) in solution, and only accurately represent the acidity from fully dissociated acids (i.e. strong acids). Acidity in ASS drainage water is not fully described by pH alone. Acidic drainage waters may also contain metal ions that produce H^+ by hydrolysis and oxidation reactions. The hydrolysis and oxidation of one mole of Fe is estimated to produce two moles of H^+ ; and one mole of Al, three moles of H^+ (Cook et al. 2000). For south-western Australian waters, few samples had

low pH values (only two samples were less than pH 4). Much of the total acidity measured in the water samples is likely to be associated with the hydrolysis of Fe and Al. To estimate the proportion of the contribution to acidity by Fe and Al we assume:

- 1 that soluble Fe and soluble AI are in the form capable of producing acidity (i.e. Fe^{2+} , AI^{3+}), and
- 2 that each mole of Fe will produce two moles of H^+ and each mole of AI will produce three moles of H^+ .

From analysis of the data in this way we deduce that metal acidity was a significant contributor to the total acidity measured. Soluble Fe and Al concentrations explained 68% and 80% of the total acidity measured for iso-group 3 and iso-group 5 in catchment samples respectively. These assumptions probably over-represent the contribution of metal hydrolysis and oxidation to the measurements of total acidity, however they highlight the importance of metal hydrolysis and oxidation to acidification of waterways further downstream of an ASS disturbance source.

| Location | рН | Iron concentration (mg L ⁻¹) | Aluminium concentration (mg L ⁻¹) | Other metals (mg L ⁻¹) | Other variables | Reference |
|--------------------------------------|---------|--|---|--|---|-------------------------|
| Magazine Creek, Qld | 2.5–4 | 8–55 ⁸ | 5–50 ⁸ | | | (Russell & Helmke 2002) |
| Firewood Creek, Qld | 2.5–4 | 5–186 ^S | 15–75 ⁸ | | Sulfate 1000–2000 mg L^{-1} Nitrate = 0.08 mg L^{-1} | (Russell & Helmke 2002) |
| | | | | | Periodic deoxygenation (<20% saturation). | |
| Trinity Inlet, Qld | 4.5-7.5 | 0.2–4 ^S | 0.05–0.35 ^S | | | (Russell & Helmke 2002) |
| Tuckean Swamp, NSW | 3–4 | 0.3–2.4 ^S | 1.8–7.3 ^S | Mn: 0.3–0.9 ^S Cu: 0.02–0.2 ^S Zn: 0.03–0.2 ^S | Sulfate: 50–250 mg L ⁻¹ Cl ⁻ :SO ₄ ²⁻ : 0.35–1.2 | (Sammut et al. 1996) |
| Rocky Mouth Creek floodplain, NSW | 3.2–3.7 | 0.7–11.2 ^T | 0.8–1.5 ^T | Mn: 0.4-1.0 ^T Zn: 0.02– 0.08 ^T | Total acidity: 41–97 Cl ⁻ :SO ₄ ²⁻ :0.36–0.7 | (Lin et al. 2004) |
| East Trinity, Qld | <4 | 3.1–120 ^S | 0.51–11 ^s | Zn: 0.19–0.3 ^S | | (Cook et al. 2000) |
| Pimpama, Qld | ~1 | 0.07–33 ^s | 2.5–20 ^S | Zn: 0.04– 0.22 ^S | | (Cook et al. 2000) |
| Tweed River, NSW | 3.2–5 | 3–8.6 ^S | 1.2–6.2 ^S | | Sulfate: 155 mg L ⁻¹ Cl ⁻ :SO ₄ ²⁻ : 0.4 | (Willett et al. 1993) |

 Table 11
 Ranges of water quality values for water reported to be affected by acid sulfate soils within Australia's eastern states

| Richmond River, NSW | 3.5–7.2 | 0.1–1.3 ^S (0.4– 4.4 ^T) | 1.6–9.7 ^S (0.3– 9.2 ^T) | | Cl ⁻ :SO ₄ ²⁻ : 0.4 –7.9 | (Roach 1997) |
|----------------------------------|---------|--|--|---|--|---------------------|
| McLeods Creek, NSW | 3.2–7.9 | 0–6.7 ^T | 0–22 [⊤] | Mn: 0.05–2.9 ^T Zn: 0–0.4 ^T | Sulfate: 864 mg L ⁻¹ Cl ⁻ :SO ₄ ²⁻ : 3.00 | (Green et al. 2006) |
| Clothiers Creek, NSW | 2.6–3.4 | 13.9–123 ^T | 48.6–254 ^T | Mn: 1.4–8.3 ^T Zn: 0.52–2.0 ^T | Sulfate: 1900 mg L ⁻¹ Cl ⁻ :SO4 ²⁻ : 0.02 | (Green et al. 2006) |
| S soluble metals determined | | | | | | |

total metals determined

10.3 Aluminium and iron toxicity

Aluminium was high at most sites, with almost all sites in the south coast and Leschenault catchments exceeding the ANZECC & ARMCANZ (2000) trigger value for 95% freshwater species protection. All is known to have deleterious effects on both plants and animals. Adverse biological effects of short-term pulses of AI and acidity have been reported for Atlantic salmon (Kroglund et al. 2008). Al is known to limit crop production by the inhibition of cell elongation and to some extent cell division. Some plants resist these effects by excreting carboxylates (e.g. malate or citrate) through their roots, which results in detoxification of AI by the formation of organic complexes (Jones 1998). Al may also be complexed with organic compounds, which reduces toxicity within humic-rich streams or wetlands. The toxicity of Al is expected to increase at extreme pH levels (<5.5 or >9) and high temperatures, and to reduce if AI forms complexes with fluoride, citrate or humic substances. Toxicity is also reduced in the presence of silicon and with high water hardness (ANZECC & ARMCANZ 2000). All is a highly abundant element within deeply weathered soils throughout southwestern Australia (Division of Soils CSIRO 1983). As such, Al may be naturally mobile within the environment, however anthropogenic activities and ASS disturbance may also contribute considerable quantities of AI. If Western Australian native flora and fauna have evolved in AIrich environments, it is likely they will be tolerant to high concentrations of AI (i.e. above current guideline values). Without undertaking ecotoxicological testing specific to Western Australian native species, we cannot comprehend how extensively the state's ecosystems are threatened by high AI concentrations.

There is no ANZECC & ARMCANZ guideline for iron. Instead, comparisons of Fe concentrations within this study have been made with new guidelines proposed in British Columbia, Canada (Phippen et al. 2008) of 1 mg L⁻¹ for total Fe, and 0.35 mg L⁻¹ for soluble Fe. One of the main arguments for including a separate guideline for total Fe was to consider both the direct toxic effects of Fe (usually in its dissolved ferrous form), and the indirect effects on habitat and species displacement that generally relate to total Fe concentrations (often as Fe hydroxide precipitates). Factors such as hardness and humic acid complexation of Fe also influence toxicity. Again, without undertaking ecotoxicological testing of Western Australian native species, the threat posed by high Fe concentrations will remain uncertain.

10.4 Chronic acidic drainage influence

Most sites in south-western Australia exhibiting 'acidification' from ASS appear as a chronic influence on waterways rather than acute expressions. Very low pH values were rarely observed in the dataset. Although a few sites were identified where strong signals of ASS exist (e.g. WOOD1, TILP012), in most cases the volume of water associated with these drains is small relative to the volume of the receiving waterbodies.

The impacts of acidic drainage are most likely to express as chronic long-term effects on aquatic ecosystems. A number of sub-lethal effects on aquatic organisms may occur as a response to acidic drainage. For example, inputs of iron were shown to increase the growth rate of the toxic cyanobacteria *Lynbya majuscula* (Ahern et al. 2006). Also, the decrease of alkalinity by acidic discharges may inhibit growth of crustaceans within estuaries (Sammut et

al. 1996). Acidification also appears to favour acid tolerant species, such as mosquitoes (McCullough & Horwitz In Press), larval biting midges (family Ceratopogonidae), water boatmen (family Corixidae) and water fleas (family Macrothricidae) (Sommer & Horwitz 2009). These effects may ultimately modify the community structure of species within the aquatic ecosystem, with subsequent effects on the trophic web.

Determining a sample's iso-group appears to provide a useful categorisation in terms of the sulfur cycle, as well as a useful screening tool to decide whether a site is likely to be receiving acidic drainage. The other advantage of this method is that the 'fingerprint' of acidic drainage still remains, even if the actual acidity has been neutralised. However, further work is required to fully understand the mixed signals, such as for iso-group 5, where multiple sources of external (non-seawater) sulfur are possible. Disentangling multiple sulfur signals seems most important for the Swan-Canning catchment, where more than 70% of sites were grouped into iso-group 5.

10.5 Sulfate-reduction in waterways

The process of sulfate-reduction efficiently removes metals from the water column, binding them into the sediment. However when this process is accelerated, the reduced sulfur is only bound in an easily oxidisable form (metastable monosulfides). Acid volatile sulfur abundance is higher in drains with gentle flow that are affected by ASS leachate (Bush et al. 2004). Monosulfide formation in drains and streams can cause major environmental problems as heavy rainfall mobilises the monosulfide, deoxygenating the water and resulting in blackwater events.

On the basis of the iso-group determination, sulfate-reduction was the dominant process for two-thirds of all samples (i.e. iso-groups 2, 4a and 4b). Of these samples, approximately two-thirds of estuarine samples and half the catchment samples were associated with extra (other than seawater) sources of sulfur (i.e. they were iso-group 4a and 4b). It is likely that the supply of extra sulfate to these waterways results in enhanced monosulfide production, however further work is required to confirm this. In estuaries, with their large natural supply of sulfate from seawater, excess sulfate is unlikely to promote further sulfate-reduction. Instead, high organic matter loading (associated with eutrophication) may enhance monosulfide production. It seems possible that within an estuary such as the Peel Harvey, the effects of eutrophication and disturbance of ASS act synergistically to produce the extremely reducing conditions observed throughout much of this region.

11 Conclusion and recommendations

Disturbance of ASS is likely to be having a chronic (rather than an acute) effect on southwestern Australian waterways. Sites likely to be influenced by acidic drainage were found in all investigated regions. Water at catchment sites was more likely to exhibit acidity than the estuarine sites (perhaps due to sulfate dilution). It seems that acidic drainage in southwestern Australia more often presents as a trickle, whereas the examples from the eastern states point to large quantities of acid being produced behind floodgates. It is possible that this difference is related to the lower rainfall experienced in Western Australia. Additionally, the sandy nature of ASS in south-western Australia (Degens & Wallace-Bell 2009) suggests that groundwater resources may be more susceptible to acidification than in other areas with clay-dominated soils. In Western Australia the impacts of acidic drainage are most likely to express as chronic long-term effects on ecosystems – potentially affecting aquatic community structure, aquatic trophic web interactions and groundwater quality.

The categorisation of samples into iso-groups, by using sulfur isotopic analysis combined with determinations of sulfate and chloride concentrations, appears to provide a useful screening tool for assessing the influence of acidic drainage. Median concentrations of many metals and phosphorus were higher in ASS-affected samples (iso-group 3) than in other samples not identified as influenced by ASS. This methodology is more sensitive for identifying acidic drainage than other indicators previously used in ASS management. Forty-one out of the 313 sites sampled were identified as hotspot sites by either sulfur isotopic analysis or by unusually high concentrations of AI and Fe, relative to the rest of the baseline dataset. These sites were found in all the geographic regions investigated in this study.

Sulfate-reduction was probably enhanced by the supply of extra sulfate (relative to seawater) in 42% of estuarine sites and 33% of catchment sites. It is likely the supply of extra sulfate to these waterways, combined with decaying organic matter (perhaps as a result of eutrophication), results in enhanced MBO production. While sulfate-reduction has a role to play in improving water quality (through the temporary binding of metals), the environmental risk is high for locations where the monosulfides are susceptible to re-suspension from heavy rainfall and floodwaters. Blackwater events result when sediment containing monosulfides are re-suspended in the water column, causing it to deoxygenate rapidly. Blackwater events on Australia's east coast have been catastrophic, with hundreds to thousands of fish and prawns reported killed (Sammut et al. 1995; Russell & Helmke 2002; ASSAY 2008).

In this study, water samples were frequently reported above ANZECC & ARMCANZ (2000) metal guideline trigger values. It is probable that various sources of metals contributed to the metals observed in waterways and estuaries in south-western Australia, including urban, agricultural and industrial influences as well as disturbed ASS sources. Al was highly prevalent, with approximately 70% of catchment samples exceeding the ANZECC & ARMCANZ (2000) trigger values. It is not known whether Al is naturally high in south-western Australia or whether this finding represents a real environmental issue.

The following recommendations are provided as a result of this work:

| Recommendation 1: | Further investigation of hotspot sites identified in this study, including development of management and/or remediation plans for these sites. |
|-------------------|--|
| Recommendation 2: | Additional work to refine the sulfur stable isotope as an indicator of ASS influence in waterways. |
| Recommendation 3: | Further work at sites which have been identified as probably having enhanced sulfate-reduction to assess the risk associated with blackwater events at these locations. |
| Recommendation 4: | Ecotoxicology specifically targeted at Western Australian native species is required to understand whether aluminium concentrations in south-western Australian waterways are presenting a widespread ecological problem. |

Appendices

Appendix A - Full site details

A1. Site name and location for estuarine sites with sampling dates for both sampling occasions, nominally called 'summer' and 'winter'.

| Site reference | Easting | Northing | Sampling date (summer) | Sampling date (winter) |
|----------------|---------|----------|---------------------------|---------------------------|
| ARM (WRC1) | 387504 | 6458161 | 19/02/2007 | 18/06/2007 |
| BLA (WRC1) | 385081 | 6456807 | 19/02/2007 | 18/06/2007 |
| KIN (WRC1) | 401768 | 6468783 | 19/02/2007 | 18/06/2007 |
| MAY (WRC1) | 396801 | 6465716 | 19/02/2007 | 18/06/2007 |
| NAR (WRC1) | 391215 | 6463014 | 19/02/2007 | 18/06/2007 |
| NIL (WRC1) | 394660 | 6464027 | 19/02/2007 | 18/06/2007 |
| RON (WRC1) | 399802 | 6467786 | 19/02/2007 | 18/06/2007 |
| STJ (WRC1) | 397308 | 6464439 | 19/02/2007 | 18/06/2007 |
| SUC (WRC1) | 401585 | 6470349 | 19/02/2007 | 18/06/2007 |
| KEN (WRC1) | 398104 | 6456676 | 20/02/2007 | 19/06/2007 |
| BAC (WRC1) | 396637 | 6456525 | 20/02/2007 | 19/06/2007 |
| CAS (WRC1) | 397591 | 6456565 | 20/02/2007 | 19/06/2007 |
| ELL (WRC1) | 401006 | 6455131 | 20/02/2007 | 19/06/2007 |
| NIC (WRC1) | 400108 | 6455732 | 20/02/2007 | 19/06/2007 |
| RIV (WRC1) | 396239 | 6455988 | 20/02/2007 | 19/06/2007 |
| SAL (WRC1) | 393509 | 6455593 | 20/02/2007 | 19/06/2007 |
| PHE58 (WRC1) | 374388 | 6388576 | 14/03/2007 | 5/07/2007 |
| PHRM9 (WRC1) | 390581 | 6394024 | 14/03/2007 | 5/07/2007 |
| PHRM4 (WRC1) | 390106 | 6393899 | 14/03/2007 | 5/07/2007 |
| PHRM2 (WRC1) | 386385 | 6394324 | 14/03/2007 | 5/07/2007 |
| PHE04 (WRC1) | 383128 | 6395061 | 14/03/2007 | 5/07/2007 |
| PHRS7 (WRC1) | 388165 | 6406598 | | 3/07/2007 |
| PHE01 (WRC1) | 375970 | 6383055 | 14/03/2007 | 5/07/2007 |
| PHE07 (WRC1) | 379243 | 6393234 | 14/03/2007 | 5/07/2007 |
| PHRS4 (WRC1) | 384316 | 6398669 | | 3/07/2007 |
| PHRS6 (WRC1) | 385996 | 6402376 | | 3/07/2007 |
| PHE31 (WRC1) | 377709 | 6377884 | 14/03/2007 | 5/07/2007 |
| PHE02 (WRC1) | 375330 | 6391875 | 14/03/2007 | 5/07/2007 |
| MPH061 (WRC1) | 375739 | 6395307 | 6/03/2007 | 13/09/2007 |
| MPH071 (WRC1) | 379199 | 6399997 | 6/03/2007 | 13/09/2007 |
| MPH083 (WRC1) | 379748 | 6398065 | 6/03/2007 | 13/09/2007 |
| MPH006 (WRC1) | 373073 | 6388550 | 7/03/2007 | 12/09/2007 |

| MPH019 (WRC1) | 377701 | 6375206 | 7/03/2007 | 12/09/2007 |
|----------------|---------|----------|---------------------------|---------------------------|
| Site reference | Easting | Northing | Sampling date (summer) | Sampling date (winter) |
| MPH025 (WRC1) | 375151 | 6389430 | 7/03/2007 | 13/09/2007 |
| MPH015 (WRC1) | 378847 | 6379013 | 8/03/2007 | 12/09/2007 |
| MPH044 (WRC1) | 383112 | 6396277 | 8/03/2007 | 14/09/2007 |
| MPH091 (WRC1) | 384200 | 6396033 | 8/03/2007 | 13/09/2007 |
| MPH040 (WRC1) | 384543 | 6393206 | 9/03/2007 | 11/09/2007 |
| MPH155 (WRC1) | 384450 | 6394812 | 9/03/2007 | 11/09/2007 |
| MPH158 (WRC1) | 385067 | 6394326 | 9/03/2007 | 11/09/2007 |
| MPH001 (WRC1) | 385992 | 6393212 | | 8/06/2007 |
| MPH002 (WRC1) | 385517 | 6392972 | | 8/06/2007 |
| MPH003 (WRC1) | 385265 | 6392982 | | 8/06/2007 |
| 6121130 (AWRC) | 390301 | 6319885 | 15/03/2007 | 11/07/2007 |
| 6121203 (AWRC) | 387979 | 6318601 | 15/03/2007 | 11/07/2007 |
| 6121162 (AWRC) | 381937 | 6317537 | 15/03/2007 | 11/07/2007 |
| 6121161 (AWRC) | 381473 | 6316299 | 15/03/2007 | 11/07/2007 |
| 6111043 (AWRC) | 376407 | 6311508 | 15/03/2007 | 11/07/2007 |
| 6121173 (AWRC) | 378719 | 6326057 | 15/03/2007 | 11/07/2007 |
| 6121166 (AWRC) | 380821 | 6315693 | 15/03/2007 | 11/07/2007 |
| 6121168 (AWRC) | 384914 | 6315194 | 15/03/2007 | 11/07/2007 |
| 6121206 (AWRC) | 379734 | 6322355 | 15/03/2007 | 11/07/2007 |
| 6121207 (AWRC) | 376978 | 6313870 | 15/03/2007 | 11/07/2007 |
| 6101072 (AWRC) | 355173 | 6281116 | 8/03/2007 | 26/07/2007 |
| 6101067 (AWRC) | 353278 | 6279048 | 8/03/2007 | 26/07/2007 |
| 6101065 (AWRC) | 351800 | 6277800 | 8/03/2007 | 26/07/2007 |
| 610019 (AWRC) | 352702 | 6278792 | 8/03/2007 | 26/07/2007 |
| 6101064 (AWRC) | 346823 | 6275036 | 8/03/2007 | 26/07/2007 |
| 6101063 (AWRC) | 347070 | 6274433 | 8/03/2007 | 26/07/2007 |
| 610020 (AWRC) | 353845 | 6279448 | 8/03/2007 | 26/07/2007 |
| BRF02 (WRC1) | 333481 | 6218227 | | 15/08/2007 |
| HIF01 (WRC1) | 331650 | 6200534 | | 15/08/2007 |
| HIF02 (WRC1) | 331060 | 6202031 | | 15/08/2007 |
| HIF03 (WRC1) | 331083 | 6203506 | | 15/08/2007 |
| HIF04 (WRC1) | 329948 | 6203551 | | 15/08/2007 |
| HIF05 (WRC1) | 330810 | 6205366 | | 15/08/2007 |
| HIF06 (WRC1) | 332512 | 6204368 | | 15/08/2007 |
| MIF01 (WRC1) | 334394 | 6205820 | | 15/08/2007 |
| MIF02 (WRC1) | 334417 | 6206863 | | 15/08/2007 |
| MIF03 (WRC1) | 335631 | 6207254 | | 15/08/2007 |
| SRF01 (WRC1) | 336527 | 6207379 | | 15/08/2007 |

| BRF01 (WRC1) | 335239 | 6208009 | | 15/08/2007 |
|----------------|---------|----------|---------------------------|---------------------------|
| WAL003 (WRC1) | 475534 | 6128782 | 24/01/2007 | 10/07/2007 |
| Site reference | Easting | Northing | Sampling date (summer) | Sampling date (winter) |
| WAL001 (WRC1) | 474613 | 6128916 | 24/01/2007 | 10/07/2007 |
| WAL006 (WRC1) | 476299 | 6126594 | 24/01/2007 | 10/07/2007 |
| WI6 (WRC1) | 535051 | 6127398 | 20/02/2007 | 24/05/2007 |
| WI30 (WRC1) | 530840 | 6126087 | 20/02/2007 | 24/05/2007 |
| WI12 (WRC1) | 539902 | 6127235 | 20/02/2007 | 24/05/2007 |
| WAL004 (WRC1) | 475299 | 6126102 | 24/01/2007 | 10/07/2007 |
| IRW003 (WRC1) | 496447 | 6127669 | 24/01/2007 | 10/07/2007 |
| BRO002 (WRC1) | 446374 | 6138504 | 23/01/2007 | 9/07/2007 |
| IRW001 (WRC1) | 498180 | 6128240 | 24/01/2007 | 10/07/2007 |
| WAL005 (WRC1) | 473701 | 6125892 | 24/01/2007 | 10/07/2007 |
| BRO004 (WRC1) | 446568 | 6134564 | 23/01/2007 | 9/07/2007 |
| BRO001 (WRC1) | 449537 | 6134997 | 23/01/2007 | 9/07/2007 |
| AOH 5 (WRC1) | 587221 | 6131995 | 19/02/2007 | 15/05/2007 |
| AOH 4 (WRC1) | 587133 | 6130708 | 19/02/2007 | 15/05/2007 |
| AOH 2 (WRC1) | 588118 | 6128413 | 19/02/2007 | 15/05/2007 |
| APRH 3 (WRC1) | 581412 | 6120955 | 19/02/2007 | 15/05/2007 |
| APRH 4 (WRC1) | 582257 | 6119330 | 19/02/2007 | 15/05/2007 |
| APRH 2 (WRC1) | 578731 | 6122409 | 19/02/2007 | 15/05/2007 |

A2. Site name, region and location of catchment sites with sampling dates of snapshot.

| Site reference | Catchment | Easting | Northing | Sampling date |
|----------------|--------------|---------|----------|---------------|
| BBDSMP | Swan Canning | 400825 | 6476349 | 3/10/2007 |
| BCBW | Swan Canning | 398979 | 6453910 | 3/10/2007 |
| BCRUS | Swan Canning | 395344 | 6455397 | 3/10/2007 |
| BICKBK 05 | Swan Canning | 404089 | 6454565 | 4/10/2007 |
| BICKBKEDS | Swan Canning | 405926 | 6454778 | 4/10/2007 |
| BLACMDS | Swan Canning | 406298 | 6472567 | 3/10/2007 |
| BNCADEN | Swan Canning | 399913 | 6468163 | 3/10/2007 |
| BSMDO | Swan Canning | 398327 | 6468780 | 4/10/2007 |
| BWD9 | Swan Canning | 392250 | 6453870 | 4/10/2007 |
| BWMDBI | Swan Canning | 397529 | 6455220 | 5/10/2007 |
| BWMDBPI | Swan Canning | 397917 | 6468415 | 3/10/2007 |
| BWMDBR | Swan Canning | 397917 | 6468415 | 5/10/2007 |
| CAMDOUT | Swan Canning | 396734 | 6466050 | 5/10/2007 |

| EBBHDS | Swan Canning | 404034 | 6505642 | 3/10/2007 |
|----------------|------------------------------|------------------|-----------|---------------|
| EBMBUS | Swan Canning | 404034 | 6483773 | 3/10/2007 |
| HRDSMW | Swan Canning Swan Canning | 406657 419366 | 6464921 | 4/10/2007 |
| Site reference | Catchment | Easting | Northing | Sampling date |
| | outonment | Lusting | itortinig | |
| HRES | Swan Canning | 403798 | 6470091 | 4/10/2007 |
| JBDG | Swan Canning | 409267 | 6474881 | 3/10/2007 |
| JBSDS | Swan Canning | 424179 | 6472664 | 4/10/2007 |
| LACDD | Swan Canning | 400532 | 6455895 | 4/10/2007 |
| LIEP6 | Swan Canning | 399380 | 6456586 | 4/10/2007 |
| MSCWW | Swan Canning | 397920 | 6457129 | 4/10/2007 |
| PAS01 | Swan Canning | 399730 | 6466720 | 4/10/2007 |
| SBMD4 | Swan Canning | 398321 | 6463354 | 4/10/2007 |
| SCCIS10 | Swan Canning | 406665 | 6437717 | 3/10/2007 |
| SLMBAIGIN | Swan Canning | 397140 | 6466698 | 5/10/2007 |
| SRBDS | Swan Canning | 403042 | 6448942 | 3/10/2007 |
| WBHW | Swan Canning | 405079 | 6459867 | 4/10/2007 |
| WCEBDS | Swan Canning | 407821 | 6472730 | 3/10/2007 |
| YBRPUS | Swan Canning | 402151 | 6456165 | 4/10/2007 |
| 613014 (AWRC) | Peel Harvey | 392689 | 6359878 | 23/10/2007 |
| 613027 (AWRC) | Peel Harvey | 385062 | 6373803 | 23/10/2007 |
| 613029 (AWRC) | Peel Harvey | 385527 | 6386327 | 24/10/2007 |
| 613031 (AWRC) | Peel Harvey | 382559 | 6369968 | 23/10/2007 |
| 613052 (AWRC) | Peel Harvey | 381689 | 6368298 | 23/10/2007 |
| 613053 (AWRC) | Peel Harvey | 384739 | 6356848 | 23/10/2007 |
| 614030 (AWRC) | Peel Harvey | 392770 | 6421040 | 24/10/2007 |
| 614063 (AWRC) | Peel Harvey | 390024 | 6400950 | 23/10/2007 |
| 614065 (AWRC) | Peel Harvey | 394839 | 6388849 | 24/10/2007 |
| 614094 (AWRC) | Peel Harvey | 393880 | 6412350 | 24/10/2007 |
| 614120 (AWRC) | Peel Harvey | 389034 | 6405484 | 23/10/2007 |
| 614121 (AWRC) | Peel Harvey | 390409 | 6417490 | 24/10/2007 |
| AMRL3 (WRC1) | Peel Harvey | 389389 | 6409823 | 24/10/2007 |
| HR01012 (WRC1) | Peel Harvey | 389367 | 6366352 | 23/10/2007 |
| HR01014 (WRC1) | Peel Harvey | 391400 | 6369611 | 24/10/2007 |
| HR02005 (WRC1) | Peel Harvey | 387802 | 6357545 | 24/10/2007 |
| MPH003 (WRC1) | Peel Harvey | 385265 | 6392982 | 25/10/2007 |
| MR01002 (WRC1) | Peel Harvey | 390365 | 6391448 | 25/10/2007 |
| MR01010 (WRC1) | Peel Harvey | 389410 | 6393338 | 25/10/2007 |
| MR01012 (WRC1) | Peel Harvey | 389473 | 6395185 | 25/10/2007 |
| | | | | |
| MR03017 (WRC1) | Peel Harvey | 386600 | 6390933 | 25/10/2007 |

| | _ | | | |
|------------------|---------------------|---------|----------|---------------|
| MR04019 (WRC1) | Peel Harvey | 385845 | 6387694 | 24/10/2007 |
| MR05004 (WRC1) | Peel Harvey | 379099 | 6382535 | 24/10/2007 |
| MR05005 (WRC1) | Peel Harvey | 382615 | 6384645 | 25/10/2007 |
| MR05010 (WRC1) | Peel Harvey | 379573 | 6379472 | 24/10/2007 |
| Site reference | Catchment | Easting | Northing | Sampling date |
| NYUND (WRC1) | Peel Harvey | 386532 | 6395596 | 25/10/2007 |
| PATRDTRIB (WRC1) | Peel Harvey | 395230 | 6393976 | 23/10/2007 |
| PHM3 (WRC1) | Peel Harvey | 395086 | 6394140 | 25/10/2007 |
| PHS7 (WRC1) | Peel Harvey | 389833 | 6405505 | 23/10/2007 |
| SR04014 (WRC1) | Peel Harvey | 392869 | 6402343 | 24/10/2007 |
| ACID01 | Leschenault | 385534 | 6276808 | 26/09/2007 |
| ACID02 | Leschenault | 375449 | 6289328 | 25/09/2007 |
| ACID03 | Leschenault | 376559 | 6287298 | 25/09/2007 |
| ACID04 | Leschenault | 368268 | 6292218 | 26/09/2007 |
| ACID05 | Leschenault | 384039 | 6299898 | 28/09/2007 |
| ACID06 | Leschenault | 391615 | 6283971 | 26/09/2007 |
| ACID07 | Leschenault | 416039 | 6307248 | 27/09/2007 |
| ACID08 | Leschenault | 411589 | 6313848 | 27/09/2007 |
| ACID09 | Leschenault | 383789 | 6307548 | 25/09/2007 |
| ACID10 | Leschenault | 366938 | 6287998 | 25/09/2007 |
| ACID11 | Leschenault | 389393 | 6325756 | 26/09/2007 |
| ACID12 | Leschenault | 403793 | 6311435 | 27/09/2007 |
| ACID13 | Leschenault | 382420 | 6293450 | 25/09/2007 |
| ACID14 | Leschenault | 399618 | 6278171 | 26/09/2007 |
| ACID15 | Leschenault | 412898 | 6289225 | 27/09/2007 |
| ACID16 | Leschenault | 380309 | 6304016 | 25/09/2007 |
| ACID17 | Leschenault | 422143 | 6308473 | 27/09/2007 |
| BRUN6 | Leschenault | 383436 | 6319879 | 26/09/2007 |
| GBC01 | Leschenault | 367393 | 6290283 | 26/09/2007 |
| GBC02 | Leschenault | 366938 | 6286498 | 25/09/2007 |
| GBC03 | Leschenault | 379829 | 6276020 | 26/09/2007 |
| LESCH01 | Leschenault | 383488 | 6314995 | 26/09/2007 |
| LESCH02 | Leschenault | 382411 | 6310960 | 26/09/2007 |
| LESCH03 | Leschenault | 386256 | 6304512 | 26/09/2007 |
| LESCH04 | Leschenault | 388922 | 6313550 | 26/09/2007 |
| LESCH05 | Leschenault | 391345 | 6334903 | 26/09/2007 |
| LESCH06 | Leschenault | 371940 | 6302173 | 27/09/2007 |
| LESCH07 | Leschenault | 373180 | 6309674 | 28/09/2007 |
| 608003 | Scott Coastal Plain | 387489 | 6200598 | 20/11/2007 |
| 608147 | Scott Coastal Plain | 390819 | 6191198 | 20/11/2007 |
| | | | | |

| 608171 | Scott Coastal Plain | 389419 | 6187018 | 20/11/2007 |
|----------------|---------------------|---------|----------|---------------|
| 609022 | Scott Coastal Plain | 333976 | 6226544 | 21/11/2007 |
| 609041 | Scott Coastal Plain | 363450 | 6225248 | 20/11/2007 |
| 609060 | Scott Coastal Plain | 339085 | 6210670 | 21/11/2007 |
| 6081009 | Scott Coastal Plain | 406189 | 6208648 | 20/11/2007 |
| Site reference | Catchment | Easting | Northing | Sampling date |
| 6081010 | Scott Coastal Plain | 381055 | 6205878 | 20/11/2007 |
| 6081026 | Scott Coastal Plain | 385598 | 6193126 | 20/11/2007 |
| 6081040 | Scott Coastal Plain | 379758 | 6190432 | 20/11/2007 |
| 6081053 | Scott Coastal Plain | 397051 | 6194911 | 20/11/2007 |
| 6091008 | Scott Coastal Plain | 367889 | 6225989 | 20/11/2007 |
| 6091044 | Scott Coastal Plain | 328588 | 6204698 | 22/11/2007 |
| 6091222 | Scott Coastal Plain | 362853 | 6204232 | 21/11/2007 |
| 6091223 | Scott Coastal Plain | 359594 | 6205436 | 21/11/2007 |
| 6091224 | Scott Coastal Plain | 347154 | 6209255 | 21/11/2007 |
| 6091302 | Scott Coastal Plain | 330774 | 6212188 | 22/11/2007 |
| 6091305 | Scott Coastal Plain | 338370 | 6213872 | 21/11/2007 |
| 6091408 | Scott Coastal Plain | 354607 | 6229702 | 21/11/2007 |
| 6091596 | Scott Coastal Plain | 342220 | 6223054 | 21/11/2007 |
| 6091735 | Scott Coastal Plain | 382943 | 6189560 | 20/11/2007 |
| 6091752 | Scott Coastal Plain | 340749 | 6207628 | 21/11/2007 |
| 6091753 | Scott Coastal Plain | 332824 | 6228166 | 21/11/2007 |
| 6091754 | Scott Coastal Plain | 329210 | 6227737 | 21/11/2007 |
| 6091755 | Scott Coastal Plain | 352910 | 6204280 | 21/11/2007 |
| 6091756 | Scott Coastal Plain | 377028 | 6196322 | 22/11/2007 |
| 6091757 | Scott Coastal Plain | 357832 | 6209811 | 22/11/2007 |
| 6091758 | Scott Coastal Plain | 331712 | 6207481 | 22/11/2007 |
| CHE | South coast | 592084 | 6138740 | 17/10/2007 |
| DML | South coast | 528789 | 6141697 | 18/10/2007 |
| FREEBOROUGH | South coast | 546653 | 6125263 | 16/10/2007 |
| HAY | South coast | 543694 | 6136740 | 16/10/2007 |
| HENNING | South coast | 555135 | 6132230 | 16/10/2007 |
| INFLOW | South coast | 555270 | 6128945 | 16/10/2007 |
| KR | South coast | 579700 | 6134127 | 17/10/2007 |
| LEAY | South coast | 533929 | 6135647 | 18/10/2007 |
| LIT | South coast | 530128 | 6127742 | 18/10/2007 |
| LOWKG | South coast | 586272 | 6131297 | 17/10/2007 |
| MANCK | South coast | 563262 | 6125233 | 16/10/2007 |
| MIT | South coast | 535925 | 6146009 | 18/10/2007 |
| MRD | South coast | 564443 | 6127486 | 16/10/2007 |
| | | | | |

| MTBARKRD | South coast | 535289 | 6141127 | 18/10/2007 |
|----------------|-------------|---------|----------|---------------|
| RD | South coast | 577005 | 6123705 | 17/10/2007 |
| RZ7 | South coast | 550369 | 6125548 | 16/10/2007 |
| SCO | South coast | 531765 | 6134382 | 18/10/2007 |
| SLE | South coast | 545938 | 6131464 | 16/10/2007 |
| SUN | South coast | 542550 | 6132572 | 16/10/2007 |
| Site reference | Catchment | Easting | Northing | Sampling date |
| TILP001 | South coast | 558735 | 6123788 | 16/10/2007 |
| TILP006 | South coast | 570845 | 6125482 | 17/10/2007 |
| TILP008 | South coast | 574290 | 6127447 | 17/10/2007 |
| TILP012 | South coast | 567877 | 6123677 | 17/10/2007 |
| TILP013 | South coast | 565565 | 6126997 | 17/10/2007 |
| TILP047 | South coast | 561215 | 6132397 | 16/10/2007 |
| WCWF | South coast | 581660 | 6133070 | 17/10/2007 |
| WOOD1 | South coast | 566613 | 6124409 | 17/10/2007 |
| YAK1 | South coast | 566613 | 6124409 | 17/10/2007 |
| YU | South coast | 580650 | 6125797 | 17/10/2007 |

Appendix ${\rm B}-{\rm Sample}$ collection and chemical analysis details

B1. Details of the sample collection and chemical analysis methods for water samples collected from estuarine and catchment sites.

| Parameter | Lab | Method code | Volume | er details Bottle | Collection details |
|--|--|---------------------------|-----------------------|------------------------|--|
| Total nitrogen (TN) | National Measurement Institute (NMI) | WL 239 | <u>type</u> 250 mL | Plastic | Unfiltered water sample. Sample bottle filled directly from grab sample to shoulder. |
| Total phosphorus (TP) | National Measurement Institute (NMI) | WL 239 | | | |
| Filtered reactive phosphorus (FRP) | National Measurement Institute (NMI) | WL 239 | 250 mL | Plastic | Filtered water sample (0.45 µM nitrocellulose membrane). |
| Nitrate and nitrite (NO _x -N) | National Measurement Institute (NMI) | WL 239 | | | Rinse bottle three times with filtered water, and |
| Ammonium (NH₄- N) | National Measurement Institute (NMI) | WL 239 | | | fill sample bottle to shoulder. |
| Dissolved organic nitrogen (DON) | National Measurement Institute (NMI) | WL 239 | | | |
| Sulfate | National Measurement Institute (NMI) | WL 119 | | | |
| Chloride | National Measurement Institute (NMI) National | WL 119 | | | |
| Colour | Measurement Institute (NMI) | | | | |
| Total acidity | National Measurement Institute (NMI) | WL 197 | 1 L | Plastic | Unfiltered water sample. Sample bottle completely filled with unfiltered water directly from grab |
| Total alkalinity | National Measurement Institute (NMI) | WL 122 | | | sample. |
| Dissolved organic carbon (DOC) | National Measurement Institute (NMI) | WL 240 | 100 mL | Dark amber glass | Filtered water sample (0.45 μM PP syringe filter disk). |
| | | | | | Sample bottle filled to shoulder. |
| Filtered metal suite (Al, As, Ca, Cd, Co, Cr, Cu, Fe, Hg, Mg, Na, | National Measurement Institute (NMI) | WL 125 WL 272 WL 41 | 125 mL | Plastic Nalgen | Filtered water sample (0.45 µM nitrocellulose membrane) |
| Mn, Mo, Ni, Pb, Se, V, Zn) | | | | | Pre-acidified sample bottle filled to shoulder. |

| Total metal suite (Al, As, Ca, Cd, | National Measurement | WL 125 WL 272 | 125 mL | Plastic Nalgen | |
|--|-------------------------|------------------|--------|-------------------|--|
| Co, Cr, Cu, Fe, Hg, Mg, Mn, Mo, Pb, Ni, Se, V, Zn) | Institute (NMI) | WL 41 | | Naigen | Pre-acidified sample bottle filled to shoulder with unfiltered sample. |

| Hardness | National Measurement Institute (NMI) | Calculated from | n Mg + Ca ir | n metal suite | |
|---|--|---|----------------|---------------|---|
| Sulfur stable isotope (in precipitated BaSO ₄) | Monash University (Geosciences) | BaCl added to preacidified water to precipitate BaSO ₄ , which is then combusted to SO ₂ gas and measured on IRMS. | 250 mL HDPE | Plastic | Field filtered sample with a 0.45 μM nitrocellulose membrane. Filtered water added to pre-acidified (nitric acid) sample bottles and filled to shoulder. Final pH ~ 2. |

WL 239 autoanalyser method based on APHA (several methods) – APHA (1998) Standard methods for the examination of water and wastewater 20th Edition
WL 119 ion chromatography
WL 197 titration (APHA 2310B)
WL 122 titration (APHA 2320B)
WL 240 (APHA 5310)
WL 125, 272, 125 AAS and ICPMS methods
IRMS = isotope ratio mass spectroscopy

Appendix C – Quality assurance and quality control

| Analyte (units) | Swan Canning | Peel Harvey | Leschenault | Scott | South coast |
|---|--------------|-------------|-------------|---------|-------------|
| Acidity (mg L ⁻¹ CaCO ₃) | 4 | 4 | <1 | 3 | 2 |
| AI (sol) (mg L ⁻¹) | <0.005 | <0.005 | <0.005 | 0.009 | <0.005 |
| As (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Ca (sol) (mg L ⁻¹) | 0.024 | 0.016 | 0.014 | 0.014 | 0.016 |
| Cd (sol) (mg L ⁻¹) | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| CI (sol) (mg L ⁻¹) | <10 | <10 | <10 | <10 | <10 |
| Co (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Colour (TCU) | <1 | 11 | 1 | <1 | <1 |
| Cr (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Cu (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Fe (sol) (mg L ⁻¹) | <0.005 | <0.005 | <0.005 | <0.005 | <0.005 |
| Hg (sol) (mg L⁻¹) | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| Mg (sol) (mg L⁻¹) | 0.025 | 0.009 | 0.008 | <0.005 | <0.005 |
| VIn (sol) (mg L⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Mo (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| DON (mg L ⁻¹) | 0.054 | 0.031 | 0.07 | <0.025 | <0.025 |
| NOx (mg L ⁻¹) | 0.019 | <0.01 | 0.028 | <0.01 | <0.01 |
| TN (mg L ⁻¹) | 0.078 | 0.049 | 0.11 | <0.025 | <0.025 |
| $NH_3 (mg L^{-1})$ | <0.01 | 0.014 | <0.01 | <0.01 | 0.011 |
| Na (sol) (mg L ⁻¹) | <0.05 | <0.05 | <0.05 | 0.088 | <0.05 |
| Ni (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |

C1. Highest recorded measurement for field blank (procedural blank) samples collected within each sampling catchment.

| TP (mg L ⁻¹) | <0.005 | <0.005 | 0.006 | <0.005 | <0.005 |
|---|--------|--------|--------|--------|--------|
| FRP (mg L^{-1}) | <0.005 | <0.005 | 0.005 | <0.005 | <0.005 |
| Pb (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| SO ₄ ²⁻ (mg L ⁻¹) | <5 | <5 | <5 | <5 | <5 |
| Se (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| V (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Zn (sol) (mg L ⁻¹) | 0.002 | 0.004 | 0.001 | 0.023 | 0.002 |

C2. Greatest difference between replicate measurements for field replicate samples within each catchment.

| Analyte (units) | Swan Canning | Peel Harvey | Leschenault | Scott | South coast |
|--|--------------|-------------|-------------|-------|-------------|
| Acidity (mg L ⁻¹ CaCO ₃) | 1 | 1 | - | - | - |
| AI (sol) (mg L ⁻¹) | 0.006 | 0.07 | 0.01 | 0.003 | 0.04 |
| AI (tot) (mg L ⁻¹) | 0.075 | 0.07 | 0.12 | 0.01 | 0.03 |
| Alkalinity (mg L ⁻¹ CaCO ₃) | 5 | 23 | 1 | - | - |
| As (sol) (mg L⁻¹) | - | - | - | - | - |
| As (tot) (mg L ⁻¹) | - | - | - | - | - |
| DOC (mg L ⁻¹) | 1 | 11 | 1 | 1 | 2 |
| Ca (sol) (mg L ⁻¹) | 1 | 2 | - | - | 0.3 |
| Ca (tot) (mg L ⁻¹) | 1 | 2 | - | - | 0.3 |
| Cd (sol) (mg L ⁻¹) | - | - | - | - | - |
| Cd (tot) (mg L ⁻¹) | - | - | - | - | - |
| CI (sol) (mg L ⁻¹) | - | 50 | - | 10 | - |
| Co (sol) (mg L ⁻¹) | - | - | - | - | - |
| Co (tot) (mg L ⁻¹) | - | - | - | - | - |

| Colour (TCU) | 10 | 3 | - | 2 | 20 |
|--------------------------------|-------|-------|-------|-------|-------|
| Cr (sol) (mg L ⁻¹) | - | - | - | - | - |
| Cr (tot) (mg L^{-1}) | - | - | - | 0.001 | - |
| Cu (sol) (mg L^{-1}) | - | - | - | - | - |
| Cu (tot) (mg L^{-1}) | - | - | - | - | - |
| δ^{34} S (‰) | 1.9 | 0.6 | 3.1 | 3.1 | 1.3 |
| Fe (sol) (mg L ⁻¹) | 0.02 | 0.05 | - | 0.03 | 0.1 |
| Fe (tot) (mg L^{-1}) | 0.2 | 0.06 | 0.2 | 0.01 | 0.1 |
| Hg (sol) (mg L^{-1}) | - | - | - | - | - |
| Hg (tot) (mg L^{-1}) | - | - | - | - | - |
| Mg (sol) (mg L^{-1}) | 1 | 0.8 | 0.1 | - | 0.4 |
| Mg (tot) (mg L^{-1}) | 0.1 | 3 | 0.1 | - | 0.1 |
| Mn (sol) (mg L ⁻¹) | 0.004 | 0.003 | 0.001 | 0.003 | 0.001 |
| Mn (tot) (mg L ⁻¹) | 0.002 | - | - | 0.001 | 0.013 |
| Mo (sol) (mg L ⁻¹) | - | - | - | - | - |
| Mo (tot) (mg L ⁻¹) | - | 0.001 | - | - | - |
| DON (mg L ⁻¹) | 0.03 | 0.08 | 0.2 | 0.022 | 0.03 |
| NOx (mg L ⁻¹) | 0.02 | 0.074 | 0.01 | 0.013 | 0.05 |
| TN (mg L ⁻¹) | 0.1 | 0.08 | 0.1 | 0.024 | 0.03 |
| $NH_3 (mg L^{-1})$ | 0.005 | 0.017 | 0.02 | 0.011 | 0.003 |
| Na (sol) (mg L ⁻¹) | 10 | 6 | - | - | 2 |
| Ni (sol) (mg L ⁻¹) | - | - | - | 0.001 | - |
| Ni (tot) (mg L ⁻¹) | - | - | - | - | - |
| TP (mg L ⁻¹) | 0.005 | 0.015 | - | - | 0.009 |
| FRP (mg L ⁻¹) | - | 0.027 | - | - | 0.006 |
| | | | | | |

| Pb (sol) (mg L ⁻¹) | - | - | - | - | - |
|---|-------|-------|-------|-------|-------|
| Pb (sol) (mg L ⁻¹) | - | - | - | - | - |
| SO ₄ ²⁻ (mg L ⁻¹) | 1 | 11 | 2 | 2 | 1 |
| Se (sol) (mg L ⁻¹) | - | - | - | - | - |
| Se (tot) (mg L ⁻¹) | - | - | - | - | - |
| V (sol) (mg L ⁻¹) | - | - | - | - | - |
| V (tot) (mg L ⁻¹) | - | - | 0.001 | - | - |
| Zn (sol) (mg L ⁻¹) | 0.001 | 0.002 | 0.002 | 0.001 | 0.001 |
| Zn (tot) (mg L ⁻¹) | 0.002 | 0.003 | 0.001 | - | - |

C3. Highest recorded measurement for field blank (procedural blank) samples collected within each estuary.

| Analyte (units) | Peel Harvey | Leschenault | Vasse | Hardy | South coast |
|---------------------------------------|-------------|-------------|---------|---------|-------------|
| Acidity (mg L⁻¹ CaCO₃) | 4 | - | 5 | - | - |
| AI (SOI) (mg L ⁻¹) | 0.034 | <0.005 | 0.012 | <0.005 | <0.005 |
| As (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Ca (sol) (mg L ⁻¹) | 0.4 | 0.06 | 0.12 | 0.015 | 0.008 |
| Cd (sol) (mg L ⁻¹) | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| CI (sol) (mg L ⁻¹) | 20 | <10 | <10 | <10 | - |
| Co (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Cr (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Cu (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Fe (sol) (mg L ⁻¹) | <0.05 | <0.05 | <0.05 | <0.05 | <0.05 |
| Hg (sol) (mg L ⁻¹) | <0.0001 | <0.0001 | <0.0001 | <0.0001 | <0.0001 |
| Mg (sol) (mg L ⁻¹) | 1.1 | 0.038 | 0.27 | 0.019 | 0.009 |

| Accessing the influence of said sulfate sails on water, | envelite in acuth wastern Australian actabusents and actuaries |
|--|--|
| - Assessing the inilience of acid suilate solls on water (| quality in south-western Australian catchments and estuaries |
| | |

| Mn (sol) (mg L⁻¹) | <0.005 | <0.005 | <0.005 | <0.005 | 0.061 |
|---------------------------------------|--------|--------|--------|--------|--------|
| Mo (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| DON (mg L^{-1}) | 0.037 | <0.025 | <0.025 | <0.025 | <0.025 |
| NOx (mg L^{-1}) | 0.01 | <0.01 | 0.01 | <0.01 | <0.01 |
| TN (mg L ⁻¹) | 0.067 | <0.025 | 0.063 | 0.085 | 0.025 |
| $NH_3 (mg L^{-1})$ | 0.013 | <0.01 | <0.01 | <0.01 | <0.01 |
| Na (sol) (mg L ⁻¹) | 8.7 | 0.17 | 1.9 | 0.062 | 0.061 |
| Ni (SOI) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| TP (mg L ⁻¹) | 0.008 | 0.005 | <0.05 | 0.01 | <0.005 |
| FRP (mg L ⁻¹) | <0.005 | 0.005 | <0.005 | 0.009 | <0.005 |
| Pb (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| SO_4^{2-} (mg L ⁻¹) | <5 | <5 | <5 | <5 | <5 |
| Se (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| V (SOI) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| Zn (sol) (mg L ⁻¹) | <0.001 | <0.001 | <0.001 | 0.007 | <0.001 |

C4. Greatest difference between replicate measurements for field replicate samples within each estuary.

| Analyte (units) | Swan Canning | Peel Harvey | Leschenault | Vasse | Hardy | South coast |
|--|--------------|-------------|-------------|-------|-------|-------------|
| Acidity (mg L ⁻¹ CaCO ₃) | | 1 | - | 2 | 1 | - |
| AI (sol) (mg L ⁻¹) | | 0.005 | 0.011 | 0.007 | 0.003 | 0.001 |
| AI (tot) (mg L ⁻¹) | | 0.029 | 0.025 | 0.025 | 0.01 | 0.018 |
| Alkalinity (mg L ⁻¹ CaCO ₃) | 30 | 5 | 10 | 10 | 2 | 10 |
| As (sol) (mg L ⁻¹) | | - | 0.001 | - | - | - |
| As (tot) (mg L ⁻¹) | | 0.003 | - | - | - | - |

| | | 0 | 0 | | | |
|--------------------------------|------|-------|-------|-------|-------|-------|
| DOC (mg L ⁻¹) | 1 | 2 | 2 | 4 | 2 | - |
| Ca (sol) (mg L ⁻¹) | | 10 | - | 10 | 2 | 10 |
| Ca (tot) (mg L ⁻¹) | | 10 | 20 | - | 2 | - |
| Cd (sol) (mg L ⁻¹) | | - | - | - | - | - |
| Cd (tot) (mg L ⁻¹) | | - | - | - | - | - |
| CI (sol) (mg L ⁻¹) | | - | 300 | 200 | 30 | |
| Co (sol) (mg L ⁻¹) | | - | - | - | - | - |
| Co (tot) (mg L ⁻¹) | | - | - | - | - | - |
| Cr (sol) (mg L ⁻¹) | | - | - | - | - | - |
| Cr (tot) (mg L ⁻¹) | | - | - | - | - | - |
| Cu (sol) (mg L ⁻¹) | | - | 0.002 | - | 0.005 | - |
| Cu (tot) (mg L ⁻¹) | | - | 0.001 | - | - | - |
| δ ³⁴ S (‰) | | 2.1 | 1.1 | 0.2 | 0.4 | 0.5 |
| Fe (sol) (mg L ⁻¹) | | 0.08 | 0.09 | - | 0.01 | - |
| Fe (tot) (mg L ⁻¹) | | 0.09 | 0.03 | - | 0.09 | - |
| Hg (sol) (mg L ⁻¹) | | - | - | - | - | - |
| Hg (tot) (mg L ⁻¹) | | - | - | - | - | - |
| Mg (sol) (mg L ⁻¹) | | 50 | 20 | 10 | 8 | 10 |
| Mg (tot) (mg L ⁻¹) | | 50 | 40 | - | 9 | 10 |
| Mn (sol) (mg L ⁻¹) | | 0.001 | 0.05 | 0.009 | - | - |
| Mn (tot) (mg L ⁻¹) | | 0.001 | 0.05 | - | 0.004 | - |
| Mo (sol) (mg L ⁻¹) | | 0.001 | - | - | - | - |
| Mo (tot) (mg L ⁻¹) | | 0.001 | - | - | - | 0.001 |
| DON (mg L ⁻¹) | 0.06 | 0.1 | 0.1 | 0.1 | 0.06 | 0.01 |
| NOx (mg L^{-1}) | - | 0.02 | 0.001 | 0.019 | 0.003 | - |
| | | | | | | |

| TN (mg L ⁻¹) | 0.1 | 0.1 | 0.6 | - | 0.06 |
|---|-------|-------|------|-------|-------|
| $NH_3 (mg L^{-1})$ | 0.006 | 0.05 | 0.09 | 0.001 | - |
| Na (sol) (mg L ⁻¹) | 330 | 190 | 180 | 40 | 200 |
| Ni (sol) (mg L ⁻¹) | - | - | - | - | - |
| Ni (tot) (mg L ⁻¹) | - | 0.001 | - | - | 0.01 |
| TP (mg L ⁻¹) | 0.002 | - | 0.12 | 0.01 | 0.006 |
| FRP (mg L ⁻¹) | 0.003 | 0.03 | - | 0.003 | 0.003 |
| Pb (sol) (mg L ⁻¹) | - | - | - | - | - |
| Pb (sol) (mg L ⁻¹) | - | - | - | - | - |
| SO ₄ ²⁻ (mg L ⁻¹) | 100 | 100 | - | 3 | |
| Se (sol) (mg L ⁻¹) | - | - | - | - | - |
| Se (tot) (mg L ⁻¹) | - | - | - | - | - |
| V (sol) (mg L ⁻¹) | - | - | - | - | - |
| V (tot) (mg L ⁻¹) | - | 0.001 | - | - | - |
| Zn (sol) (mg L ⁻¹) | 0.001 | - | - | - | - |
| Zn (tot) (mg L ⁻¹) | 0.009 | 0.005 | - | 0.008 | 0.002 |

Appendix D - Guidelines trigger values per estuary

D1. Comparison of Swan-Canning estuary water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection high-reliability TV (metals, metalloids) unless otherwise stated. N = 32 (16 for total metals). Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|---|
| Aluminium (total and soluble) (mg L ⁻¹) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 50% total; 0% soluble. |
| Arsenic (total and soluble) (mg L ⁻¹) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L ⁻¹) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Guideline not exceeded. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Guideline not exceeded. |
| Copper (total and soluble) (mg L ⁻¹) | ≥0.0013 ^M | Samples above guideline: total and soluble Cu 3%. |
| Iron (total and soluble) (mg L ⁻¹) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 13% total; 3% soluble. |
| Mercury (total and soluble) (mg L^{-1}) | ≥0.0004 ^M | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L^{-1}) | ≥0.011 | Guideline not exceeded. |
| Lead (total and soluble) (mg L^{-1}) | ≥0.0034 | Guideline not exceeded. |
| Selenium (total and soluble) (mg L^{-1}) | ≥0.0011 | Guideline not exceeded. |
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Guideline not exceeded. |

| Zinc (total and soluble) | ≥0.008 | Guideline not exceeded. |
|--------------------------|--------|-------------------------|
| (mg L ⁻¹) | | |

^A Low reliability ANZECC & ARMCANZ guideline value.
 ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)
 ^M Marine ANZECC & ARMCANZ guideline value.
 ^s indicates the guideline triggered mainly in summer.
 ^w indicates the guideline triggered mainly in winter.

Comparison of Peel-Harvey estuary water quality with ANZECC & ARMCANZ D2. (2000) trigger values for freshwater 95% species protection high-reliability TV (metals, metalloids) unless otherwise stated. N = 51 (27 for total metals). Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|--|
| Aluminium (total and soluble) (mg L^{-1}) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 93% total; 14% soluble. 33% are 4x trigger value. |
| Arsenic (total and soluble) (mg L ⁻¹) | ≥0.013 | Samples above guideline: 30% total; 0% soluble. |
| Cadmium (total and soluble) (mg L ⁻¹) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Samples above guideline: 7% total; 3% soluble. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Guideline not exceeded. |
| Copper (total and soluble) (mg L^{-1}) | ≥0.0013 ^M | Guideline not exceeded. |
| Iron (total and soluble) (mg L^{-1}) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 4% total; 2% soluble. |
| Mercury (total and soluble) (mg L^{-1}) | ≥0.0004 ^M | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L^{-1}) | ≥0.011 | Guideline not exceeded. |
| Lead (total and soluble) (mg L ⁻¹) | ≥0.0034 | Guideline not exceeded. |

| Selenium (total and soluble) (mg L ⁻¹) | ≥0.0011 | Guideline not exceeded. |
|---|---------|-------------------------|
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Guideline not exceeded. |
| Zinc (total and soluble) (mg L ⁻¹) | ≥0.008 | Guideline not exceeded. |

^A Low reliability ANZECC & ARMCANZ guideline value. ^B Ambient water quality guidelines for iron, Province of British Columbia (2008) ^M Marine ANZECC & ARMCANZ guideline value.

^s indicates guideline triggered mainly in summer.

D3. Comparison of Leschenault Estuary water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection high-reliability TV (metals, metalloids) unless otherwise stated. N = 20 (10 for total metals). Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|---|
| Aluminium (total and soluble) (mg L^{-1}) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 83% total; 35% soluble. |
| Arsenic (total and soluble) (mg L^{-1}) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Samples above guideline: 10% total; 5% soluble |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Samples above guideline: 20% total; 10% soluble. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Guideline not exceeded. |
| Copper (total and soluble) (mg L^{-1}) | ≥0.0013 ^M | Samples above guideline: 20% total ^W ; 0% soluble. |
| Iron (total and soluble) (mg L^{-1}) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 50% total; 25% soluble. |
| Mercury (total and soluble) (mg L^{-1}) | ≥0.0004 ^M | Guideline not exceeded. |
| Manganese (total and soluble) (mg L⁻¹) | ≥1.900 | Guideline not exceeded. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |

| Nickel (total and soluble) (mg L^{-1}) | ≥0.011 | Guideline not exceeded. |
|---|---------|---|
| Lead (total and soluble) (mg L^{-1}) | ≥0.0034 | Guideline not exceeded. |
| Selenium (total and soluble) (mg L ⁻¹) | ≥0.0011 | Guideline not exceeded. |
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Samples above guideline: 10% total; 5% soluble. |
| Zinc (total and soluble) (mg L ⁻¹) | ≥0.008 | Guideline not exceeded. |

^A Low reliability ANZECC & ARMCANZ guideline value.
 ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)
 ^M Marine ANZECC & ARMCANZ guideline value.
 ^W indicates the guideline triggered mainly in winter.

Comparison of Vasse-Wonnerup estuary water quality with ANZECC & ARMCANZ D4. (2000) trigger values for freshwater 95% species protection high-reliability TV (metals, metalloids) unless otherwise stated. N = 14 (7 for total metals). Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|--|
| Aluminium (total and soluble) (mg L^{-1}) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 57% total; 14% soluble ^w . |
| Arsenic (total and soluble) (mg L ⁻¹) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L ⁻¹) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Guideline not exceeded. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Guideline not exceeded. |
| Copper (total and soluble) (mg L ⁻¹) | ≥0.0013 ^M | Guideline not exceeded. |
| Iron (total and soluble) (mg L⁻¹) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 43% total; 14% soluble. |
| Mercury (total and soluble) (mg L ⁻¹) | ≥0.0004 ^M | Guideline not exceeded. |

| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. |
|---|---------|---|
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L ⁻¹) | ≥0.011 | Guideline not exceeded. |
| Lead (total and soluble) (mg L ⁻¹) | ≥0.0034 | Guideline not exceeded. |
| Selenium (total and soluble) (mg L^{-1}) | ≥0.0011 | Guideline not exceeded. |
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Guideline not exceeded. |
| Zinc (total and soluble) (mg L ⁻¹) | ≥0.008 | Samples above guideline: 14% total; 0% soluble. |

^A Low reliability ANZECC & ARMCANZ guideline value.
 ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)
 ^M Marine ANZECC & ARMCANZ guideline value.
 ^W indicates the guideline triggered mainly in winter.

Comparison of Hardy Inlet water quality with ANZECC & ARMCANZ (2000) trigger D5. values for freshwater 95% species protection high-reliability TV (metals, metalloids) unless otherwise stated. N = 12 (only sampled once). Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|--|
| Aluminium (total and soluble) (mg L^{-1}) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 75% total; 25% soluble. |
| Arsenic (total and soluble) (mg L ⁻¹) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Guideline not exceeded. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Guideline not exceeded. |
| Copper (total) (mg L ⁻¹) | ≥0.0013 ^M | Samples above guideline: 8% total; 0% soluble. |

| Iron (total and soluble) (mg $L^{-1})^{B}$ | ≥1.000 (total) ≥0.35 (soluble) | Guideline not exceeded. | |
|---|-----------------------------------|-------------------------|--|
| Mercury (total and soluble) (mg L ⁻¹) | ≥0.0004 ^M | Guideline not exceeded. | |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. | |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. | |
| Nickel (total and soluble) (mg L ⁻¹) | ≥0.011 | Guideline not exceeded. | |
| Lead (total and soluble) (mg L^{-1}) | ≥0.0034 | Guideline not exceeded. | |
| Selenium (total and soluble) (mg L^{-1}) | ≥0.0011 | Guideline not exceeded. | |
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Guideline not exceeded. | |
| Zinc (total and soluble) (mg L^{-1}) | ≥0.008 | Guideline not exceeded. | |
| A Low reliability ANZECC & ARMCANZ guideling value | | | |

^A Low reliability ANZECC & ARMCANZ guideline value.
 ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)
 ^M Marine ANZECC & ARMCANZ guideline value.

D6. Comparison of south coast estuaries' water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection high-reliability TV (metals, metalloids) unless otherwise stated. N = 12 (only sampled once). Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|--|
| Aluminium (total and soluble) (mg L^{-1}) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 47% total; 26% soluble. |
| Arsenic (total and soluble) (mg L^{-1}) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Guideline not exceeded. |

| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Guideline not exceeded. |
|--|-----------------------------------|--|
| Copper (total and soluble) (mg L ⁻¹) | ≥0.0013 ^M | Guideline not exceeded. |
| Iron (total and soluble) (mg $L^{-1})^{B}$ | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 0% total; 3% soluble. |
| Mercury (total and soluble) (mg L ⁻¹) | ≥0.0004 ^M | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L ⁻¹) | ≥0.011 | Guideline not exceeded. |
| Lead (total and soluble) (mg L^{-1}) | ≥0.0034 | Guideline not exceeded. |
| Selenium (total and soluble) (mg L^{-1}) | ≥0.0011 | Guideline not exceeded. |
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Guideline not exceeded. |
| Zinc (total and soluble) (mg L^{-1}) | ≥0.008 | Guideline not exceeded. |

^A Low reliability ANZECC & ARMCANZ guideline value.
 ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)
 ^M Marine ANZECC & ARMCANZ guideline value.

Appendix E – Guideline trigger values per catchment

E1. Comparison of Swan-Canning catchment water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection high-reliability TV, unless otherwise stated. N = 30. Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|--|
| Aluminium (total and soluble) (mg L ⁻¹) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 77% total; 45% soluble. |
| Arsenic (total and soluble) (mg L ⁻¹) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Samples above guideline: 7% total; 7% soluble. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Samples above guideline: 7% total; 0% soluble. |
| Copper (total and soluble) (mg L^{-1}) | ≥0.0014 | Samples above guideline: 3% total; 3% soluble. |
| Iron (total and soluble) (mg L ⁻¹) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 33% total; 14% soluble. |
| Mercury (total and soluble) (mg L^{-1}) | ≥0.0006 | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L ⁻¹) | ≥0.011 | Samples above guideline: 3% total; 3% soluble. |
| Lead (total and soluble) (mg L ⁻¹) | ≥0.0034 | Guideline not exceeded. |
| Selenium (total and soluble) (mg L^{-1}) | ≥0.0011 | Guideline not exceeded. |
| Vanadium (total and soluble) (mg L^{-1}) | ≥0.006 | Guideline not exceeded. |
| Zinc (total and soluble) | ≥0.008 | Samples above guideline: 7% total; 7% |

| $(mg L^{-1})$ | soluble. |
|---------------|----------|
|---------------|----------|

^A Low reliability ANZECC & ARMCANZ guideline value. ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)

E2. Comparison of Peel-Harvey catchment water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection high-reliability TV, unless otherwise stated. N = 31. Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|---|
| Aluminium (total and soluble) (mg L ⁻¹) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 100% total; 71% soluble. |
| Arsenic (total and soluble) (mg L^{-1}) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Samples above guideline: 13% total; 10% soluble. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Samples above guideline: 13% total; 0% soluble. |
| Copper (total and soluble) (mg L ⁻¹) | ≥0.0014 | Samples above guideline: 6% total; 3% soluble. |
| Iron (total and soluble) (mg L^{-1}) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 65% total; 39% soluble. |
| Mercury (total and soluble) (mg L^{-1}) | ≥0.0006 | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Samples above guideline: 3% total; 0% soluble. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L^{-1}) | ≥0.011 | Guideline not exceeded. |
| Lead (total and soluble) (mg L^{-1}) | ≥0.0034 | Guideline not exceeded. |
| Selenium (total and soluble) (mg L^{-1}) | ≥0.0011 | Guideline not exceeded. |

| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Samples above guideline: 16% total; 13% soluble. |
|---|--------|--|
| Zinc (total and soluble) (mg L ⁻¹) | ≥0.008 | Samples above guideline: 6% total; 3% soluble. |

^A Low reliability ANZECC & ARMCANZ guideline value. ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)

E3. Comparison of Leschenault catchment water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection high-reliability TV, unless otherwise stated. N = 28. Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|---|
| Aluminium (total and soluble) (mg L ⁻¹) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 100% total; 89% soluble. |
| Arsenic (total and soluble) (mg L^{-1}) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Samples above guideline: 38% total; 18% soluble. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Samples above guideline: 4% total; 0% soluble. |
| Copper (total and soluble) (mg L^{-1}) | ≥0.0014 | Samples above guideline: 21% total; 14% soluble. |
| Iron (total and soluble) (mg L^{-1}) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 68% total; 21% soluble. |
| Mercury (total and soluble) (mg L^{-1}) | ≥0.0006 | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L^{-1}) | ≥0.011 | Guideline not exceeded. |
| Lead (total and soluble) (mg L ⁻¹) | ≥0.0034 | Guideline not exceeded. |

| Selenium (total and soluble) (mg L ⁻¹) | ≥0.0011 | Guideline not exceeded. |
|---|---------|--|
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Samples above guideline: 7% total; 0% soluble. |
| Zinc (total and soluble) (mg L^{-1}) | ≥0.008 | Guideline not exceeded. |

^A Low reliability ANZECC & ARMCANZ guideline value.
 ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)

E4. Comparison of Scott Coastal Plain catchment water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection highreliability TV, unless otherwise stated. N = 28. Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|--|
| Aluminium (total and soluble) (mg L^{-1}) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 86% total; 54% soluble. |
| Arsenic (total and soluble) (mg L ⁻¹) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Samples above guideline: 25% total; 25% soluble. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Samples above guideline: 11% total; 4% soluble. |
| Copper (total and soluble) (mg L ⁻¹) | ≥0.0014 | Samples above guideline: 4% total; 4% soluble. |
| Iron (total and soluble) (mg L ⁻¹) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 46% total; 29% soluble. |
| Mercury (total and soluble) (mg L ⁻¹) | ≥0.0006 | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Samples above guideline: 4% total; 4% soluble. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |
| Nickel (total and soluble) (mg L ⁻¹) | ≥0.011 | Guideline not exceeded. |

| Lead (total and soluble) (mg L ⁻¹) | ≥0.0034 | Guideline not exceeded. |
|---|---------|-------------------------|
| Selenium (total and soluble) (mg L ⁻¹) | ≥0.0011 | Guideline not exceeded. |
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Guideline not exceeded. |
| Zinc (total and soluble) (mg L^{-1}) | ≥0.008 | Guideline not exceeded. |

^A Low reliability ANZECC & ARMCANZ guideline value.
 ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)

E5. Comparison of south coast catchment water quality with ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection high-reliability TV, unless otherwise stated. N = 29. Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc.

| Variable (units) | Trigger level | Samples exceeding trigger values |
|--|---|--|
| Aluminium (total and soluble) (mg L^{-1}) | ≥0.055 (pH>6.5) ≥0.008 (pH<6.5) ^A | Samples above guideline: 97% total; 93% soluble. |
| Arsenic (total and soluble) (mg L ⁻¹) | ≥0.013 | Guideline not exceeded. |
| Cadmium (total and soluble) (mg L^{-1}) | ≥0.0002 | Guideline not exceeded. |
| Cobalt (total and soluble) (mg L^{-1}) ^A | ≥0.0014 | Samples above guideline: 14% total; 14% soluble. |
| Chromium (using Cr (VI) trigger) (total and soluble) (mg L ⁻¹) | ≥0.001 | Samples above guideline: 17% total; 10% soluble. |
| Copper (total and soluble) (mg L ⁻¹) | ≥0.0014 | Samples above guideline: 3% total; 0% soluble. |
| Iron (total and soluble) (mg L⁻¹) ^B | ≥1.000 (total) ≥0.35 (soluble) | Samples above guideline: 83% total; 72% soluble. |
| Mercury (total and soluble) (mg L ⁻¹) | ≥0.0006 | Guideline not exceeded. |
| Manganese (total and soluble) (mg L ⁻¹) | ≥1.900 | Guideline not exceeded. |
| Molybdenum (total and soluble) (mg L ⁻¹) ^A | ≥0.034 | Guideline not exceeded. |

| Nickel (total and soluble) (mg L ⁻¹) | ≥0.011 | Guideline not exceeded. |
|---|---------|--|
| Lead (total and soluble) (mg L ⁻¹) | ≥0.0034 | Guideline not exceeded. |
| Selenium (total and soluble) (mg L ⁻¹) | ≥0.0011 | Guideline not exceeded. |
| Vanadium (total and soluble) (mg L ⁻¹) | ≥0.006 | Samples above guideline: 3% total; 0% soluble. |
| Zinc (total and soluble) (mg L^{-1}) | ≥0.008 | Samples above guideline: 3% total; 3% soluble. |

^A Low reliability ANZECC & ARMCANZ guideline value. ^B Ambient water quality guidelines for iron, Province of British Columbia (2008)

Appendix F - Guideline trigger values exceeded by site

F1. Catchment sites at which metals and nutrients exceeded ANZECC & ARMCANZ guideline trigger values. The ANZECC & ARMCANZ (2000) trigger values for freshwater 95% species protection and lowland river ecosystem stress are used. Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc. Normal text indicates value exceeded trigger value, **bold** text indicates value was 2x trigger value, and **red** text indicates 10x trigger value.

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|---------------|-----------------|---------------------------------|-----------------------------|-------------------------------------|
| BBDSMP | 5 | Swan Canning | Al, Fe | <mark>Al</mark> , Fe | TN |
| BCBW | 5 | Swan Canning | AI | AI, Cr | NOx , TN, TP , FRP |
| BCRUS | 5 | Swan Canning | AI | AI, Fe | - |
| BICKBK 05 | 5 | Swan Canning | AI | AI | NOx |
| BICKBKEDS | 5 | Swan Canning | - | AI | NOx |
| BLACMDS | 4a | Swan Canning | - | Fe | NOx |
| BNCADEN | 5 | Swan Canning | AI | AI | NOx , TN, TP , FRP |
| BSMDO | 5 | Swan Canning | AI | AI | NOx, TN |
| BWD9 | 5 | Swan Canning | AI, Co, <mark>Zn</mark> | Al, Co, Fe, <mark>Zn</mark> | NOx, TN, TP, FRP |
| BWMDBI | 5 | Swan Canning | n.d. | AI | NOx |
| BWMDBPI | 5 | Swan Canning | Al, Co, Fe | Al, Co, Cr, Fe | NOx, NH4 ⁺ |
| BWMDBR | 5 | Swan Canning | AI | AI | NOx |
| CAMDOUT | 5 | Swan Canning | - | Al, Fe | NOx |

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|---------------|-----------------|---------------------------------|----------------------------|--|
| EBBHDS | 2 | Swan Canning | AI, | Al, Fe | TN, TP, FRP |
| EBMBUS | 2 | Swan Canning | AI, Fe | AI, Fe | TN, TP, FRP |
| HRDSMW | 2 | Swan Canning | - | AI | - |
| HRES | 5 | Swan Canning | - | Al, Fe | NOx, NH4 ⁺ |
| JBDG | 4a | Swan Canning | - | - | NOx |
| JBSDS | 2 | Swan Canning | - | AI | - |
| LACDD | 5 | Swan Canning | - | - | NOx, TN |
| LIEP6 | 5 | Swan Canning | - | - | NOx, TN, TP, FRP |
| MSCWW | 5 | Swan Canning | - | AI | NOx , TN, NH4 , TP , FRP |
| PAS01 | 5 | Swan Canning | AI , Cu, Zn | AI, Cu, Zn | NOx, TN |
| SBMD4 | 5 | Swan Canning | - | AI | TP, FRP |
| SCCIS10 | 5 | Swan Canning | AI, | AI, Fe | NOx, TN, TP, FRP |
| SLMBAIGIN | 5 | Swan Canning | - | AI | NOx, TP, FRP |
| SRBDS | 3 | Swan Canning | - | AI | NOx |
| WBHW | 5 | Swan Canning | - | - | NOx, TN |
| WCEBDS | 4a | Swan Canning | - | - | - |
| YBRPUS | 5 | Swan Canning | - | - | NOx |

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-------------------|---------------|-------------|---------------------------------|-------------------------------|--|
| 613014 | 4a | Peel Harvey | AI | AI | - |
| 613027 | 2 | Peel Harvey | AI, Fe | AI, Cr, Fe | TN, TP , FRP |
| 613029 | 2 | Peel Harvey | - | AI | TN |
| 613031 | 2 | Peel Harvey | - | AI, Fe | ТР |
| 613052 | 5 | Peel Harvey | AI | AI, Fe | TP |
| 613053 | 5 | Peel Harvey | AI | AI, Cr | TN, NH₄ ⁺ , TP , FRP |
| 614030 | 5 | Peel Harvey | - | AI, Fe | TP, FRP |
| 614063 | 4a | Peel Harvey | AI, Fe | AI, Fe | TN, TP, FRP |
| 614065 | 2 | Peel Harvey | - | AI | - |
| 614094 | 5 | Peel Harvey | AI | AI, Fe, Cr | - |
| 614120 | n.d. | Peel Harvey | <mark>Al</mark> , Fe | AI, Cu, Fe | TN, NH₄ ⁺ , TP , FRP |
| 614121 | 5 | Peel Harvey | - | Al, Fe, Zn | TN, TP, FRP |
| AMRL3 | 2 | Peel Harvey | Al, Fe | AI, Fe | TN, TP , FRP |
| HR01012 (WRC1) | 4a | Peel Harvey | AI | AI | NOx |
| HR01014 | 2 | Peel Harvey | Al, Fe | AI, Fe | ТР |
| HR02005 | 4a | Peel Harvey | AI, Cu | AI, Cr | TN, TP , FRP |
| MPH003 | 3 | Peel Harvey | Al, Co, Zn | Al, Co | - |
| MR01002 | 2 | Peel Harvey | AI | AI | TN |
| MR01010 | 2 | Peel Harvey | Al, Fe | AI , Co, Fe , V | TN, TP , FRP |
| MR01012 | 1 | Peel Harvey | ΑΙ | AI, Fe | TN, TP , FRP |
| MR03017 | 4b | Peel Harvey | - | ΑΙ | TN |
| MR04014 | 2 | Peel Harvey | AI, Co | AI , Co, Fe | TN, NH4, TP |
| MR04019 | 2 | Peel Harvey | V | AI | TN, TP , FRP |
| MR05004 | 2 | Peel Harvey | AI, Fe | AI, Fe | TN, TP , FRP |

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|---------------|-------------|---------------------------------|--|--|
| MR05005 | n.d. | Peel Harvey | AI, Fe, V | AI, Fe, V | TN, NH₄⁺, TP , FRP |
| MR05010 | 2 | Peel Harvey | Al, Fe, Zn | AI, Fe, Zn | TN, TP , FRP |
| NYUND | 2 | Peel Harvey | - | AI | TN, TP |
| PATRDTRIB | 4b | Peel Harvey | Al , Co , Fe , Mn | Al, Co , Cu, Fe , Mn, V | TN, NH₄⁺, TP , FRP |
| PHM3 | 2 | Peel Harvey | - | Al, Fe | - |
| PHS7 | 2 | Peel Harvey | AI, Fe, V | Al, Cr, Fe, V | TN, NH₄ ⁺ , TP , <mark>FRP</mark> |
| SR04014 | 1 | Peel Harvey | AI, Fe, V | AI, Fe, V | TN, NH₄ ⁺ , TP , FRP |
| ACID01 | 4a | Leschenault | AI | AI | - |
| ACID02 | 5 | Leschenault | AI | AI, Co, Fe | TN , NH₄⁺ , TP, FRP |
| ACID03 | 2 | Leschenault | Al, Co , Fe | Al, Co, Fe | TN , NH₄⁺ , TP, FRP |
| ACID04 | 2 | Leschenault | AI, Co, Cu, Fe | AI, Co , Cu, Fe , V | NOx, TN , NH₄⁺ , TP, FRP |
| ACID05 | 5 | Leschenault | AI | AI | NOx |
| ACID06 | 4a | Leschenault | ΑΙ | AI, Fe | NOx |
| ACID07 | 1 | Leschenault | AI, Co | AI, Co | - |
| ACID08 | 2 | Leschenault | - | AI | - |
| ACID09 | | Leschenault | AI | AI, Fe | TN, NH4 ⁺ , TP, FRP |
| ACID10 | 4a | Leschenault | AI | AI, Fe | NOx, TN , NH₄⁺ , TP, FRP |
| ACID11 | 4b | Leschenault | AI | AI, Co, Fe | NOx, TN, NH₄⁺ , TP |
| ACID12 | 2 | Leschenault | AI | AI | - |
| ACID13 | 4a | Leschenault | AI, Cu | AI, Cr, Cu, Fe | NOx |

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|---------------|------------------------|---------------------------------|--|---|
| ACID14 | 4a | Leschenault | AI | AI | NOx |
| ACID15 | 1 | Leschenault | AI | AI | NOx |
| ACID16 | 5 | Leschenault | AI , Cu, Fe | <mark>AI</mark> , Co, Cu, Fe , V | TN , NH₄ ⁺ , TP, FRP |
| ACID17 | 2 | Leschenault | AI, Co, Fe | AI, Co, Fe | TN, NH 4 ⁺ |
| BRUN6 | 4b | Leschenault | AI | AI, Fe | NOx, TN, TP |
| GBC01 | 4a | Leschenault | AI , Cu, Fe | AI , Co, Cu, Fe | NOx, TN , NH₄⁺ , TP, FRP |
| GBC02 | 5 | Leschenault | AI | AI, Co, Fe | TP |
| GBC03 | 4a | Leschenault | AI | AI, Fe | NOx |
| LESCH01 | 4b | Leschenault | AI | AI, Cu | - |
| LESCH02 | 4b | Leschenault | AI, Co | AI, Co Fe | NOx, TN, TP |
| LESCH03 | 4a | Leschenault | AI | AI, Cu, Fe | NOx |
| LESCH04 | 4b | Leschenault | AI | AI, Fe | TP |
| LESCH05 | 3 | Leschenault | AI, | AI , Co, Fe | NOx, TN , NH₄ ⁺ , TP, FRP |
| LESCH06 | 2 | Leschenault | AI, Fe | AI , Fe | TN, NH₄ ⁺ , TP, FRP |
| LESCH07 | 1 | Leschenault | - | AI | NOx, TN, NH4 ⁺ |
| 608003 | 2 | Scott Coastal Plain | - | Al, Fe | - |
| 608147 | 2 | Scott Coastal Plain | AI | AI | - |
| 608171 | 2 | Scott Coastal Plain | AI, Cu | Al, Cu | - |
| 609022 | 4b | Scott Coastal Plain | - | AI | - |

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|---------------|------------------------|-----------------------------------|-----------------------------------|--------------------------------------|
| 609041 | 2 | Scott Coastal Plain | - | - | - |
| 609060 | 5 | Scott Coastal Plain | - | - | - |
| 6081009 | 2 | Scott Coastal Plain | AI | AI | - |
| 6081010 | 2 | Scott Coastal Plain | - | Al, Fe | - |
| 6081026 | 2 | Scott Coastal Plain | <mark>Al</mark> , Fe | <mark>Al</mark> , Fe | - |
| 6081040 | 2 | Scott Coastal Plain | AI | AI | - |
| 6081053 | 2 | Scott Coastal Plain | AI | Al, Cr | - |
| 6091008 | 2 | Scott Coastal Plain | - | AI | - |
| 6091044 | 2 | Scott Coastal Plain | Fe | Fe | - |
| 6091222 | 4b | Scott Coastal Plain | Al, Co, Cr, Fe | Al, Co, Cr, Fe | TN, TP |
| 6091223 | 2 | Scott Coastal Plain | AI , Co, Fe , Mn | AI , Co, Fe , Mn | TN, TP |
| 6091224 | 5 | Scott Coastal Plain | Co | Al, Co | NOx , TN, NH₄ ⁺ |
| 6091302 | 2 | Scott Coastal Plain | AI | AI, Fe | - |
| 6091305 | 4a | Scott Coastal Plain | AI | AI, Fe | NOx |
| 6091408 | 2 | Scott Coastal Plain | AI | AI | - |
| 6091596 | 2 | Scott Coastal Plain | Al, Co, Fe | Al, Co, Cr, Fe | - |
| 6091735 | 1 | Scott Coastal Plain | AI | AI | - |

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-------------|---------------|------------------------|---------------------------------|----------------------------|---|
| 6091752 | 1 | Scott Coastal Plain | Al, Fe | AI, Fe | TN, TP |
| 6091753 | 2 | Scott Coastal Plain | - | - | NOx |
| 6091754 | 4a | Scott Coastal Plain | - | AI | NOx |
| 6091755 | 3 | Scott Coastal Plain | AI, Co, Fe | AI , Co, Fe | ТР |
| 6091756 | 2 | Scott Coastal Plain | AI | AI | TN |
| 6091757 | n.d. | Scott Coastal Plain | Co | Al, Co, Fe | TN , NH4 ⁺ , TP |
| 6091758 | 5 | Scott Coastal Plain | Al, Co, Fe | <mark>Al</mark> , Co, Fe | - |
| CHE | 4b | Albany | - | AI | NOx |
| DML | 2 | Albany | AI | AI | - |
| FREEBOROUGH | 4b | Albany | AI, Fe | <mark>Al</mark> , Fe | TN, NH4 ⁺ , TP, FRP |
| HAY | 2 | Albany | AI | AI | - |
| HENNING | 2 | Albany | AI, Fe | AI, Fe | - |
| INFLOW | 2 | Albany | <mark>Al</mark> , Fe | Al, Fe | TN, NH₄ ⁺ , TP , FRP |
| KR | 2 | Albany | AI, Fe | AI, Fe | TP |
| LEAY | 4b | Albany | AI, Fe | Al, Fe | NH_4^+ |
| LIT | 4a | Albany | AI, Fe | AI, Fe | TP |
| LOWKG | 3 | Albany | AI, Fe | AI, Fe | TN, TP , FRP |
| MANCK | 2 | Albany | AI, Fe | Al, Fe | TN, TP, FRP |
| MIT | 4b | Albany | AI, Co | Al, Co | - |
| MRD | 3 | Albany | AI, Fe | AI, Fe | TN, TP , FRP |
| MTBARKRD | 3 | Albany | AI, Cr, Fe | Al, Cr, Fe | - |

| Site name | lso- group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|---------------|--------|---------------------------------|---|--|
| RD | 5 | Albany | AI | Al, Fe | NOx , TN, NH₄ ⁺ , TP , FRP |
| RZ7 | 2 | Albany | AI, Fe | AI, Fe | TN, TP, FRP |
| SCO | 4a | Albany | AI, Fe | AI, Fe | - |
| SLE | 4b | Albany | AI, Fe | AI, Fe | TP |
| SUN | 4b | Albany | AI, Fe | AI, Fe | TN, TP , FRP |
| TILP001 | 3 | Albany | AI, Co , Fe | AI, Co, Cu, Fe | TN, TP , FRP |
| TILP006 | 2 | Albany | AI | AI, Fe | TN, NH₄ ⁺ , TP , FRP |
| TILP008 | 2 | Albany | AI, Fe | AI, Fe | NOx, TP |
| TILP012 | 5 | Albany | AI, Co, Fe, | AI, Co, Fe | TN, NH_4^+ |
| TILP013 | 2 | Albany | AI, Cr, Fe | AI, Cr, Fe | TN , TP |
| TILP047 | 2 | Albany | Al, Fe, Zn | <mark>Al, Cr, Fe</mark> , V, <mark>Zn</mark> | TN, TP |
| WCWF | 2 | Albany | AI, Fe | AI, Fe | NOx, TP |
| WOOD1 | 5 | Albany | Al, Co, Cr, Fe | AI, Co, Cr, Fe | - |
| YAK1 | 5 | Albany | AI, | Al, Cr, Fe | NOx , TN, NH ₄ ⁺ , TP |
| YU | 5 | Albany | AI | AI | NOx, TN, TP |

F2. Estuarine sites at which metals and nutrients exceeded ANZECC & ARMCANZ trigger values. The ANZECC & ARMCANZ (2000) trigger values for 95% species protection – freshwater or marine (see Section 7.3) – and estuarine ecosystem stress were used. Trigger values were adjusted by the hardness algorithm for cadmium, chromium, copper, nickel, lead and zinc. Normal text indicates value exceeded trigger value, **bold** text indicates value was 2x trigger value, and **red** text indicates 10x trigger value.

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-----------------|---------------------------------|----------------------------|---|
| 616094-s | 4b | Swan Canning | - | n.d | TP, FRP |
| 616094-w | 2 | Swan Canning | - | AI, | NO _x , TN, NH₄⁺, TP, FRP |
| ARM-s | 5 | Swan Canning | - | n.d | TP, FRP |
| ARM-w | 4a | Swan Canning | - | - | FRP |
| BAC-s | 4b | Swan Canning | - | n.d | TP, FRP |
| BAC-w | 1 | Swan Canning | - | AI, | NO _x , TN, NH₄⁺ , TP |
| BLA-s | 4a | Swan Canning | - | n.d | TP, FRP |
| BLA-w | n.d. | Swan Canning | - | - | NH ₃ |
| CAS-s | 5 | Swan Canning | - | n.d | TN, TP , FRP |
| CAS-w | 1 | Swan Canning | - | AI | NO _x , TN, NH ₃ , TP, FRP |
| ELL-s | 5 | Swan Canning | - | n.d | TN, TP, FRP |
| ELL-w | 5 | Swan Canning | - | AI, Fe, | NO _x , TN, NH₄⁺, TP, FRP |
| KIN-s | 5 | Swan Canning | - | n.d | TN, TP, FRP |
| KIN-w | n.d. | Swan Canning | - | AI | NO _x , TN, NH₄⁺ , TP, FRP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|------------------|-----------|-----------------|---------------------------------|----------------------------|---|
| MAY-s | 5 | Swan Canning | - | n.d | TN, TP , FRP |
| MAY-w | 4a | Swan Canning | - | | NH4 ⁺ , TP, FRP |
| NAR-s | 4a | Swan Canning | - | n.d | TP, FRP |
| NAR-w | 1 | Swan Canning | - | AI, | FRP |
| NIC-s | 4b | Swan Canning | - | n.d | TP, FRP |
| NIC-w | 5 | Swan Canning | Cu | AI, Cu | NO _x , TN, NH₄ ⁺ , TP, FRP |
| NIL-s | 5 | Swan Canning | - | n.d | TN, NH₄ ⁺ , TP , FRP |
| NIL-w | 1 | Swan Canning | - | - | NH4 ⁺ , TP, FRP |
| RIV-s | 5 | Swan Canning | - | n.d | TN, TP , FRP |
| RIV-w | 1 | Swan Canning | - | - | NH4 ⁺ , TP, FRP |
| RON-s | 4a | Swan Canning | - | n.d | TN, TP , FRP |
| RON-w | 1 | Swan Canning | - | - | NO _x , TN, NH₄⁺ , TP, FRP |
| SAL-s | 4a | Swan Canning | - | n.d | TP, FRP |
| SAL-w | 1 | Swan Canning | - | - | NH4 ⁺ , TP, FRP |
| STJ-s | 4a | Swan Canning | - | n.d | TN, NH4 ⁺ , TP , FRP |
| STJ-w | 4b | Swan Canning | - | - | NH4 ⁺ , TP, FRP |
| SUC ^S | 2 | Swan Canning | - | n.d | TN, TP, FRP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-----------------|---------------------------------|----------------------------|---|
| SUC-w | 1 | Swan Canning | - | AI | NO _x , TN, NH₄⁺ , TP, FRP |
| MPH001-w | 1 | Peel Harvey | - | AI, Co | TN, TP, FRP |
| MPH002-w | 2 | Peel Harvey | - | AI | TN, TP, FRP |
| MPH003-w | 1 | Peel Harvey | - | AI | NO _x , TN, NH4 ⁺ , TP, FRP |
| MPH006-s | 4a | Peel Harvey | - | n.d. | - |
| MPH006-w | n.d. | Peel Harvey | - | Al, As | - |
| MPH015-s | 1 | Peel Harvey | - | n.d. | TP |
| MPH015-w | 3 | Peel Harvey | AI | AI, As | TN, TP |
| MPH019-s | 1 | Peel Harvey | - | n.d. | TN, TP |
| MPH019-w | 1 | Peel Harvey | AI | AI, As | TN, TP |
| MPH025-s | 5 | Peel Harvey | - | n.d. | - |
| MPH025-w | 1 | Peel Harvey | - | AI, As | - |
| MPH040-s | 4b | Peel Harvey | - | n.d. | TN, TP |
| MPH040-w | 4b | Peel Harvey | - | AI | TN, NH₄ ⁺ , TP, FRP |
| MPH044-s | 2 | Peel Harvey | - | n.d. | TN, TP, FRP |
| MPH044-w | 2 | Peel Harvey | - | Al, As | TP |
| MPH061-s | 1 | Peel Harvey | - | n.d. | FRP |
| MPH061-w | 2 | Peel Harvey | - | As | - |
| MPH071-s | 1 | Peel Harvey | - | n.d. | FRP |
| MPH071-w | 4a | Peel Harvey | - | AI, As | - |
| MPH083-s | 2 | Peel Harvey | - | n.d. | TP, FRP |
| MPH083-w | 2 | Peel Harvey | AI | AI, As | - |
| MPH091-s | 4a | Peel Harvey | - | n.d. | TP, FRP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-------------|---------------------------------|----------------------------|--|
| MPH091-w | 1 | Peel Harvey | AI, Fe | AI, Fe | NO _x , TN, NH4 ⁺ , TP, FRP |
| MPH155-s | 4b | Peel Harvey | - | n.d. | TN, TP |
| MPH155-w | 2 | Peel Harvey | AI | AI | NO _x , TN, NH₄ ⁺ , TP, FRP |
| MPH158-s | 1 | Peel Harvey | - | n.d. | TN, TP, FRP |
| MPH158-w | 2 | Peel Harvey | AI | AI | NO _x , TN, NH₄⁺, TP, FRP |
| PHE01-s | 1 | Peel Harvey | - | n.d. | TP, FRP |
| PHE01-w | 5 | Peel Harvey | - | AI | NO _x , NH₄⁺ , SRP |
| PHE02-s | 4b | Peel Harvey | - | n.d. | FRP |
| PHE02-w | 5 | Peel Harvey | - | AI | - |
| PHE04-s | 1 | Peel Harvey | - | n.d. | FRP |
| PHE04-w | 4a | Peel Harvey | - | AI | NO _x , TN, NH₄ ⁺ , TP, FRP |
| PHE07-s | 1 | Peel Harvey | - | n.d. | FRP |
| PHE07-w | 4a | Peel Harvey | - | AI | NH ₃ , FRP |
| PHE31-s | 1 | Peel Harvey | - | n.d. | TN, TP, FRP |
| PHE31-w | 4a | Peel Harvey | - | AI | NH₄ ⁺ , FRP |
| PHE58-s | 2 | Peel Harvey | - | n.d. | FRP |
| PHE58-w | n.d. | Peel Harvey | - | AI | FRP |
| PHRM2-s | 4a | Peel Harvey | AI | n.d. | TN, TP, FRP |
| PHRM2-w | 2 | Peel Harvey | - | AI | NO _x , TN, NH4 ⁺ , TP, FRP |
| PHRM4-s | 4a | Peel Harvey | - | n.d. | FRP (TN/TP n.d.) |
| PHRM4-w | 2 | Peel Harvey | - | AI | NO _x , TN, NH₄⁺ , TP, FRP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-------------|---------------------------------|------------------------------|---|
| PHRM9-s | 2 | Peel Harvey | - | n.d. | FRP (TN/TP n.d.) |
| PHRM9-w | 2 | Peel Harvey | - | AI | NO _x , TN, NH₄⁺ , TP, FRP |
| PHRS4-s | 3 | Peel Harvey | - | n.d. | TN, NH₄⁺, TP , FRP |
| PHRS4-w | 1 | Peel Harvey | - | AI | FRP |
| PHRS6-s | 4a | Peel Harvey | - | n.d. | TN, NH₄ ⁺ , TP, <mark>FRP</mark> |
| PHRS6-w | 1 | Peel Harvey | - | - | TN, TP, FRP |
| PHRS7-s | 1 | Peel Harvey | Со | n.d. | TN, NH₄ ⁺ , TP, FRP |
| PHRS7-w | 4a | Peel Harvey | - | AI | TN , TP , FRP |
| 6111043-s | 4a | Leschenault | - | n.d. | NH4 ⁺ , FRP |
| 6111043-w | 5 | Leschenault | Al, Fe | AI, Fe | <mark>NO</mark> x, TN, NH₄ ⁺ , TP, FRP |
| 6121130-s | 2 | Leschenault | - | n.d. | FRP |
| 6121130-w | 4a | Leschenault | Al, Fe, Cd | Al, Cd | NO _x , TN, FRP |
| 6121161-s | 1 | Leschenault | - | AI (only Al, Fe analysed) | NO _x , TN, NH₄⁺ , TP , FRP |
| 6121161-w | 3 | Leschenault | AI, Fe | AI, Co, Fe, Cu | <mark>NO</mark> x, TN, NH₄ ⁺ , TP, FRP |
| 6121162-s | 2 | Leschenault | - | n.d. | NO _x , TN, <mark>TP</mark> , FRP |
| 6121162-w | 3 | Leschenault | Al, Co, Fe | AI, Co, Fe | <mark>NO</mark> x, TN, NH₄ ⁺ , TP, FRP |
| 6121166-s | 4b | Leschenault | - | AI (only Al, Fe analysed) | NH₄⁺, TP, FRP |
| 6121166-w | 4a | Leschenault | Al, Fe | AI, Fe | NO _x , TN, NH4 ⁺ , TP, FRP |
| 6121168-s | 2 | Leschenault | - | n.d. | NH_4^+ , FRP |
| 6121168-w | 4a | Leschenault | - | AI , Fe | <mark>NO</mark> x, TN, NH₄⁺, TP, FRP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-------------------|---------------------------------|----------------------------|--|
| 6121173-s | 1 | Leschenault | Co, V | n.d. | TN, NH₄ ⁺ , TP, FRP |
| 6121173-w | 3 | Leschenault | - | AI | TN, NH₄ ⁺ , TP, FRP |
| 6121203-s | 4b | Leschenault | AI | n.d. | NO _x , TN, NH₄⁺, TP, FRP |
| 6121203-w | 5 | Leschenault | AI | AI , Fe, Cu, V | <mark>NO</mark> x, TN, NH₄ ⁺ , TP, FRP |
| 6121206-s | 4a | Leschenault | - | n.d. | NH4 ⁺ , FRP |
| 6121206-w | 4a | Leschenault | - | - | FRP |
| 6121207-s | 1 | Leschenault | - | n.d. | FRP |
| 6121207-w | 5 | Leschenault | - | - | NH4 ⁺ , FRP |
| 610019-s | 1 | Vasse Wonnerup | - | n.d. | TN, NH4⁺, TP |
| 610019-w | 4a | Vasse Wonnerup | - | - | NO _x , TN, NH4 ⁺ , TP |
| 610020-s | 1 | Vasse Wonnerup | - | n.d. | NH4⁺ , TP |
| 610020-w | 5 | Vasse Wonnerup | Fe | Fe , Zn | NO _x , TN, NH ₄ ⁺ , TP |
| 6101063-s | 4b | Vasse Wonnerup | - | n.d. | TN, TP |
| 6101063-w | 4a | Vasse Wonnerup | - | - | TN, NH ₄ ⁺ , TP |
| 6101064-s | 4b | Vasse Wonnerup | - | n.d. | TN |
| 6101064-w | 5 | Vasse Wonnerup | - | Fe | NO _x , TN, NH4 ⁺ , TP |
| 6101065-s | 2 | Vasse Wonnerup | - | n.d. | NH_4^+ |
| 6101065-w | 4a | Vasse Wonnerup | Fe | Fe | NO _x , TN, NH ₄ ⁺ , TP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-----------------------|---------------------------------|--------------------------------|--|
| 6101067-s | 1 | Vasse Wonnerup | - | n.d. | TN, TP |
| 6101067-w | 4a | Vasse Wonnerup | - | - | - |
| 6101072-s | 4b | Vasse Wonnerup | - | n.d. | TN, TP |
| 6101072-w | 4a | Vasse Wonnerup | - | - | NH_4^+ |
| BRF01-w | 2 | Hardy | Al, Fe | AI | NO_x , TN, NH4 ⁺ , TP , FRP |
| BRF02-w | 2 | Hardy | - | AI | NO _x , TN |
| HIF01-w | 2 | Hardy | - | - | NO _x , TN |
| HIF02-w | 2 | Hardy | - | AI | NO _x , TN |
| HIF03-w | 2 | Hardy | - | AI | NO _x , TN |
| HIF04-w | 2 | Hardy | - | AI | NO _x , TN |
| HIF05-w | 2 | Hardy | - | - | NO _x , TN |
| HIF06-w | 2 | Hardy | - | - | NO _x , TN |
| MIF01-w | 2 | Hardy | - | AI | NO _x , TN |
| MIF02-w | 2 | Hardy | - | AI | <mark>NO</mark> _x , TN, TP, FRP |
| MIF03-w | 4a | Hardy | AI, Cu, Fe | AI | NO _x , TN, NH4 ⁺ , TP, FRP |
| SRF01-w | 4a | Hardy | AI, Fe | AI | NO _x , TN, TP, FRP |
| AOH 2-s | 5 | South coast estuaries | - | n.d. | FRP |
| AOH 2-w | 4b | South coast estuaries | - | - (only Al, Fe analysed) | FRP |
| AOH 4-s | 5 | South coast estuaries | - | n.d. | FRP |
| AOH 4-w | n.d. | South coast estuaries | - | - (only Al, Fe analysed) | FRP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-----------------------|---------------------------------|--------------------------------|--------------------|
| AOH 5-s | 1 | South coast estuaries | - | n.d. | FRP |
| AOH 5-w | 1 | South coast estuaries | - | - (only Al, Fe analysed) | FRP |
| APRH 2-s | 1 | South coast estuaries | - | n.d. | FRP |
| APRH 2-w | 1 | South coast estuaries | - | - (only Al, Fe analysed) | FRP |
| APRH 3-s | 4a | South coast estuaries | - | n.d. | FRP |
| APRH 3-w | n.d. | South coast estuaries | | (only Al, Fe analysed) | FRP |
| APRH 4-s | 4a | South coast estuaries | | n.d. | FRP |
| APRH 4-w | 1 | South coast estuaries | - | - (only Al, Fe analysed) | FRP |
| BRO001-s | 1 | South coast estuaries | AI | n.d. | - |
| BRO001-w | n.d. | South coast estuaries | - | AI | FRP |
| BRO002-s | 1 | South coast estuaries | AI | n.d. | - |
| BRO002-w | n.d. | South coast estuaries | AI | AI | FRP |
| BRO004-s | 1 | South coast estuaries | Al | n.d. | - |
| BRO004-w | n.d. | South coast estuaries | - | AI | FRP |
| IRW001-s | 1 | South coast estuaries | - | n.d. | FRP |
| IRW001-w | n.d. | South coast estuaries | - | - | FRP |
| IRW003-s | 1 | South coast estuaries | - | n.d. | FRP |

| Site name | lso-group | Region | Exceeded metals (soluble) | Exceeded metals (total) | Exceeded nutrients |
|-----------|-----------|-----------------------|---------------------------------|----------------------------|-------------------------------|
| IRW003-w | n.d. | South coast estuaries | AI | AI | FRP |
| NI 10-s | 1 | South coast estuaries | - | n.d. | - |
| NI 10-w | n.d. | South coast estuaries | AI | AI | FRP |
| NI 5-s | 2 | South coast estuaries | - | n.d. | - |
| NI 5-w | n.d. | South coast estuaries | AI | AI | FRP |
| WAL001-s | 4b | South coast estuaries | - | n.d. | - |
| WAL001-w | n.d. | South coast estuaries | AI | AI | TN, TP, FRP |
| WAL003-s | 1 | South coast estuaries | - | n.d. | NH_4^+ |
| WAL003-w | n.d. | South coast estuaries | AI, Fe | AI | TP, FRP |
| WAL004-s | 1 | South coast estuaries | - | n.d. | - |
| WAL004-w | n.d. | South coast estuaries | AI | AI | FRP |
| WI12-s | 2 | South coast estuaries | - | n.d. | NH_4^+ , FRP |
| WI12-w | 2 | South coast estuaries | (only Al, Fe analysed) | - | TN, NH₄ ⁺ , FRP |
| WI30-s | 1 | South coast estuaries | - | n.d. | - |
| WI30-w | 1 | South coast estuaries | (only Al, Fe analysed) | - | NH_4^+ , FRP |
| WI6-s | 2 | South coast estuaries | - | n.d. | FRP |
| WI6-w | 1 | South coast estuaries | (only Al, Fe analysed) | - | FRP |

Appendix G - Sulfur stable isotope ratios measured in fertilisers, detergent and rainfall

G1. Sulfur stable isotope ratios (per mil, ‰) measured for various sources of 'oxidised sulfur'.

| Source | Manufacturer | Product name | del34S |
|------------|-----------------------|---------------------------------|-----------|
| Fertiliser | Yates | Thrive water soluble plant food | 2.5 |
| | Scotts | Osmocote multi-purpose | 2.9 |
| | | Osmocote plus roses | 2.0 |
| | | Lawn builder | 8.3 |
| | Cresco | Lawn fertiliser | 0.7 |
| | Richgro | Sulfate of potash | -0.1 |
| | CSBP | Sulfate of ammonia | 2.2 |
| | | Super phosphate | 1.1 |
| | | Coastal super | 2.3 |
| | | GSOP | 18.7 |
| Detergent | United Laboratories | Duo laundry detergent | 21 |
| Rainfall | (collected in Perth m | netropolitan area) | 18.2-20.0 |

Acronyms

| ANOVA | Analysis of variance |
|-----------|--|
| ANZECC | Australian and New Zealand Environment and Conservation Council |
| ARMCANZ | Agriculture and Resource Management Council of Australia and New Zealand |
| ASS | Acid sulfate soils |
| DEC | Department of Conservation and Environment |
| DoE | Department of Environment |
| DoW | Department of Water |
| МВО | Monosulfidic black ooze |
| PERMANOVA | Permutational multivariate analysis of variance |

Glossary

| Abiotic | Non-living chemical and physical factors in the environment. |
|-----------------------------|--|
| Acids, fully dissociated | Acids that fully dissociate in water are called strong acids; for example, $H_2SO_{4(aq)} \rightarrow 2H^+_{(aq)} + SO_4^{2^-}_{(aq)}$. |
| Acids, weak | Acids that do not fully dissociate when dissolved in water are called weak acids. Vinegar is a commonly known weak acid. These acids, when dissolved in water, form an equilibrium where both reactants and products are present. The equilibrium means that if some H^+ ions are removed (i.e. by neutralisation), more will be formed to maintain the equilibrium. |
| Acidification | The process by which soil or water becomes more acid (decreasing pH). |
| Alkalinity | A measure of a solution's ability to resist changes in pH due to the addition of an acid. In natural waters this usually relates to the amount of bicarbonate, carbonate, and hydroxide compounds present in the water. |
| Algal blooms | The rapid excessive growth of algae, generally caused by high nutrient levels and favourable conditions. Water-column deoxygenation may occur when the algae die. |
| Analyte | A substance or chemical constituent that is determined in an analytical procedure. |
| Anaerobic | Without air, where 'air' is generally used to mean oxygen. |
| Anoxic | An absence of oxygen, an extreme form of hypoxia or 'low oxygen' (anoxic 0% and hypoxia <40% dissolved oxygen saturation in solution). |
| Anthropogenic | Originating from the activity of humans. |
| Baseline data | Data or measurements collected as a starting point, generally before a program begins. |
| Bimodal distribution | A statistical distribution having two modes – indicating a mixing of two populations such as the salinity from a dataset containing a mix of fresh and marine waters. |
| Blackwater event | Rapid deoxygenation of a waterway due to the high oxygen demand of the mobilised organic-rich matter (may also contain monosulfides). Blackwater events are usually triggered by a rainfall event. |
| Blank sample | Clean samples of distilled de-ionised water prepared in the field or laboratory, treated as normal samples to determine if contamination occurs during sampling, handling or analysis. |
| Box plots | A graph summarising the distribution of a set of data values using |

| | summary data: minimum, lower quartile (25%), median, upper quartile (75%), maximum and often outliers observations. Also known as a box- and-whisker diagram or plot. |
|------------------------------|--|
| Buffering | Resisting changes in pH when a small amount of strong acid or base is added to a solution. Common buffering agents are carbonate or bicarbonates. Seawater contains high quantities of these compounds. |
| Chloride to sulfate ratio | The ratio of the mass of chloride to sulfate, used as an indicator of changes in sulfate. Chloride is considered a conservative ion that has few additional sources except seawater. The ratio of seawater is normally ~7.2:1. Additional sulfate from acidic drainage can reduce this ratio to <4:1. Sulfate removed by sulfate reduction can produce ratios of >25:1. |
| Correlation | Indicates the strength and direction of the linear relationship between two random variables. |
| Dewatering | The removal of groundwater during sewerage and other below-ground construction projects. This is often carried out during the site development phase of a major construction project due to a high water table. |
| Dredging | Dredging is an excavation activity carried out at least partly underwater, in shallow seas, estuaries or freshwater areas to keep waterways navigable. Bottom sediments are collected and disposed of at a different location. |
| Eutrophic | Waters rich in mineral and organic nutrients that promote plant life, especially algae, which can cause a reduction in dissolved oxygen as the algal population collapse and biological oxygen demand increases. |
| Fish kill | The sudden and mass death of aquatic animals due to the introduction of pollutants or the reduction of dissolved oxygen concentration in a waterbody. |
| Fractionation | A separation process in which a mixture (solid, liquid, solute, suspension or isotope) is divided up in a number of smaller quantities (fractions) and the composition changes according to a gradient. For example, fractionation of sulfur by sulfate-reducing bacteria, where bacteria use the lighter sulfur isotope (³² S) preferentially to the heavier sulfur isotope (³⁴ S), which results in a change in isotopic composition of the resulting sulfide relative to the starting material. |
| Guidelines | Values or ranges of acceptable or unacceptable levels of a chemical, beyond which a management response is usually triggered. |
| Holocene period | A geological epoch at the end of the last Ice Age which began approximately 10 000 years ago and is characterised by the development of human civilisations. |

| Hotspots | In the context of this project: areas which show clear signals of acidity by either iso-group classification or extremely high concentrations of iron and/or aluminium |
|----------------------------------|--|
| Hydrolysis | Chemical reaction involving water and the oxidation of a compound. |
| lso-group | In the context of this project: groupings that categorise the signal of acidic drainage using a combination of both sulfur stable isotope and sulfate and chloride analysis. |
| Isotope | One of several nuclides having the same number of protons in their nuclei, but with different numbers of neutrons, resulting in a very slight variation in the mass of the element. |
| Isotopically light | The substance which is enriched in the lighter isotope relative to the composition of another substance; for example, low δ^{34} S value (relative to another). |
| Leaching | The loss of mineral and organic solutes from the soil matrix due to percolation. It is a mechanism of soil formation and acidic drainage. |
| LOR | Limit of reporting. The lower limit of reliability given by the laboratory responsible for carrying out the analysis. Greater than the limit of detection (LOD). |
| Median | The middle value of an ordered set of values. It indicates the centre line of a distribution, while the mean (average) can be skewed by a single high or low value. |
| Metal mobilisation | Metals such as aluminium, iron, cadmium and zinc that are bound to organic or mineral components of the soil matrix which when dissolved can be redistributed to other areas of the landscape. |
| Multi- dimensional scaling | A multivariate statistical technique which aims to develop spatial structure from numerical data by estimating the differences and similarities between analytical units. The resulting locations are commonly displayed in a 2D or 3D visualisation. |
| Multi-parameter probe | Often refers to a handheld water quality meter that measures parameters in the field (in situ). Parameters can include: pH, temperature, dissolved oxygen, electrolytic conductivity, redox, turbidity and salinity. Water quality data is collected by submersing the sensor in the water to indicate environmental conditions. |
| Non-parametric | Statistical procedures which are not based on the dimensional units of the measured observation, rather these procedures usually use the rank order of observations. |
| Oxidation | The loss of electrons accompanied by an increase in oxidation state. This process does not necessarily require the presence of oxygen. |
| рН | A log scale for indicating the acidity of a solution in terms of hydrogen |

ion concentration.

- **Precipitate** The solid formed when two solutions are mixed together which have an insoluble product.
- **Redox** In aqueous solutions, the reduction potential is the tendency of the solution to either gain or lose electrons and is measured in volts (V), millivolts (mV), or Eh (1 Eh = 1 mV). Because the absolute potentials are difficult to accurately measure, reduction potentials are defined relative to the standard hydrogen electrode which is arbitrarily given a potential of 0.00 V.
- **Reduction** The gain of electrons accompanied by a decrease in oxidation state.
- ReplicateA second sample taken from the same site at the same time, usingsampleexactly the same methods. They are representative of the same
environmental condition and used to evaluate the variability of the
environment and to establish how reproducible the sample is.
- SalinityMeasures the concentration of dissociated salts in a solution
(commonly Na⁺, Ca⁺², Mg⁺², Cl⁻, CO₃⁻²), recorded in practical salinity
units (psu) or parts per thousand (ppt). Salinity ranges from fresh 0-<5,
brackish 5-25, and saline >25 ppt. Marine waters are generally 32-38
ppt.
- Sulfides Group of minerals that are compounds of one or more metallic elements combined with the non-metallic element sulfur; for example, pyrite FeS₂, or galena PbS.
- **Surface waters** Waterways that include estuaries, rivers, drains, lakes and wetlands. Does not include groundwater or pore water.
- Sulfate-
reductionThe microbially catalysed process which converts sulfate $(SO_4^{2^-}_{(aq)})$ to
sulfide $(S^{2^-}_{(aq)})$.
- **Sulfur isotope** Forms of sulfur with the same number of protons but differing numbers of neutrons; for example, S³⁴ with one additional neutron per molecule is the second-most abundant (4.2%) of sulfur isotopes (c.f. S³² abundant 95% the common form).
- **Trophic web** A set of interconnected relationships by which energy and materials circulate within an ecosystem. The food web is divided into two broad categories: the grazing web materials typically pass from plants to plant eaters (herbivores) to flesh eaters (carnivores) (plants) and the detrital web (organic debris); the detrital web materials pass from plant and animal matter to bacteria and fungi (decomposers), then to detrital feeders (detritivores), and then to their predators (carnivores).

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